Toxicology Management Review

Medical Toxicology 3: 33-58 (1988) 0112-5966/88/0001-0033/\$13.00/0 © ADIS Press Limited All rights reserved.

Summary

Oral Activated Charcoal in the Treatment of Intoxications Role of Single and Repeated Doses

Pertti J. Neuvonen and Klaus T. Olkkola

Department of Clinical Pharmacology, University of Helsinki and University Central Hospital, Helsinki

34

Contents

Summary	2.4
1. Factors Affecting the Antidotal Efficacy of Activated Charcoal	35
1.1 Effects of Physicochemical Properties	
1.2 Effects of Pharmaceutical Formulation	
1.3 Effects of Chemical Nature and Formulation of Agents Ingested in Intoxications	35
1.4 Effects of Charcoal Dose	35
1.5 Effect of pH	36
1.6 Gastrointestinal Contents	37
1.7 Effects of Stability of the Charcoal-Poison Complex	38
1.8 Time-Effect Relationships	
1.9 Effects of Purgatives and Whole-Bowel Irrigation	
2. Mechanisms of Enhanced Drug and Toxin Elimination by Multiple Charcoal Doses	
3. Effect of Single and Repeated Doses on the Absorption and Elimination of	
Drugs in Humans	41
3.1 Antipyretic Analgesics	41
3.2 Other Analgesics	
3.3 Hypnosedatives and Anticonvulsants	44
3.4 Antidepressants	
3.5 Other Psychopharmaca	45
3.6 Cardiac Glycosides	46
3.7 Antiarrhythmic Drugs	
3.8 β-Adrenoceptor Antagonists	47
3.9 Oral Antihyperglycaemic Drugs	47
3.10 Bronchodilating Drugs	47
3.11 Antimicrobial Agents	48
3.12 Hormones	49
3.13 Other Drugs	
3.14 Metals, Alcohols and Other Substances	
4. Relative Efficacy of Activated Charcoal, Emetics, and Gastric Lavage in	
Inhibiting Drug Absorption	52
5. Toxicity and Side Effects of Activated Charcoal	
6. Conclusions	
6.1 Role of Single Doses in Treatment of Intoxications	
6.2 Role of Repeated Doses in Treatment of Intoxications	
	-

Summary

Activated charcoal has an ability to adsorb a wide variety of substances. This property can be applied to prevent the gastrointestinal absorption of various drugs and toxins and to increase their elimination, even after systemic absorption.

Single doses of oral activated charcoal effectively prevent the gastrointestinal absorption of most drugs and toxins present in the stomach at the time of charcoal administration. Known exceptions are alcohols, cyanide, and metals such as iron and lithium. In general, activated charcoal is more effective than gastric emptying. However, if the amount of drug or poison ingested is very large or if its affinity to charcoal is poor, the adsorption capacity of activated charcoal can be saturated. In such cases properly performed gastric emptying is likely to be more effective than charcoal alone.

Repeated dosing with oral activated charcoal enhances the elimination of many toxicologically significant agents, e.g. aspirin, carbamazepine, dapsone, dextropropoxyphene, cardiac glycosides, meprobamate, phenobarbitone, phenytoin and theophylline. It also accelerates the elimination of many industrial and environmental intoxicants.

In acute intoxications 50 to 100g activated charcoal should be administered to adult patients (to children, about 1 g/kg) as soon as possible. The exceptions are patients poisoned with caustic alkalis or acids which will immediately cause local tissue damages. To avoid delays in charcoal administration, activated charcoal should be a part of first-aid kits both at home and at work. The 'blind' administration of charcoal neither prevents later gastric emptying nor does it cause serious adverse effects provided that pulmonary aspiration in obtunded patients is prevented.

In severe acute poisonings oral activated charcoal should be administered repeatedly, e.g. 20 to 50g at intervals of 4 to 6 hours, until recovery or until plasma drug concentrations have fallen to non-toxic levels. In addition to increasing the elimination of many drugs and toxins even after their systemic absorption, repeated doses of charcoal also reduce the risk of desorbing from the charcoal-toxin complex as the complex passes through the gastrointestinal tract. Charcoal will not increase the elimination of all substances taken. However, as the drug history in acute intoxications is often unreliable, repeated doses of oral activated charcoal in severe intoxications seem to be justified unless the toxicological laboratory has identified the causative agent as not being prone to adsorption by charcoal.

The role of repeated doses of oral activated charcoal in chronic intoxications has not been clearly defined. Charcoal seems able to accelerate the elimination of many industrial and environmental toxicants like dioxins, polychlorinated biphenyls and possibly also some heavy metals, including their radioactive isotopes. Further studies will be needed to define the value of repeated doses of oral activated charcoal in chronic intoxications.

Activated charcoal is an insoluble powder produced by pyrolysis of organic material. It is able to adsorb a wide variety of drugs and toxic agents onto its surface.

The capacity of charcoal to bind chemicals has been recognised for centuries and the first systematic studies of charcoal as an antidote were performed in the early 1800s. During the following hundred years, several studies on activated charcoal were published, but in many countries and in many hospitals its use as an antidote has not been accepted until recently.

One reason for this neglect has been the lack of suitable formulations, a situation which has pre-

vented the use of activated charcoal in adequate amounts. Furthermore, until the 1980s there have been only a few experimental or clinical human studies on the antidotal effect of *high* doses of activated charcoal. The effect of high single and repeated doses of charcoal on the absorption and elimination of various drugs has been studied intensively during the last 10 years. Various aspects of the antidotal use of activated charcoal have been reviewed lately (Cooney 1980; Levy 1982; Neuvonen 1982; Olkkola 1985a; Park et al. 1986; Pond 1986; Spector et al. 1986). As a result, the initial management in particular of intoxicated patients has changed.

Well adsorbed	Moderately adsorbed	Poorly or clinically inadequately adsorbed
Aflatoxins	Aspirin and other salicylates	Cyanide
Amphetamine	DDT	Ethanol
Antidepressants	Disopyramide	Ethylene glycol
Antiepileptics	Kerosene, benzene and dichlorethane	Iron
Antihistamines	Malathion	Lithium
Atropine	Many 'high-dose' NSAIDs e.g. tolfenamic acid	Methanol
Barbiturates	Mexiletine	Strong acids and alkalis
Benzodiazepines	Paracetamol (acetaminophen)	
β-Blocking agents	PCB-compounds	
Chloroquine and primaquine	Phenol	
Cimetidine	Syrup of ipecacuanha	
Dapsone	Tolbutamide, chlorpropamide, carbutamide,	
Dextropropoxyphene and other opioids	tolazamide	
Digitalis glycosides		
Ergot alkaloids		
Frusemide		
Glibenclamide and glipizide		
Glutethimide		
Indomethacin		
Meprobamate		
Nefopam		
Phenothiazines		
Phenylbutazone		
Phenylpropanolamine		
Piroxicam		
Quinidine and quinine		
Strychnine		
Tetracyclines		
Theophylline		

Table I. Adsorption of drugs and other substances to activated charcoal in vitro

1. Factors Affecting the Antidotal Efficacy of Activated Charcoal

1.1 Effects of Physicochemical Properties

Adsorption of chemicals onto charcoal is dependent on several factors. The physicochemical properties of charcoal are of vital importance for its antidotal efficacy. The major determinants of these properties are the pore size and the surface area of charcoal. If all pores are large enough for the drug to enter then the adsorption capacity of charcoal is proportional to its surface area. Modern activated charcoals have a large surface area (1000 to 3500 m²/g). A usual antidotal dose for adults

has an area of more than 10 football fields! Theoretically, recently developed 'superactive' charcoals (area up to $3500 \text{ m}^2/\text{g}$) should have a better antidotal efficacy than the conventional ones. There is some evidence for this both *in vitro* and *in vivo* (Cooney 1977a; Cooney & Kane 1980; Chung et al. 1982; Neuvonen et al. 1983d; Park et al. 1984), but the potential advantages in clinical practice of these oil-based charcoals are not clear.

1.2 Effects of Pharmaceutical Formulation

Water suspensions of charcoal are superior to coated charcoal tablets in antidotal use. Uncoated

tablets, as well as plain powdered or specially granulated forms of activated charcoal, can be suspended in water. Adding of thickening and flavouring agents (bentonite and carboxymethylcellulose, saccharin, sorbitol, cherry extract, etc.) to charcoal formulations has resulted in either reduced (De Neve 1976; Mathur et al. 1976a,b; Mayersohn et al. 1977; Yancy et al. 1977), increased (Gwilt & Perrier 1976; Picchioni et al. 1982), or unaffected (Cooney & Roach 1979; Scholtz et al. 1978) adsorption of drugs to activated charcoal.

Some formulations contain significant amounts of ethanol to prevent bacterial growth, sodium bicarbonate to help mixing, or other agents in addition to activated charcoal. These may have some consequences when given in high doses to intoxicated patients.

1.3 Effects of Chemical Nature and Formulation of Agents Ingested in Intoxications

Activated charcoal has the ability to adsorb a wide variety of substances onto its surface, but some substances such as ethanol, iron, lithium and cyanides are not adsorbed in clinically significant amounts (table I). Unfortunately, *in vitro* studies cannot be applied directly in clinical practice. The antidotal efficacy of activated charcoal in humans has to be established by direct human studies.

There seem to be no systematic studies on the ability of activated charcoal to adsorb agents from different pharmaceutical formulations. In healthy volunteers the absorption of a slowly absorbed formulation of phenylpropanolamine 50mg was reduced by 50% only when 25g of charcoal (charcoal-drug ratio 500 : 1) was given immediately after it (Neuvonen & Olkkola 1986). The absorption of 50mg phenylpropanolamine was reduced by 80% when a small dose of 2g of charcoal (charcoal-drug ratio 40 : 1) was ingested immediately after a rapidly absorbing phenylpropanolamine formulation (Tsuchiya & Levy 1972). The apparent discrepancy between these 2 studies is most probably due to the differences in the pharmaceutical formulation

of phenylpropanolamine used. The inhibition of the absorption of some other prolonged release formulations (theophylline, valproate) too, has been somewhat lower by a single dose of charcoal than might have been expected (Neuvonen et al. 1983b,c). In some cases the addition of sorbitol or other laxatives to an oral regimen of activated charcoal may decrease serum drug concentrations after the ingestion of slow release formulations (Goldberg et al. 1987).

1.4 Effects of Charcoal Dose

Completeness of adsorption depends greatly on the ratio of charcoal to poison, because according to the mass law there is an equilibrium between free and adsorbed poison. The higher the charcoalpoison ratio, the more complete is the adsorption. At a ratio of 10:1, 90 to 100% of most substances are adsorbed in optimum in vitro conditions. In acute intoxications the amount and type of poison taken is largely unknown. The less charcoal given, the more easily is its adsorption capacity saturated (figs 1 and 2). Therefore, in the initial management of intoxicated patients large doses (50 to 100g) of charcoal should be used (Neuvonen & Olkkola 1984a; Olkkola 1985b). On the other hand, too large doses of charcoal may cause vomiting and aspiration of charcoal by obtunded patients.

Huge daily doses of 200 to 400g activated charcoal should be given only in well controlled conditions, and only in severe poisoning if necessary. It should be realised that some formulations of activated charcoal may contain considerable amounts of sodium bicarbonate (e.g. 'Medicoal', 1.5g sodium bicarbonate per 5g charcoal), sorbitol (up to 70%) or ethanol, which in high doses may have significant gastrointestinal and systemic side effects.

1.5 Effect of pH

Environmental pH affects the adsorption capacity of activated charcoal. Compounds are best adsorbed to charcoal in their undissociated form,

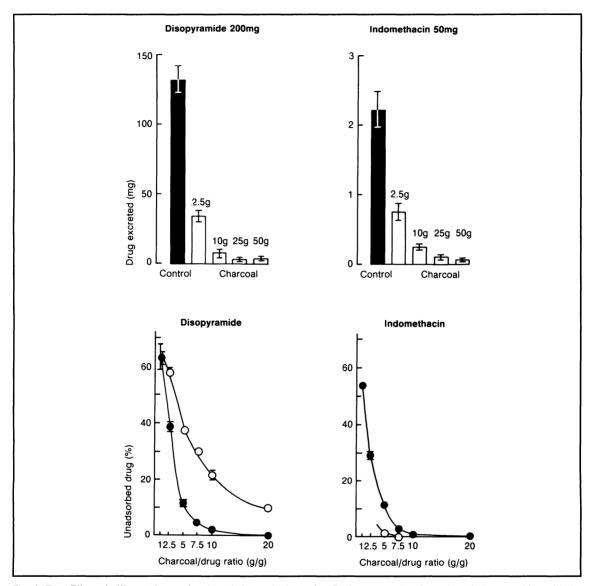


Fig. 1. Top: Effect of different doses of activated charcoal, given after 5 minutes, on the absorption of disopyramide and indomethacin, reflected as their cumulative excretion into urine over 72 hours. Mean \pm SEM in 6 volunteers. Bottom: The percentages of unabsorbed disopyramide and indomethacin at pH 1.2 (\odot) and pH 7.0 (\bullet) and at various charcoal-drug ratios. Mean \pm SEM of 3 experiments. From Neuvonen & Olkkola (1984a).

acids at a low pH and bases at a high pH (Andersen 1947; Hauge & Willamann 1927). However, the change in gastric pH has no significant effect on the antidotal efficacy of oral activated charcoal (Olkkola & Neuvonen 1984a).

1.6 Gastrointestinal Contents

Gastrointestinal contents, probably like any other competing solutes, impair the adsorption of drugs to activated charcoal (Andersen 1948). However, the effect of gastrointestinal contents on the antidotal efficacy of charcoal is complicated. Although the presence of food in the stomach of patients with drug overdosages somewhat impairs the adsorption capacity of charcoal, it gives charcoal more time to effectively adsorb drugs in the gastrointestinal canal (fig. 3), possibly by slowing the gastric emptying rate (Olkkola & Neuvonen 1984b).

Concomitant ethanol ingestion is common in overdosages. Ethanol is an organic solvent and could thus affect the antidotal efficacy of charcoal. Ethanol impairs the adsorption capacity of charcoal *in vitro* but it has only minimal effects in humans (Neuvonen et al. 1984). Accordingly, there should be no hesitation in the administration of activated charcoal to bind other toxic compounds despite the potential ethanol ingestion. Unfortunately, charcoal does not prevent the absorption of ethanol.

1.7 Effects of Stability of the Charcoal-Poison Complex

The binding of drugs and poisons to activated charcoal is a reversible process, meaning that the

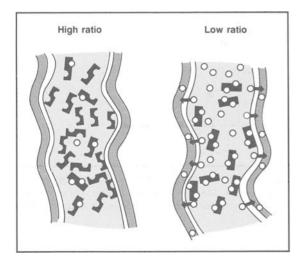


Fig. 2. Schematic representation of the effect of high and low charcoal-drug ratios on systemic drug absorption. At low ratios the amount of free drug (\bigcirc) and the subsequent absorption from the gastrointestinal tract (\square) into the blood stream (\square) is higher than at high ratios when there is sufficient charcoal (\blacksquare) to adsorb the free drug.

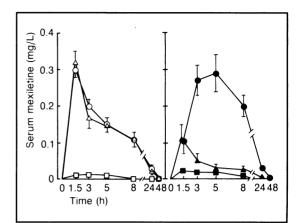


Fig. 3. Effect of activated charcoal and eating on the absorption of 200mg mexiletine, measured by serum mexiletine concentration. Mean \pm SEM in 6 volunteers. From Olkkola & Neuvonen (1984b). *Key:* \bigcirc = control; \square = mexiletine + 25g charcoal; \triangle = mexiletine + charcoal 1 hour later; \bullet = mexiletine + food; \blacksquare = mexiletine + food + 25g charcoal; \triangle = mexiletine + food + charcoal 1 hour later.

compound once adsorbed can also desorb (Bainbridge et al. 1977). This has also been demonstrated in humans with certain drugs (Levy & Tsuchiya 1972; Neuvonen & Olkkola 1986; Neuvonen et al. 1978; Olkkola 1985b). The excretion of salicylates into urine from 24 to 72 hours after administration of aspirin has been higher when taken with activated charcoal than without (fig. 4). The apparent half-life of drugs has been prolonged when ingested with a single dose of charcoal (Alvan 1973). This indirectly indicates desorption from charcoal leading to subsequent absorption of the drug. However, if adequately high doses of activated charcoal are used the desorption is seldom significant in clinical situations, but it can somewhat increase the total amount of drug or poison absorbed. The use of repeated doses of charcoal increases its efficacy (Crome et al. 1977; Dawling et al. 1978) and reduces the risk of desorption.

1.8 Time-Effect Relationships

Delay in the administration of charcoal after drug ingestion no doubt impairs the antidotal efficacy of oral activated charcoal. Accordingly, charcoal should be given as soon as possible and should be a part of first-aid kits both at home and at work.

The rate of absorption of most orally administered agents is directly related to the rate at which these agents pass from the stomach to the intestine (Nimmo 1979). The factors affecting gastric emptying rate and the formulation of the agent ingested thus modify the antidotal efficacy of oral activated charcoal. The absorption of drugs in life-threatening overdosages may be considerably prolonged (Rosenberg et al. 1981). Therefore, there is no exact time at which charcoal should no longer be administered to prevent gastrointestinal absorption in intoxications.

1.9 Effects of Purgatives and Whole-Bowel Irrigation

Theoretically, the quicker a poison passes through the gastrointestinal tract the less will be its desorption, provided that hyperperistalsis and increased fluid secretion do not facilitate desorption. It has been claimed that purgative-enhanced propulsion of charcoal-poison complex would improve the antidotal efficacy of oral activated charcoal. Thus, many authors recommend the use of saline purgatives as an adjunct to charcoal to hasten the elimination of charcoal-poison complex (Boehnert et al. 1985; Cupit & Temple 1984; Daunderer 1983; Krenzelok 1985; Minocha et al. 1985; Oderda 1979; Rumack 1980; Shannon et al. 1986; Teschke 1984).

In animals, purgatives have either improved (Chin et al. 1981; Gaudreault et al. 1985; Laass 1980; Picchioni et al. 1982) or diminished (Galey et al. 1987; Van de Graaff et al. 1982) the antidotal efficacy of charcoal. Most studies in humans have failed to demonstrate any substantial benefit from the combined use of purgatives and charcoal (Easom et al. 1982; Galinsky & Levy 1984; Mayersohn et al. 1977; Neuvonen & Olkkola 1986; Sketris et al. 1982) [fig. 5].

Goldberg et al. (1987) were able to increase the efficacy of repeated doses of oral activated charcoal with sorbitol. They administered charcoal or charcoal combined to sorbitol to healthy volunteers who

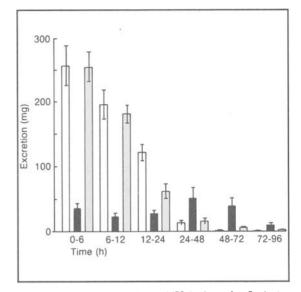


Fig. 4. Effect of activated charcoal (50g), given after 5 minutes (\blacksquare or 1 hour (\square , on the absorption of aspirin 1000mg, measured by the excretion of salicylate into urine during various periods. Mean \pm SEM in 6 volunteers. From Neuvonen et al. (1978). Open bar (\square) indicates aspirin excretion without charcoal.

had taken 1200mg slow release theophylline 6 hours earlier. However, 2 of the 9 subjects developed severe adverse effects (from sorbitol) requiring medical intervention during the charcoal-sorbitol phase. In another recent study (Berg et al. 1987), sorbitolinduced diarrhoea may have accelerated the onset of the effect of charcoal on the elimination of intravenously administered phenobarbitone, although the overall benefit from sorbitol was minimal.

To date, the routine use of purgatives in combination with activated charcoal does not appear to be indicated. Even high doses of activated charcoal given as watery suspensions do not cause constipation in most patients. In fact, charcoal suspensions – without any laxatives – often cause diarrhoea. In some instances the use of laxatives may promote the evacuation of depot formulations or other slowly absorbed drugs from the gastrointestinal tract and thus have a beneficial effect together with activated charcoal. However, the benefits of combining, for example, sorbitol with charcoal must be weighed against the potential risks for the individual patient.

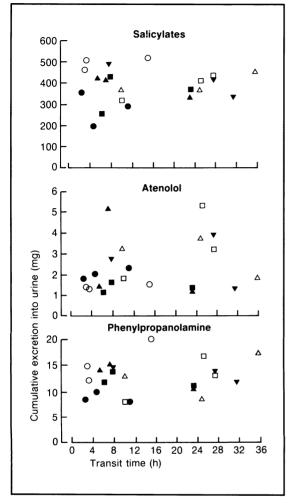


Fig. 5. Correlation between the gastrointestinal transit time (modified by the use of oral magnesium citrate, and rectal metoclopramide and bisacodyl) and the antidotal efficacy of charcoal reflected as the cumulative excretion of salicylates, atenolol and phenylpropanolamine into urine over 36 hours. Different symbols refer to 7 subjects. Activated charcoal (25g) was ingested 5 minutes after aspirin (1000mg), atenolol (100mg) and phenylpropanolamine (50mg). From Neuvonen & Olkkola (1986).

There are some reports of the effect of wholebowel irrigation on the absorption of drugs and toxic substances from the gastrointestinal tract (Tenenbein et al. 1986). As far as we know, there are no studies on the combined use of whole-bowel irrigation and oral activated charcoal. Theoretically, ingestion of charcoal prior to whole-bowel irrigation, or whole-bowel irrigation with some charcoal added to the irrigation solution, could enhance the efficacy of both charcoal and whole-bowel irrigation.

2. Mechanisms of Enhanced Drug and Toxin Elimination by Multiple Charcoal Doses

Many drugs and toxic agents are excreted into the gastrointestinal tract as parent substances, as active metabolites, or as conjugates, which may liberate active compounds in the gastrointestinal tract. The excretion may occur in gastric juice, bile, pancreatic secretions or other gastrointestinal fluids (McKinnon et al. 1986). Also, the diffusion through the mucosal ('dialysis') membrane from capillaries into the gut is possible. Irrespective of the exact mechanism of the excretion of individual substances, very many of these compounds are then reabsorbed into the blood. Activated charcoal effectively and practically irreversibly adsorbs many of those compounds in the gastrointestinal canal and prevents their reabsorption. The interruption of the enterohepatic and enteroenteric circulation by multiple oral doses of activated charcoal then accelerates the rate of drug elimination (fig. 6).

In some cases the binding of drugs and toxins to a single dose of activated charcoal is far from complete, although the charcoal has been given at an appropriate time. This may be due, for example, to inadequate administration of charcoal, to low affinity of the ingested poisons to charcoal, to the reversibility of adsorption of that particular substance, or to the saturation of the adsorbing capacity of charcoal by the drug or other gastrointestinal tract contents. Furthermore, the release rates of drugs from various pharmaceutical formulations and in various intoxications may vary considerably. This makes the adsorption to a single charcoal dose sometimes less complete than when multiple charcoal doses were given.

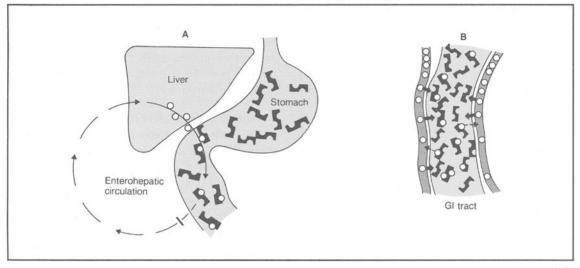


Fig. 6. Orally administered repeat-dose activated charcoal (Lan) prevents: (A) the enterohepatic circulation of drugs (O), and (B) favours their diffusion from blood () into gastrointestinal lumen by preventing back-diffusion ('gastrointestinal dialysis').

3. Effect of Single and Repeated Doses on the Absorption and Elimination of Drugs in Humans

3.1 Antipyretic Analgesics

3.1.1 Aspirin

In vitro, aspirin is adsorbed only moderately to activated charcoal. In humans, the absorption of therapeutic aspirin doses (1 to 5g) is reduced by 50 to 85% with 20 to 50g activated charcoal (Levy & Tsuchiya 1972; Neuvonen et al. 1978) [fig. 7].The peak serum concentrations are reduced more effectively, but because of the desorption of aspirin from the aspirin-charcoal complex (fig. 4) the reduction in total absorption is smaller.

Repeated doses of charcoal enhance the elimination of salicylates. This has been shown both in experimental animals (Wogan et al. 1986) and in intoxicated patients (Boldy & Vale 1986; Hillman & Prescott 1985; Prescott et al. 1986). Some charcoal preparations contain considerable amounts of sodium bicarbonate which may contribute to the shortened elimination half-life of salicylates by increasing urinary pH and excretion of salicylates. In any case, repeated doses of charcoal are indicated in aspirin poisoning, as they also reduce the risk of desorption.

3.1.2 Paracetamol

The adsorption of paracetamol to activated charcoal has been studied both in vitro and in vivo (Dordoni et al. 1973; Galinsky & Levy 1984; Levy & Houston 1976; Neuvonen et al. 1983c). The efficacy of activated charcoal alone has been only moderate in experimental studies (fig. 7) and in paracetamol poisonings saturation of the adsorption capacity of charcoal is likely to occur. In severe paracetamol poisoning specific antidotes, acetylcysteine or methionine, must be used in order to prevent hepatic necrosis. In vitro these antidotes are adsorbed to charcoal to some extent (Klein-Schwarz & Oderda 1980). However, this adsorption does not seem to be clinically significant and does not invalidate the concomitant oral administration of charcoal and acetylcysteine (North 1981a; Rybolt et al. 1986).

The elimination half-life of paracetamol is prolonged in severe poisoning. At least theoretically, charcoal may bind paracetamol and its toxic metabolites and thus reduce their hepatic toxicity.

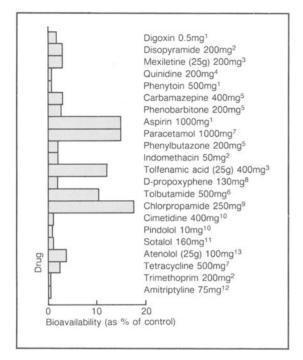


Fig. 7. Inhibition of drug absorption by activated charcoal (Carbomix). Charcoal 50g (except where indicated) or water (control) was ingested 5 minutes after the drug. Mean values in 5 to 7 volunteers. *Reference key*: 1 Neuvonen et al. 1978; 2 Neuvonen & Olkkola 1984a; 3 Olkkola & Neuvonen 1984b; 4 Neuvonen et al. 1984; 5 Neuvonen & Elonen 1980; 6 Neuvonen et al. 1983b; 7 Neuvonen et al. 1983c; 8 Kärkkäinen & Neuvonen 1985; 9 Neuvonen & Kärkkäinen 1983; 10 Neuvonen & Olkkola 1984b; 11 Kärkkäinen & Neuvonen 1984; 12 Kärkkäinen & Neuvonen 1986; 13 Neuvonen & Olkkola 1986.

3.1.3 Indomethacin

Activated charcoal adsorbs indomethacin effectively both *in vitro* and in humans (figs 1 and 7). Saturation of the adsorption capacity of charcoal in indomethacin poisoning is unlikely (Neuvonen & Olkkola 1984a). Large amounts of conjugated indomethacin are excreted into the bile and reabsorbed. It is therefore to be expected that repeated doses of charcoal would enhance its elimination, but this has not been studied.

3.1.4 Phenylbutazone and Other Pyrazolone Derivatives

Activated charcoal inhibits very effectively the absorption of phenylbutazone (fig. 7). The effect of

charcoal on the elimination of this drug is only moderate (fig. 8) despite its long elimination halflife (Neuvonen & Elonen 1980). The effect of charcoal on oxyphenbutazone and azapropazone absorption is probably similar to its effect on phenylbutazone absorption.

3.1.5 Tolfenamic Acid (Fenamates)

The affinity of tolfenamic acid to activated charcoal is good, and charcoal moderately prevents the absorption of tolfenamic acid (fig. 7) and mefenamic acid in humans (El-Bahie et al. 1985; Olk-kola & Neuvonen 1984a,b). In experimental animals charcoal has also reduced the toxicity of mefenamic acid (Glazko 1967).

The elimination half-life of fenamates is rather short, but some of the metabolites appear to undergo significant enterohepatic circulation and slow elimination. Repeated doses of activated charcoal may enhance the elimination of the fenamate metabolites.

3.1.6 Piroxicam

The absorption of 20mg piroxicam was reduced by 98% by 50g activated charcoal. Repeated oral doses of charcoal shortened the elimination halflife of piroxicam from the control value of 48 hours to 22 hours (Laufen et al. 1984).

3.1.7 Other Antipyretic Analgesics

Activated charcoal is likely to adsorb sulindac, ibuprofen, naproxen and most other antipyretic analgesics, and inhibit their absorption from the gastrointestinal tract. It is to be expected that the enterohepatic circulation of sulindac (Dujovne et al. 1983), and possibly of some other antipyretic analgesics or their metabolites, can be interrupted by activated charcoal. Accordingly, repeated doses of charcoal could enhance the elimination of these drugs.

3.2 Other Analgesics

3.2.1 Dextropropoxyphene

The adsorption of dextropropoxyphene to charcoal is good *in vitro* (Corby & Decker 1968). Activated charcoal 4g, given over 10 to 50 minutes

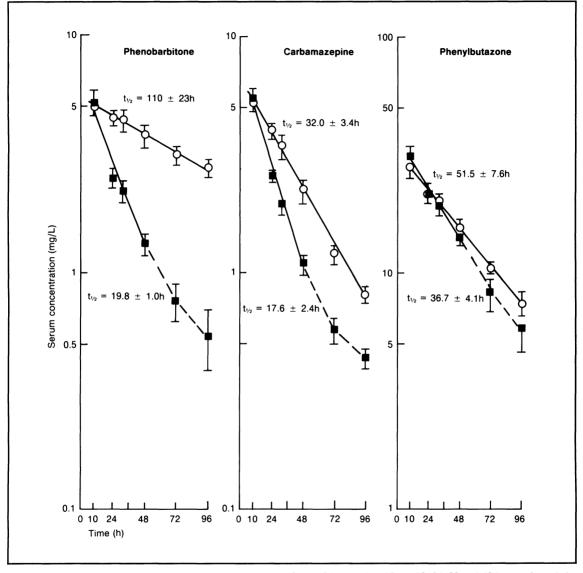


Fig. 8. Effect of activated charcoal (**II**) given in multiple doses (50g at 10 hours, then 17g at 12, 24, 36 and 48 hours after administration of drug), compared with control (\bigcirc) on t_{1/2} of phenobarbitone, carbamazepine and phenylbutazone. Mean \pm SEM in 5 volunteers. From Neuvonen & Elonen (1980).

after the intake of dextropropoxyphene, reduced its gastrointestinal absorption by 50% (Chernish et al. 1972). 50g charcoal, given immediately after 130mg dextropropoxyphene, reduced its absorption by 97% (Kärkkäinen & Neuvonen 1985) [fig. 7]. It is obvious that 50g activated charcoal can adsorb more

than 90% of the dextropropoxyphene present in the stomach of intoxicated patients at the time of the administration of charcoal. Furthermore, repeated doses of oral charcoal will enhance the elimination of both dextropropoxyphene and norpropoxyphene (Kärkkäinen & Neuvonen 1985).

3.2.2 Opiates and Other Opioid Analgesics

There is no reason to believe that activated charcoal would not adsorb morphine, codeine, pentazocine, oxycodone, tilidine (Cordonnier et al. 1986), buprenorphine, methadone and other related substances and inhibit their absorption from the gastrointestinal canal. Because at least methadone and buprenorphine undergo enterohepatic circulation and have a long elimination half-life, charcoal is likely to enhance their elimination, too.

3.2.3 Nefopam

Nefopam has caused several fatal intoxications. It has a good affinity to activated charcoal both *in vitro* and in animals. In mice it has reduced acute toxicity by a factor of 5 (Neuvonen et al. 1983d). The effect of charcoal on the elimination of nefopam is unknown.

3.3 Hypnosedatives and Anticonvulsants

3.3.1 Barbiturates

Activated charcoal binds most barbiturates very effectively. For instance, 50g activated charcoal adsorbed 97% of phenobarbitone (200mg) present in the stomach (Neuvonen & Elonen 1980) [fig. 7].

Normally, phenobarbitone has an elimination half-life of about 100 hours, which repeated doses of oral activated charcoal have shortened to about 20 hours (Neuvonen & Elonen 1980) [fig. 8]. The effect of charcoal has also been shown in intoxicated patients and following intravenous administration of phenobarbitone (Berg et al. 1982, 1987; Goldberg et al. 1985a; Linden et al. 1983; Pond et al. 1984; Vale et al. 1986). Repeated doses of oral activated charcoal should also hasten recovery from phenobarbitone poisoning (Boldy et al. 1986; Goldberg & Berlinger 1982; Prescott et al. 1986; Vale et al. 1986), although that effect was not seen in a controlled study, despite a clear effect on phenobarbitone clearance (Pond et al. 1984). The effect of charcoal on the elimination of other barbiturates has not been studied systematically. In some intoxicated patients elimination has been unaffected by administration of charcoal (Neuvonen 1982).

Orally administered activated charcoal has resulted in a great shortening (80 to 90%), both in half-life and sleep time, in mice following intravenous injection of phenobarbitone (and also of methyprylone, glutethimide, ethchlorvynol and methaqualone), but no significant effect on sleep time following amylobarbitone (amobarbital) or pentobarbitone was noted (Adler et al. 1986).

3.3.2 Meprobamate

Activated charcoal effectively adsorbs meprobamate and inhibits its absorption. Given in multiple doses, charcoal enhances the elimination of meprobamate in overdosages (Hassan 1986; Linden & Rumack 1984).

3.3.3. Benzodiazepines

Charcoal inhibits the absorption of benzodiazepines. Small repeated doses of charcoal did not seem to affect the elimination of diazepam (Korttila et al. 1976). However, 40g oral activated charcoal given 4-hourly seemed to shorten the elimination half-life of diazepam greatly in an intoxicated patient (Traeger & Haug 1986).

3.3.4 Glutethimide

In vitro, glutethimide is efficiently adsorbed to activated charcoal (Decker et al. 1968). Charcoal inhibits the gastrointestinal absorption of glutethimide and probably also increases its elimination (Adler et al. 1986; Fiser et al. 1971; Hayden & Comstock 1975).

3.3.5 Phenytoin

Charcoal effectively inhibits the gastrointestinal absorption of phenytoin. Activated charcoal 50g reduced the gastrointestinal absorption of 500mg phenytoin by 98 to 99% when given after 5 minutes (fig. 7), and by 80% when given 1 hour later (Neuvonen et al. 1978). Because of the slow absorption of phenytoin, charcoal is a useful antidote even several hours after ingestion.

Phenytoin metabolism saturates even at therapeutic doses, and in intoxications elimination is very slow. Repeated doses of charcoal may enhance its elimination (Prescott et al. 1986), although this effect was not clear in another study (Pond et al. 1984).

3.3.6 Carbamazepine

Charcoal inhibits the absorption of carbamazepine effectively in humans. Over 95% of the carbamazepine dose (400mg) present in the stomach was adsorbed to 50g charcoal (Neuvonen & Elonen 1980) [fig. 7]. Repeated charcoal doses have increased the elimination rate of carbamazepine by about 50% in healthy volunteers (Neuvonen & Elonen 1980) [fig. 8] and in intoxicated patients (Boldy et al. 1987; Heath & Van Loo 1986; Vale et al. 1986).

3.3.7 Valproate

The absorption of valproate is inhibited moderately by oral activated charcoal, but not as effectively as that of carbamazepine, phenobarbitone and phenytoin (Neuvonen et al. 1983b). Repeated doses of charcoal also seem to increase its elimination (Prescott et al. 1986).

3.4 Antidepressants

3.4.1 Nortriptyline

Single dose charcoal effectively inhibits the gastrointestinal absorption of nortriptyline but seems to prolong its elimination half-life (Alvan 1973). This suggests desorption of nortriptyline from the charcoal-drug complex, which can be prevented by the administration of repeated doses of activated charcoal (Crome et al. 1977; Dawling et al. 1978). Repeated doses of charcoal may also increase the elimination of nortriptyline to some extent (Kärkkäinen & Neuvonen 1986).

3.4.2 Amitriptyline

Charcoal binds amitriptyline efficiently both *in vitro* and in humans. In healthy volunteers 99% of amitriptyline (75mg) present in the stomach was adsorbed to 50g activated charcoal (Kärkkäinen & Neuvonen 1986) [fig. 7].

Repeated doses of charcoal shortened the elimination half-life of amitriptyline by 20% in healthy volunteers. Elimination of its active metabolite, nortriptyline, was shortened by 40% (Kärkkäinen & Neuvonen 1986). It has been claimed that repeated doses of oral activated charcoal would reduce the elimination half-life of amitriptyline from 40 hours to 4 to 10 hours in intoxicated patients (Swartz & Sherman 1984), but the kinetic data reported do not justify that conclusion. The effect of charcoal on the elimination of amitriptyline is obviously not so dramatic (Prescott et al. 1986).

3.4.3 Imipramine

Charcoal effectively prevents the gastrointestinal absorption of imipramine but did not enhance its elimination (Goldberg et al. 1985b). This may be partly due to the elimination half-life (9 to 10 hours) of imipramine, which is relatively short in comparison with the elimination half-lives of amitriptyline and nortriptyline.

3.4.4 Doxepin

A single dose of 15g activated charcoal administered 30 minutes after doxepin 50mg inhibited 50% of its absorption but caused an apparent prolongation of the elimination half-life. This was due to the desorption of doxepin from charcoal. Repeated doses of charcoal did not reduce the elimination half-life of doxepin, but the clearance of desmethyldoxepin was increased by 57% (Scheinin et al. 1985).

3.4.5 Other Antidepressants

It is to be expected that activated charcoal also inhibits the absorption of other tricyclic and tetracyclic antidepressants. Thus, the immediate administration of charcoal in overdosages with these drugs is certainly indicated and in severe poisoning repeated dosing should be carried out.

3.5 Other Psychopharmaca

3.5.1 Phenothiazines

In vitro, activated charcoal adsorbs the phenothiazines tested well. As little as 100mg activated charcoal mixed with 50mg promazine before ingestion reduced absorption by 60% (Sorby 1965). There is no reason to expect that charcoal would not inhibit the absorption of other phenothiazines even better when used in adequate doses. Several phenothiazines have a relatively long elimination half-life and at least some enterohepatic circulation. Accordingly, repeated doses of oral activated charcoal may be useful in acute intoxications.

3.6 Cardiac Glycosides

3.6.1 Digoxin

Charcoal adsorbs digoxin effectively both *in vitro* and in humans. About 98% of therapeutic digoxin doses (0.5mg) was adsorbed to 50g activated charcoal (Neuvonen et al. 1978) [fig. 7]. Activated charcoal may be the most effective treatment in inhibiting the gastrointestinal absorption of digoxin, since the total amount of digoxin ingested is small (less than 100mg) even in severe digoxin poisonings. Thus, the risk of saturation of the adsorption capacity of charcoal is small.

Digoxin undergoes significant enterohepatic cycling in the body, and this can be interrupted by repeated doses of oral activated charcoal. The elimination half-life of digoxin has been shortened by about 50% (fig. 9) by repeat-dose charcoal (Boldy et al. 1985; Lalonde et al. 1985).

3.6.2 Other Cardiac Glycosides

Activated charcoal effectively adsorbs digitoxin. The relative efficacy of charcoal in digitoxin poisoning is about at the same level as in digoxin poisoning. Digitoxin has a more significant enterohepatic circulation than digoxin, and repeated doses of charcoal have shortened its elimination half-life by 90% (Pond et al. 1981).

The effect of charcoal on the absorption of other cardiac glycosides is probably at the same level as with digoxin and digitoxin. Repeated doses of charcoal may also enhance to some extent the elimination of other cardiac glycosides (Belz & Bader 1974).

3.7 Antiarrhythmic Drugs

3.7.1 Quinidine

Activated charcoal inhibits the gastrointestinal absorption of quinidine very effectively. Charcoal 50g given 5 minutes after quinidine (200mg) re-

Fig. 9. Serum digoxin concentration during control (\bullet) and charcoal (\blacksquare) phases. Mean \pm SEM in 10 subjects. From Lalonde et al. (1985).

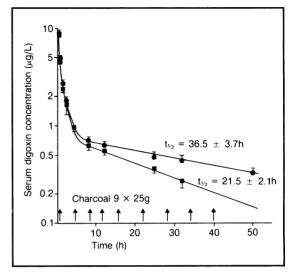
duced its absorption by 99% (Neuvonen et al. 1984) [fig. 7]. The adsorption capacity of charcoal is unlikely to be saturated even in severe quinidine poisoning. Charcoal most likely inhibits the absorption of quinidine more effectively than gastric lavage or ipecac-induced vomiting. Its effect on quinidine elimination is not known, but the elimination of quinidine's optical isomer, quinine, is accelerated by charcoal (Prescott et al. 1986).

3.7.2 Disopyramide

In humans 50g of charcoal prevented the absorption of over 90% of therapeutic doses (200mg) of disopyramide (Neuvonen & Olkkola 1984a). The efficacy of charcoal is also likely to be at least moderate in intoxications, but it should be given in as high doses as feasible to minimise the potential risk of saturation of its adsorption capacity (figs 1 and 7). The effect of charcoal on disopyramide elimination has not been studied.

3.7.3 Mexiletine

Oral activated charcoal inhibits the absorption of mexiletine well in humans. Activated charcoal 25g given 5 minutes after 200mg mexiletine inhib-



ited absorption by over 95% (Olkkola & Neuvonen 1984b) [figs 3 and 7]. Data concerning the effect of charcoal on elimination of mexiletine are lacking.

3.7.4 Other Antiarrhythmic Drugs

Activated charcoal is likely to adsorb most antiarrhythmic drugs and prevent the gastrointestinal absorption of, for example, verapamil, nifedipine, diltiazem, propafenone and amiodarone. The absorption of flecainide 200mg was almost completely prevented when activated charcoal was ingested immediately (Nitsch et al. 1987).

Amiodarone has an exceptionally long elimination half-life and a significant enterohepatic circulation. Thus, repeated doses of oral activated charcoal should be effective in overdosages.

3.8 β -Adrenoceptor Antagonists

In humans, 50g activated charcoal reduced the absorption of atenolol, pindolol and sotalol, given in therapeutic doses, by over 90% (Kärkkäinen & Neuvonen 1984; Neuvonen & Olkkola 1984b, 1986) [fig. 7]. Syrup of ipecac allowed 30-fold greater absorption of pindolol compared with charcoal (Neuvonen & Olkkola 1984b) [fig. 10]. Because of the varying potencies of the β -blocking agents, the amounts ingested in acute overdosages usually range from 100mg (e.g. pindolol) to 10g (e.g. atenolol and sotalol). Based on adsorption studies *in vitro*, it is obvious that the risk of saturation of the adsorption capacity of charcoal is small if the intoxication is caused by a β -blocking agent used therapeutically in small amounts.

Repeated doses of charcoal have been shown to increase elimination of sotalol (Kärkkäinen & Neuvonen 1985) and nadolol (Du Souich et al. 1982).

3.9 Oral Antihyperglycaemic Drugs

Based solely on limited *in vitro* experiments at low pH, where tolbutamide is only sparingly soluble (Decker et al. 1968), activated charcoal was recently erroneously claimed to be ineffective in poisonings caused by tolbutamide (Boyd 1982; Derlet & Albertson 1986). Charcoal effectively adsorbs *in vitro* carbutamide, chlorpropamide, glibenclamide (glyburide), glipizide, tolazamide and tolbutamide (Kannisto & Neuvonen 1984). The

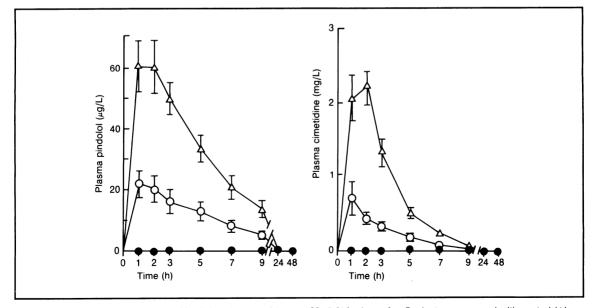


Fig. 10. Effect of activated charcoal (50g) [\bullet] and syrup of ipecac (20ml) [\bigcirc], given after 5 minutes, compared with control (\triangle), on the absorption of pindolol (10mg) and cimetidine (400mg) as measured by plasma concentration. Mean \pm SEM in 7 subjects. From Neuvonen & Olkkola (1984b).

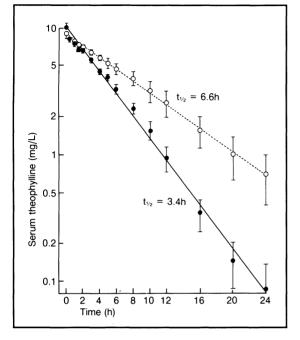


Fig. 11. Serum theophylline concentrations after intravenous infusion of aminophylline (6 mg/kg) with (\bullet) and without (\bigcirc) activated charcoal (40g at 0 hours and 20g at 2, 4, 6, 9 and 12 hours). Mean \pm SEM in 5 subjects. From Berlinger et al. (1983).

adsorption of other sulphonylureas is most likely similar; the affinity of the second generation sulphonylureas to charcoal is especially good. Due to their high potency, therapeutic doses are small, only some milligrams. This results in high charcoal-drug ratios, even in potentially severe overdosages. Accordingly, charcoal is expected to be very effective in intoxications caused by second generation sulphonylureas.

The immediate administration of 50g charcoal inhibited 80 to 90% of the absorption of therapeutic chlorpropamide and tolbutamide doses in healthy volunteers (Neuvonen & Kärkkäinen 1983; Neuvonen et al. 1983b) [fig. 7]. Due to high therapeutic doses of tolbutamide and chlorpropamide, the adsorption capacity of charcoal is likely to be saturated in severe intoxications caused by them (but not by glibenclamide or glipizide).

Repeated doses of charcoal did not enhance the elimination of chlorpropamide (Neuvonen & Kärkkäinen 1983). The effect of charcoal on the

elimination of other sulphonylureas has not been studied.

The effect of activated charcoal on the absorption and elimination of biguanides, like buformin, metformin and phenformin, is unknown. On the other hand, repeated doses of charcoal seem to accelerate the elimination of M79175, a new inhibitor of aldose reductase (Arimori et al. 1987).

3.10 Bronchodilating Drugs

Activated charcoal reduces the gastrointestinal absorption of theophylline (Sintek et al. 1979), which is often used as a slowly absorbed preparation. In these cases the efficacy of single charcoal doses is not sufficient to prevent toxicity (Lim et al. 1986; Neuvonen et al. 1983c) and hence repeated doses of charcoal should be used in severe theophylline poisoning to prevent prolonged absorption.

Oral activated charcoal increases the elimination of both orally and intravenously administered theophylline (Gal et al. 1984; Goldberg et al. 1987; Park et al. 1983, 1984; Radomski et al. 1984; Rygnestad et al. 1986 [fig. 11]. However, as theophylline elimination is relatively fast (elimination half-life, 6 to 9 hours), increased elimination may not be as important as it is in poisonings with drugs which have long elimination half-lives. It may be more important to prevent the prolonged gastrointestinal absorption by repeated charcoal doses (fig. 12).

The effect of charcoal on the absorption and elimination of salbutamol (albuterol), terbutaline, and other sympathomimetic drugs is unknown. There is no reason to believe that charcoal would not be effective in preventing absorption of sympathomimetics from the gastrointestinal tract, particularly as their therapeutic doses are small.

3.11 Antimicrobial Agents

Antimicrobial agents are seldom a problem in clinical toxicology. Therefore, interest in the effect of charcoal on the absorption and elimination of these agents has been limited. Oral activated char-

coal inhibits the gastrointestinal absorption of, for example, tetracycline, doxycycline and trimethoprim (Neuvonen & Olkkola 1984a; Neuvonen et al. 1983c; Venho et al. 1978). It also inhibits the absorption of para-aminosalicylic acid, but at low charcoal-drug ratios its adsorption capacity is saturated (Olkkola 1985b). In healthy volunteers, the absorption of isoniazid was poorly inhibited if charcoal was delayed 60 minutes (Scolding et al. 1986). In animals, at a charcoal-drug ratio of 8:1, charcoal has effectively reduced the absorption and toxicity of isoniazid and chloroquine (Chin et al. 1970, 1973; Picchioni et al. 1966). Due to the long half-life of chloroquine, multiple dosing of oral charcoal seems to be worth trying in patients with acute or chronic intoxications.

Dapsone is an antimicrobial agent which can cause severe poisoning (Elonen et al. 1979; Reigart et al. 1982). Activated charcoal reduces the gastrointestinal absorption of dapsone and increases the elimination of both dapsone and the metabolite, monoacetyldapsone (Neuvonen et al. 1980). The efficacy of repeat-dose oral charcoal in dapsone intoxications is comparable to that of haemodialysis (Neuvonen et al. 1983a) [fig. 13].

Clioquinol (iodochlorhydroxyquin) seems to undergo enterohepatic circulation in considerable amounts, at least in experimental animals (Kotaki et al. 1984). Because of its potential role in subacute myelo-optic neuropathy, repeated doses of oral activated charcoal should be used in clioquinol overdosages to increase elimination.

3.12 Hormones

Hormones do not have much toxicological significance. Charcoal adsorbs steroid hormones and interferes with the enterohepatic circulation of, for example, oestriol (Heimer & Englund 1986). The concurrent administration of oral contraceptives

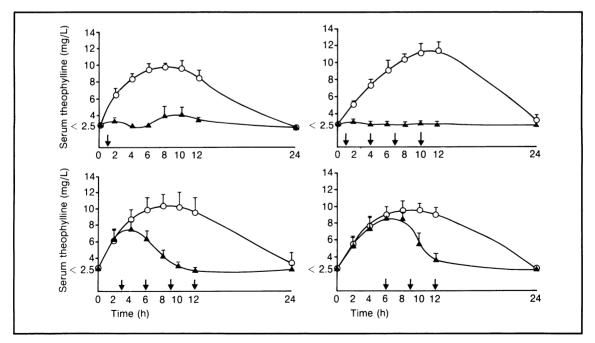


Fig. 12. Serum theophylline concentrations in children (mean age 12 years) after single doses of slow release theophylline (10 mg/ kg), with (Δ) or without (\bigcirc) activated charcoal. During the charcoal phase of the crossover study theophylline ingestion was followed by single or repeated doses (1 g/kg) of activated charcoal as indicated by arrows. Mean \pm SEM in 5 children in each group. From Lim et al. (1986).

Fig. 13. Effect of orally administered charcoal on serum concentrations of dapsone (\bullet) and monoacetyldapsone (MADDS) [O] and on urinary excretion rate of dapsone (\Box) in a patient intoxicated with dapsone (70 × 100mg). The first control half-lives on days 1 to 4 are based on the data between 3 haemodialyses ($\uparrow \uparrow \uparrow$). The second half-lives are based on the data during charcoal administration (20g 4 times/day) on days 4 to 7. From Neuvonen et al. (1983a).

and charcoal can result in the loss of contraceptive efficacy.

3.13 Other Drugs

3.13.1 Phenylpropanolamine

Phenylpropanolamine has a good affinity to activated charcoal *in vitro* (Tsuchiya & Levy 1972). In humans the absorption of phenylpropanolamine is inhibited well, or at least moderately. Efficacy seems to be dependent on the pharmaceutical formulation of phenylpropanolamine. When a single dose of 25g charcoal was ingested 5 minutes after 50mg phenylpropanolamine as a slowly absorbed formulation, absorption was inhibited by only 50% (Neuvonen & Olkkola 1986). The efficacy of charcoal has been considerably better when a rapidly absorbed formulation is employed (Tsuchiya & Levy 1972). Most probably, charcoal should be given in multiple doses in poisonings caused by slow release formulations of phenylpropanolamine.

3.13.2 Histamine Antagonists

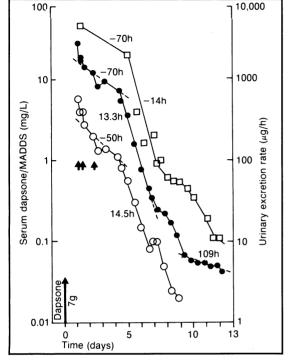
Many classical histamine antagonists (H_1 -receptor blockers) have caused severe intoxications. The antihistamines are probably well adsorbed to activated charcoal. Activated charcoal (50g) given within 5 minutes of 200mg diphenylhydramine reduced its absorption by 95% in healthy volunteers (Guay et al. 1984). Astemizole and its active metabolites have an exceptionally long halflife of several days and their elimination could possibly be accelerated by repeat-dose charcoal. The toxicity of H₂-receptor blocking agents is relatively low and, in any case, activated charcoal effectively prevents the gastrointestinal absorption of cimetidine, for example (Neuvonen & Olkkola 1984b) [figs 7 and 10].

3.13.3 Diuretics

The epoxide metabolites of frusemide (furosemide) may cause hepatic necrosis in high overdoses. A small dose of charcoal (8g) completely prevented (99.5%) the absorption of a therapeutic dose (40mg) of frusemide (Neuvonen et al. 1987). Thus, the usual antidotal dose of charcoal binds even high toxic doses of frusemide well. The absorption of thiazide diuretics can probably also be prevented by activated charcoal. Some thiazide diuretics and chlorthalidone undergo significant enterohepatic circulation, which could be affected by repeat-dose charcoal.

3.13.4 Propantheline

Activated charcoal 5g ingested simultaneously with 45mg propantheline significantly reduced the pharmacological effects, suggesting an inhibition of propantheline absorption (Chaput de Saintonge & Herxheirner 1971).



3.13.5 Methotrexate

Activated charcoal adsorbs methotrexate well, both *in vitro* and in humans. Repeated doses of oral charcoal also increase its elimination (Gadgil et al. 1982), which could have some clinical significance with high dose methotrexate treatment in patients with renal failure.

3.13.6 Cyclosporine

Oral activated charcoal is able to adsorb cyclosporine and to increase its elimination (Honcharic & Anthone 1985).

3.13.7 Syrup of Ipecac

Charcoal adsorbs the active ingredients of syrup of ipecac (Cooney 1978). However, when a 5-minute delay occurred between the administration of syrup of ipecac (60ml, i.e. twice the usual dose) and activated charcoal (50g), ipecac-induced emesis occurred in 80% of subjects (Krenzelok et al. 1986). In a clinical study, activated charcoal (50g), given 10 minutes after syrup of ipecac (60ml), did not prevent the emetic effect in any of the 10 intoxicated patients (Freedman et al. 1987).

3.13.8 Acetylcysteine and Methionine

Although *in vitro* acetylcysteine and methionine are adsorbed slightly to activated charcoal (Klein-Schwarz & Oderda 1980) the absorption of acetylcysteine is not prevented by oral charcoal in man (North et al. 1981a; Rybolt et al. 1986).

3.14 Metals, Alcohols and Other Substances

3.14.1 Metals

The efficacy of charcoal in human metal poisoning has not been studied. Some metallic salts, mercuric chloride for example, seem to be fairly well adsorbed *in vitro* (Andersen 1945). Orally administered multiple-dose charcoal may increase the elimination of some metals from the body. However, most metals, including lithium and iron, are not efficiently adsorbed to activated charcoal.

3.14.2 Alcohols

Activated charcoal adsorbs *in vitro* up to 300 to 400mg ethanol per gram of charcoal (Andersen 1947; Smith et al. 1967). In dogs, charcoal significantly reduced (or possibly only postponed) the absorption of ethanol (North et al. 1981b). However, in humans charcoal does not significantly affect the absorption of ethanol (Hulten et al. 1986; Minocha et al. 1986; Neuvonen et al. 1984). Although some reviews (Cupit & Temple 1984) still claim that ethanol is well adsorbed to charcoal, activated charcoal is not an effective antidote against ethanol intoxication.

The efficacy of charcoal in methanol poisoning seems to be comparable to its low efficacy in ethanol poisoning (Picchioni 1970). However, there are few experimental data on the effect of charcoal on methanol toxicity.

In animals, activated charcoal seems to reduce the mortality in ethylene glycol poisoning (Szabuniewicz et al. 1975), even though *in vitro*, charcoal adsorbs ethylene glycol relatively inefficiently (Cooney 1977b). This apparent discrepancy may be caused by the toxic metabolites of ethylene glycol, which in acute poisoning may be of major importance (Jacobsen & McMartin 1986).

3.14.3 Other Substances

Activated charcoal binds well to paraquat *in vitro.* In animals charcoal is at least as effective as Fuller's earth in reducing the toxicity of paraquat (Gaudreault et al. 1985; Okonek et al. 1982), but its antidotal efficacy in human paraquat poisoning has not been studied. The absorption and toxicity of T-2 mycotoxin is reduced by activated charcoal even if some time (e.g. 3 hours in the rat) has elapsed since exposure (Galey et al. 1987).

High doses of oral activated charcoal also reduce gastrointestinal absorption of kerosene, benzene and dichlorethane in rats (Chin et al. 1969; Laass 1980). The effect of repeated doses of charcoal on the elimination of paraquat, kerosene, benzene, dichlorethane and other organic solvents is unknown.

Activated charcoal adsorbs many other substances well (table I), including nicotine, strychnine and aflatoxins (Decker & Corby 1980). However, cyanide is not adsorbed significantly to charcoal. Even pretreatment with charcoal does not reduce the toxicity of potassium cyanide in mice (Neuvonen, unpublished results).

4. Relative Efficacy of Activated Charcoal, Emetics, and Gastric Lavage in Inhibiting Drug Absorption

Induced emesis seems to be more efficient than gastric lavage in emptying the stomach (Abdallah & Tye 1967; Arnold et al. 1959; Boxer et al. 1969; Corby et al. 1967). The validity of these older studies has been questioned because of the later technical development in orogastric tubes. It has been claimed that currently used large orogastric tubes are able to remove larger volumes of gastric contents than the previously used tubes (Burke 1972; Wheeler-Usher et al. 1986). However, the question remains whether gastric emptying should be performed at all.

Kulig et al. (1985) have demonstrated that syrup of ipecacuanha does not alter the clinical course of poisoned patients who are alert on presentation to hospital. Recent experimental studies have shown that oral activated charcoal is superior to emetics in reducing the absorption of that fraction of the drug which is still present in the stomach when charcoal or emetic is administered (Curtis et al. 1984; Neuvonen & Olkkola 1984b; Neuvonen et al. 1983c) [figs 10 and 14]. In these studies therapeutic doses (from 10 to 2000mg) of various drugs have been used. Despite the apparent handicap of experimental studies when compared with clinical intoxications, the data are relevant to very many intoxications: most drugs will cause fatal intoxications in doses of a few grams, or even in much smaller doses. However, it is obvious that gastric emptying is likely to be more effective than activated charcoal alone in those poisonings where the amount of the drug ingested is large (e.g. aspirin and paracetamol poisonings) leading to saturation of the adsorption capacity of charcoal, or where the affinity of the particular agent to activated charcoal is poor (e.g. iron, table I).

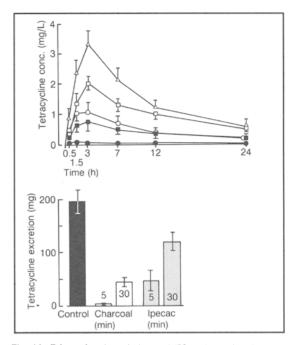


Fig. 14. Effect of activated charcoal (50g) given after 5 () or 30 (\bigcirc) minutes, and syrup of ipecac (20ml) given after 5 () or 30 minutes (\square), compared with tetracycline alone (\triangle) on the absorption of tetracycline (500mg), reflected as serum concentration and 4B hours' cumulative excretion of tetracycline. Mean \pm SEM in 3 volunteers. Tetracycline excretion was significantly less after charcoal than in the control group (p < 0.01), and was significantly less with charcoal after 5 minutes than ipecac after 5 minutes (p < 0.05) or charcoal after 30 minutes than ipecac after 30 minutes (p < 0.01). From Neuvonen et al. (1983c).

The immediate administration of activated charcoal before other time-consuming procedures should be considered in most intoxications. The efficacy of charcoal is expected to be good in poisonings where adsorption capacity will not be saturated, and its administration does not prevent gastric emptying being performed later. Charcoal in the stomach can adsorb drugs which otherwise could be absorbed to systemic circulation during transport to hospital and during the gastric emptying procedures.

5. Toxicity and Side Effects of Activated Charcoal

Activated charcoal has not of itself been associated with specific toxicity (Nau et al. 1958a,b, 1962). If: uraemic patients continuous treatment with oral charcoal 20 to 50 g/day over periods of 4 to 20 months has not resulted in any significant side effects (Yatzidis 1972), although rapid ingestion of large doses of charcoal may cause vomiting. Constipation or sometimes diarrhoea have occurred in some subjects receiving activated charcoal as a watery suspension.

Major complications associated with the use of activated charcoal have been due to the aspiration of charcoal and gastric contents (Harsch 1986; Pollack et al. 1981). There has been at least 1 case of charcoal-containing empyema (Justiniani et al. 1985) and 1 case of intestinal obstruction caused by huge amounts (300g) of charcoal (Anderson & Ware 1987). Possible constipation caused by activated charcoal can be treated with sorbitol, paraffin oil or lactulose. It should be noted that activated charcoal adsorbs many laxatives (e.g. sennosides) and prevents their action.

Some commercially available charcoal preparations contain large amounts of sorbitol or sodium bicarbonate. Repeated administration of these preparations in high doses may cause severe adverse effects (hypotension, electrolyte disturbances), themselves requiring medical intervention (Goldberg et al. 1987). Thus, the use of various extra agents in combination with activated charcoal must be weighed against the potential risks.

6. Conclusions

6.1 Role of Single Doses in Treatment of Intoxications

Single doses of oral activated charcoal effectively (or in some cases very effectively) prevent the gastrointestinal absorption of that fraction of most drugs and poisons which is present in the stomach when charcoal is given. Known exceptions are alcohols, cyanide, and metals such as iron and lithium.

In general, activated charcoal is superior to emetics in reducing absorption. If the amount of drug or poison ingested is large (e.g. aspirin and paracetamol poisonings) or if its affinity to charcoal is poor (e.g. iron poisoning), the adsorption capacity of activated charcoal can be saturated. In those instances, properly performed gastric emptying is likely to be more effective than charcoal alone.

In acute intoxications the drug history is often unreliable and thus, to be on the safe side, overtreatment is necessary. Although there is no exact time after which charcoal should not be administered to prevent gastrointestinal absorption, activated charcoal should be administered orally to acutely poisoned patients (adults 50 to 100g, children 1 g/kg) as soon as possible. The only exceptions are probably patients poisoned with caustic alkalis and acids. To avoid unnecessary delays in administration, activated charcoal should be a part of firstaid kits both at home and at work. In obtunded patients charcoal can be administered with a gastric tube, but tracheal intubation may be needed prior to administration. The 'blind' administration of charcoal neither prevents later gastric emptying nor does it cause serious adverse effects, as long as pulmonary aspiration in obtunded patients is prevented. Accordingly, there is nothing to prevent the liberal administration of charcoal in the immediate management of a poisoned patient.

6.2 Role of Repeated Doses in Treatment of Intoxications

6.2.1 Acute Intoxications

Repeated dosing with oral activated charcoal enhances the elimination of aspirin, carbamazepine, cyclosporine, dapsone, dextropropoxyphene, digitoxin, digoxin, meprobamate, nadolol, nortriptyline, phenobarbitone, phenytoin, piroxicam, valproate, sotalol and theophylline, to give some examples. It is to be expected that the efficacy of charcoal in accelerating the elimination of many other substances will be shown in the near future. Repeated doses of charcoal also reduce the risk of poison desorbing from the charcoal-poison complex.

Repeated dosing with oral activated charcoal seems to be indicated in severe poisonings until recovery or until plasma concentrations have fallen to non-toxic levels. Often doses of 20 to 50g activated charcoal as a watery suspension are given every 4 to 6 hours for 1 to 2 days. Constipation is seldom a problem but laxatives, such as sorbitol or lactulose, can be given with charcoal.

6.2.2 Chronic Intoxications

Repeated doses of oral activated charcoal may prove very useful in accelerating the elimination of many industrial and environmental toxicants, like dioxins, polychlorinated biphenyls, and some heavy metals and their radioactive isotopes, thus reducing the long term adverse effects of these toxicants. Moderate doses of activated charcoal, e.g. 10 to 20g every 6 to 8 hours, can be used for several months without significant side effects, although possible interference with gastrointestinal absorption of essential therapeutic drugs should be kept in mind during charcoal use. Further studies are needed to establish the value of repeated doses of oral activated charcoal in various types of chronic intoxications.

References

- Abdallah AH, Tye A. A comparison of efficacy of emetic drugs and stomach lavage. American Journal of Diseases of Children 113: 571-575, 1967
- Adler LJ, Waters DH, Gwilt PR. The effect of activated charcoal on mouse sleep times induced by intravenously administered hypnotics. Biopharmaceutics and Drug Disposition 7: 421-429, 1986
- Alvan G. Effect of activated charcoal on plasma levels of nortriptyline after single doses in man. European Journal of Clinical Pharmacology 5: 236-238, 1973
- Andersen AH. Experimental studies on the pharmacology of activated charcoal, II: the effect of pH on the adsorption by charcoal from aqueous solutions. Acta Pharmacologica 3: 199-218, 1947
- Andersen AH. Experimental studies on the pharmacology of activated charcoal. III: Adsorption from gastro-intestinal contents. Acta Pharmacologica 4: 275-284, 1948
- Anderson IM, Ware C. Syrup of ipecacuanha. British Medical Journal 294: 578, 1987
- Arimori K, Mishima M, Iwaoku R, Nakano M. Evaluation of orally administered activated charcoal on intestinal dialysis of intravenously administered M79175, an aldose reductase inhibitor, in rats. Journal of Pharmacobio-Dynamics 10: 243-249, 1987
- Arnold FJ, Hodges JB, Barta RA, Spector S, Sunshine I, et al. Evaluation of the efficacy of lavage and induced emesis in treatment of salicylate poisoning. Pediatrics 23: 286-301, 1959
- Bainbridge CA, Kelly EL, Walking WD. In vitro adsorption of acetaminophen onto activated charcoal. Journal of Pharmaceutical Sciences 4: 480-483, 1977
- Belz GB, Bader H. Effect of oral charcoal on plasma levels of intravenous methyl proscillaridin. Klinische Wochenschrift 52: 1134-1135, 1974
- Berg MJ, Berlinger WG, Goldberg MJ, Spector R, Johnson GF. Acceleration of the body clearance of phenobarbital by oral

activated charcoal. New England Journal of Medicine 307: 642-644, 1982

- Berg MJ, Rose JQ, Wurster DE, Rahman S, Fincham RW, et al. Effect of charcoal and sorbitol-charcoal suspension on the elimination of intravenous phenobarbital. Therapeutic Drug Monitoring 9: 41-47, 1987
- Berlinger WG, Spector R, Goldberg MJ, Johnson GF, Quee CK, et al. Enhancement of theophylline clearance by oral activated charcoal. Clinical Pharmacology and Therapeutics 33: 351-354, 1983
- Boehnert MT, Lewander W, Gaudreault P, Lovejoy FH. Advances in clinical toxicology. Pediatric Clinics of North America 32: 193-211, 1985
- Boldy D, Vale JA. Treatment of salicylic poisoning with repeated oral charcoal. British Medical Journal 291: 1472, 1986
- Boldy DA, Heath A, Ruddock S, Vale JA, Prescott LF. Activated charcoal for carbamazepine poisoning. Lancet 1: 1027, 1987
- Boldy DAR, Smart V, Vale JA. Multiple doses of charcoal in digoxin poisoning. Lancet 2: 1076-1077, 1985
- Boldy DAR, Vale JA, Prescott LF. Treatment of phenobarbitone poisoning with repeated oral administration of activated charcoal. Quarterly Journal of Medicine 235: 997-1002, 1986
- Boxer L, Anderson FP, Rowe SS. Comparison of ipecac-induced emesis with gastric lavage in the treatment of acute salicylate ingestion. Pediatric Pharmacology and Therapeutics 5: 800-803, 1969
- Boyd JR. Drug Facts and Comparisons, p. 1839, Lippincott, Philadelphia-Toronto, 1982
- Burke M. Gastric lavage and emesis in the treatment of ingested poisons: a review and clinical study of lavage in ten adults. Resuscitation 1: 91-105, 1972
- Chaput de Saintonge DM, Herzheimer A. Activated charcoal impairs propantheline absorption. European Journal of Clinical Pharmacology 4: 52-53, 1971
- Chernish SM, Wolen RL, Rodda BE. Adsorption of propoxyphene hydrochloride by activated charcoal. Clinical Toxicology 5: 317-329, 1972
- Chin L, Picchioni AL, Bourn WM, Laird HE. Optimal antidotal dose of activated charcoal. Toxicology and Applied Pharmacology 26: 103-108, 1972
- Chin L, Picchioni AL, Duplisse BR. Comparative antidotal effectiveness of activated charcoal, Arizona montmorillonite, and evaporated milk. Journal of Pharmaceutical Sciences 58: 1353-1358, 1969
- Chin L, Picchioni AL, Duplisse BR. The action of activated charcoal on poisons in the digestive tract. Toxicology and Applied Pharmacology 16: 786-799, 1970
- Chin L, Picchioni AL, Gillespie T. Saline cathartics plus activated charcoal as antidotal treatments. Ciinical Toxicology 18: 865-871, 1981
- Chung DC, Murphy JE, Taylor TW. In vivo comparison of the adsorption capacity of 'superactive' charcoal and fructose with activated charcoal and fructose. Journal of Toxicology – Clinical Toxicology 19: 219-224, 1982
- Cooney D. A superactive charcoal for antidotal use in poisonings. Journal of Clinical Toxicology 11 (4): 387-390, 1977a
- Cooney DO. Activated charcoal: antidotal and other medical uses, Marcel Dekker Inc., New York 1980
- Cooney DO. In vitro evidence for ipecac inactivation by activated charcoal. Journal of Pharmaceutical Sciences 67: 426-430, 1978
- Cooney DO. The treatment of ethylene glycol poisoning with activated charcoal. IRCS Medical Sciences 5: 265, 1977b
- Cooney DO, Kane RP. Superactive charcoal adsorbs as fast as standard antidotal charcoal. Journal of Toxicology 16: 123-125, 1980
- Cooney DO, Roach M. Sucrose as a sweetener for activated charcoal. American Journal of Hospital Pharmacy 36: 797-798, 1979
- Corby DG, Decker WJ. Antidote for propoxyphene. Journal of the American Medical Association 203: 1074, 1968

- Corby DG, Lisciandro RC, Lehman RH, Decker WJ. The efficacy of methods used to evacuate the stomach after acute ingestions. Pediatrics 5: 871-874, 1967
- Cordonnier J, Van den Heede M, Bruynocghe P, Heyndrickx A. Adsorption of tilidine-HCL by activated charcoal: an *in vitro* and *in vivo* study. 3rd World Congress of the World Federation of Associations of Clinical Toxicology and Poison Control Centres, Brussels, 27-30 Aug, p.6, 1986
- Crome P, Dawling S, Braithwaite RA, Masters J, Walkey R. Effect of activated charcoal on absorption of nortriptyline. Lancet 2: 1203-1205, 1977
- Cupit GC, Temple AR. Gastrointestinal decontamination in the management of the poisoned patients. Emergency Medicine Clinics of North America 2: 15-29, 1984
- Curtis RA, Barone J, Giacona N. Efficacy of ipecac and activated charcoal/cathartic in prevention of salicylate absorption in a simulated overdose. Archives of Internal Medicine 144: 48-52, 1984
- Daunderer M. Medizinalkohle das älteste und wichtigste Gegengift. Fortschritte der Medizin 101: 697-700, 1983
- Dawling S, Crome P, Braithwaite R. Effect of delayed administration of activated charcoal on nortriptyline absorption. European Journal of Clinical Pharmacology 14: 445-447, 1978
- Decker WJ, Combs HF, Corby DG. Adsorption of drug and poison by activated charcoal. Toxicology and Applied Pharmacology 13: 454-469, 1968
- Decker WJ, Corby DG. Activated charcoal adsorbs aflatoxin B₁. Veterinary and Human Toxicology 22: 388-389, 1980
- De Neve R. Antidotal efficacy of activated charcoal in presence of jam, starch and milk. American Journal of Hospital Pharmacy 33: 965-966, 1976
- Derlet RW, Albertson TE. Activated charcoal past, present and future. Western Journal of Medicine 145: 493-496, 1986
- Dordoni P, Willson RA, Thompson RPH, Williams R. Reduction of absorption of paracetamol by activated charcoal and cholestyramine: a possible therapeutic measure. British Medical Journal 3: 86-87, 1973
- Dujovne CA, Pitterman A, Vincek WC, Doprinska MR, Davies RO, et al. Enterohepatic circulation of sulindac and metabolites. Clinical Pharmacology and Therapeutics 32: 172-177, 1983.
- Du Souich P, Caillé P, Larochelle P. Reduction of nadolol plasma half-life by activated charcoal and antibiotics in man. Clinical Pharmacology and Therapeutics 31: 222, 1982
- Easom JM, Caraccio TR, Lovejoy Jr FH. Evaluation of activated charcoal and magnesium citrate in the prevention of aspirin absorption in humans. Clinical Pharmacy 1: 154-156, 1982
- El-Bahie N, Allen EM, Williams J, Routledge PA. The effect of activated charcoal and hyoscine butylbromide alone and in combination on the absorption of mefenamic acid. British Journal of Clinical Pharmacology 19: 836-838, 1985
- Elonen E, Neuvonen PJ, Halmekoski J, Mattila MJ. Acute dapsone intoxication: a case with prolonged symptoms. Clinical Toxicology 14: 79-85, 1979
- Fiser RH, Maetz HM, Treuting JJ, Decker WJ. Activated charcoal in barbiturate and glutethimide poisoning of the dog. Pediatric Pharmacology and Therapeutics 6: 1045-1047, 1971
- Freedman GE, Pasternak S, Frenzelok EP. A clinical trial using syrup of ipecac and activated charcoal concurrently. Annals of Emergency Medicine 16: 164-166, 1987
 Gadgil SD, Damile SR, Advani SH, Vaidya AG. Effect of acti-
- Gadgil SD, Damile SR, Advani SH, Vaidya AG. Effect of activated charcoal on the pharmacokinetics of high-dose methotrexate. Cancer Treatment Reports 66: 1169-1171, 1982
- Gal P, Miller A, McCur JD. Oral activated charcoal to enhance theophylline elimination in an acute overdose. Journal of the American Medical Association 251: 3130-3131, 1984
- Galey FD, Lambert RJ, Busse M, Buck WB. Therapeutic efficacy of superactive charcoal in rats exposed to oral lethal doses of T-2 toxin. Toxicon 25: 493-499, 1987

- Galinsky RE, Levy G. Evaluation of activated charcoal-sodium sulfate combination for inhibition of acetaminophen absorption and repletion of inorganic sulfate. Clinical Toxicology 22: 21-30, 1984
- Gaudreault P, Friedman PA, Lovejoy FH. Efficacy of activated charcoal and magnesium citrate in the treatment of oral paraquat intoxication. Annals of Emergency Medicine 14: 123-125, 1985
- Glazko AJ. Pharmacology of the fenemates, III: metabolic disposition. Annals of Physiological Medicine 9: 23-26, 1967
- Goldberg MJ, Berlinger WG. Treatment of phenobarbital overdose with activated charcoal. Journal of the American Medical Association 247: 2400-2401, 1982
- Goldberg MJ, Berlinger WG, Park GD. Activated charcoal in phenobarbital overdose. Journal of the American Medical Association 253: 1120-1121, 1985a
- Goldberg MJ, Park GD, Spector R, Fischer LJ, Feldman RD. Lack of effect of oral activated charcoal on imipramine clearance. Clinical Pharmacology and Therapeutics 38: 350-353, 1985b
- Goldberg MJ, Spector R, Park GD, Johnson GF, Roberts P. The effect of sorbitol and activated charcoal on serum theophylline concentrations after slow-release theophylline. Clinical Pharmacology and Therapeutics 41: 108-111, 1987
 Guay DRP, Meatherall RC, Macaulay PA, Yeung C. Activated
- Guay DRP, Meatherall RC, Macaulay PA, Yeung C. Activated charcoal adsorption of diphenylhydramine. International Journal of Clinical Pharmacology, Therapy and Toxicology 22: 395-400, 1984
- Gwilt PR, Perrier D. Influence of 'thickening' agents on the antidotal efficacy of activated charcoal. Clinical Toxicology 9: 89-92, 1976
- Harsch HH. Aspiration of activated charcoal. New England Journal of Medicine 314: 318, 1986
- Hassan E. Treatment of meprobamate overdose with repeated oral doses of activated charcoal. Annals of Emergency Medicine 15: 73-76, 1986
- Hauge SM, Willamann JJ. Effect of pH on adsorption by carbons. Ind Eng Chem 19: 943-953, 1927
- Hayden JW, Comstock EG. Use of activated charcoal in acute poisoning. Clinical Toxicology 8: 515-533, 1975
- Heath A, Van Loo T. Multiple dose oral activated charcoal therapy in carbamazepine overdose. 3rd World Congress of the World Federation of Associations of Clinical Toxicology and Poison Control Centres, Brussels 27-30 August, 1986, Abstract No. 054, 1986
- Heimer GM, Englund DE. Enterohepatic recirculation of oestriol: inhibition by activated charcoal. Acta Endocrinologica 113: 93-95, 1986
- Hillman RJ, Prescott LF. Treatment of salicylate poisoning with repeated oral charcoal. British Medical Journal 291: 1472, 1985
- Honcharic N, Anthone S. Activated charcoal in acute cyclosporin overdose. Lancet 1: 1051, 1985
- Hulten BA, Heath A, Mellstrand T, Hedner T. Does alcohol adsorb to activated charcoal? Human Toxicology 5: 211-212, 1986
- Jacobsen D, McMartin KE. Methanol and ethylene glycol poisonings: mechanism of toxicity, clinical course, diagnosis and treatment. Medical Toxicology 1: 309-334, 1986 Justiniani FR, Hippalegaonkar R, Martinez LO. Charcoal-con-
- Justiniani FR, Hippalegaonkar R, Martinez LO. Charcoal-containing empyema complicating treatment for overdose. Chest 87: 404-405, 1985
- Kannisto H, Neuvonen PJ. Adsorption of sulfonylureas onto activated charcoal in vitro. Journal of Pharmaceutical Sciences 73: 253-256, 1984
- Kärkkäinen S, Neuvonen PJ. Effect of oral charcoal and urine pH on dextropropoxyphene pharmacokinetics. International Jounal of Pharmacology, Therapy and Toxicology 23 (4): 219-225, 1985
- Kärkkäinen S, Neuvonen PJ. Effect of oral charcoal and urine pH on sotalol pharmacokinetics. International Journal of Clinical Pharmacology, Therapy and Toxicology 22 (8): 441-446, 1984
- Kärkkäinen S, Neuvonen PJ. Pharmacokinetics of amitriptyline influenced by oral charcoal and urine pH. International Jour-

nal of Clinical Pharmacology, Therapy and Toxicology 24: 326-332, 1986

- Klein-Schwarz W, Oderda G. Absorption of oral antidotes for acetaminophen poisoning (methionine and N-acetylcysteine) by activated charcoal. Paper presented to 1980 National Poison Center Network Annual Symposium, Pittsburgh, Pennsylvania, June 10, 1980
- Korttila K, Mattila MJ, Linnoila M. Prolonged recovery after diazepam sedation: the influence of food, charcoal ingestion and injection rate on the effects of intravenous diazepam. British Journal of Anaesthesia 48: 333-340, 1976
- Kotaki H, Yamamura Y, Tanimura Y, Saitoh Y, Fujio N, et al. Enterohepatic circulation of clioquinol in the rat. Journal of Pharmacobio-Dynamics 7: 420-425, 1984
- Krenzelok EP. Gastrointestinal transit times of cathartics used with activated charcoal. Clinical Pharmacology 4: 446-448, 1985
- Krenzelok EP, Freedman GE, Pasternak S. Preserving the emetic effect of syrup of ipecac with concurrent activated charcoal administration: a preliminary study. Clinical Toxicology 24: 159-166, 1986
- Kulig K, Bar-Or D, Cantrill SV, et al. Management of acutely poisoned patients without gastric emptying. Annals of Emergency Medicine 14: 562-567, 1985
- Laass W. Therapy of acute oral poisonings by organic solvents: treatment by activated charcoal in combination with laxatives. Archives of Toxicology 4 (Suppl.): 406-409, 1980
- Lalonde RS, Deshpande R, Hamilton PP, McLean WM, Greenway DC. Acceleration of digoxin clearance by activated charcoal. Clinical Pharmacology and Therapeutics 37: 367-371, 1985
- Laufen H, Leifold M, Riedel KD. The effect of activated charcoal on the gastrointestinal absorption and reabsorption of piroxicam in dog and man. In Aiache & Hirtz (Eds) Biopharmaceutics and pharmacokinetics, 2nd European Congress, Spain, April 24-27, pp. 401-403, 1984
- Levy G. Gastrointestinal clearance of drugs with activated charcoal. New England Journal of Medicine 307: 676-678, 1982
- Levy G, Houston JB. Effect of activated charcoal on acetaminophen absorption. Pediatrics 3: 432-435, 1976
- Levy G, Tsuchiya T. Effect of activated charcoal on aspirin absorption in man. Clinical Pharmacology and Therapeutics 13: 317-322, 1972
- Lim DT, Singh P, Nourtsis S, Crutz RD. Absorption inhibition and enhancement of elimination of sustained-releasee theophylline tablets by oral activated charcoal. Annals of Emergency Medicine 15: 1303-1307, 1986
- Linden CH, Lewis PK, Rumack BH. Phenobarbital overdosage: treatment with multiple doses activated charcoal. Abstract No. A-7. Abstracts of the Annual Scientific Meeting of the American Academy of Clinical Toxicologists, the American Association of Poison Control Centers, and the American Board of Medical Toxicology, Boston, Aug 6-11, 1983
- Linden CH, Rumack BH. Enhanced elimination of meprobamate by multiple doses of activated charcoal. Veterinary and Human Toxicology 26 (Suppl. 2): 47, 1984
- Mathur LK, Jaffe JM, Colaizzi JL. Effect of carboxymethylcellulose on the adsorptive capacity of charcoal. American Journal of Hospital Pharmacy 33: 1122, 1976a
- Mathur LK, Jaffe JM, Colaizzi JL, Moriarty RW. Activated charcoal-carboxymethylcellulose gel formulation as an antidotal agent for orally ingested aspirin. American Journal of Hospital Pharmacy 33: 717-719, 1976b
- Mayersohn M, Perrier D, Picchioni AL. Evaluation of a charcoalsorbitol mixture as an antidote for oral aspirin overdose. Clinical Toxicology 11: 561-567, 1977
- McKinnon RS, Desmond PV, Harman PJ, Kamm M, Ghabrial H, et al. Studies on the mechanisms of action of activated charcoal on theophylline pharmacokinetics. Journal of Pharmacy and Pharmacology 39: 522-525, 1987

Minocha A, Herold DA, Barth JT, Gideon DA, Spyker DA. Ac-

tivated charcoal in oral ethanol absorption: lack of effect in humans. Clinical Toxicology 24: 225-234, 1986

- Minocha A, Krenzelok EP, Spyker DA. Dosage recommendations for activated charcoal-sorbitol treatment. Journal of Toxicology – Clinical Toxicology 23: 579-587, 1985
- Nau CA, Neal J, Stembridge V. A study of the physiological effects of carbon black. I: Ingestion. Archives of Industrial Health 17: 21-28, 1985a
- Nau CA, Neal J, Stembridge V. A study of the physiological effects of carbon black. II: Skin contact. Archives of Industrial Health 18: 511-520, 1985b
- Nau CA, Neal J, Stembridge V, Cooley RN. Physiological effects of carbon black. IV: Inhalation. Archives of Environmental Health 4: 415-431, 1962
- Neuvonen PJ. Clinical pharmacokinetics of oral activated charcoal in acute intoxications. Clinical Pharmacokinetics 7: 465-489, 1982
- Neuvonen PJ, Elfving SM, Elonen E. Reduction of absorption of digoxin, phenytoin and aspirin by activated charcoal in man. European Journal of Clinical Pharmacology 13: 213-218, 1978
- Neuvonen PJ, Elonen E. Effect of activated charcoal on absorption and elimination of phenobarbitone, carbamazepine and phenylbutazone in man. European Journal of Clinical Pharmacology 17: 51-57, 1980
- Neuvonen PJ, Elonen E, Haapanen EJ. Acute dapsone intoxication: clinical findings and effect of oral charcoal and hemodialysis on dapsone elimination. Acta Medica Scandinavica 214: 215-220, 1983a
- Neuvonen PJ, Elonen E, Mattila MJ. Oral activated charcoal and dapsone elimination. Clinical Pharmacology and Therapeutics 6: 823-827, 1980
- Neuvonen PJ, Kannisto H, Hirvisalo El. Effect of activated charcoal on absorption of tolbutamide and valproate in man. European Journal of Clinical Pharmacology 24: 243-246, 1983b
- Neuovnen PJ, Kannisto H, Lankinen S. Capacity of two forms of activated charcoal to adsorb nefopam *in vitro* and to reduce its toxicity *in vivo*. Clinical Toxicology 21: 333-342, 1983d
- Neuvonen PJ, Kärkkäinen S. Effects of charcoal, sodium bicarbonate and ammonium chloride on chlorpropamide kinetics. Clinical Pharmacology and Therapeutics 33: 386-393, 1983
- Neuvonen PJ, Kivistö K, Hirvisalo EL. Effect of resins and charcoal on digoxin, carbamazepine and furosemide absorption. Abstract 0.339, Xth International Congress of Pharmacology, Sydney, Australia, 1987
- Neuvonen PJ, Olkkola KT. Activated charcoal and syrup of ipecac in prevention of cimetidine and pindolol absorption in man after administration of metoclopramide as antiemetic agent. Clinical Toxicology 22: 103-114, 1984b
- Neuvonen PJ, Olkkola KT. Effect of dose of charcoal on the absorption of disopyramide, indomethacin and trimethoprim by man. European Journal of Clinical Pharmacology 26: 761-767, 1984a
- Neuvonen PJ, Olkkola KT. Effect of purgatives on antidotal efficacy of oral activated charcoal. Human Toxicology 5: 255-263, 1986
- Neuvonen PJ, Olkkola KT, Alanen T. Effect of ethanol and pH on the adsorption of drugs to activated charcoal: studies *in vitro* and in man. Acta Pharmacologica et Toxicologica 54: 1-7, 1984
- Neuvonen PJ, Vartiainen M, Tokola O. Comparison of activated charcoal and ipecac syrup in prevention of drug absorption. European Journal of Clinical Pharmacology 24: 557-562, 1983c
- Nimmo WS. Gastric emptying and drug absorption. In Prescott & Nimmo (Eds) Drug absorption, pp. 11-20, ADIS Press, Hong Kong, 1979
- Nitsch J, Köhler U, Luderitz B. Hemmung der Flecainidresorption durch Aktivkohle. Zeitschrift fur Kardiologie 76: 289-291, 1987
- North DS, Peterson RG, Krenzelok EP. Effect of activated char-

coal administration on acetylcysteine serum levels in humans. American Journal of Hospital Pharmacy 38: 1022-1024, 1981a

- North DS, Thompson JD, Peterson CD. Effect of activated charcoal on ethanol blood levels in dogs. American Journal of Hospital Pharmacy 38: 864-866, 1981b
- Oderda GM. Clinical Toxicology. In Hofindal & Hirschman (Eds) Clinical pharmacy and therapeutics, 2nd ed. pp. 1-21, Williams & Wilkins, London, 1979
- Okonek S, Setyadharma H, Borchert A, Krienke EG. Activated charcoal is as effective as Fuller's earth or bentonite in paraquat poisoning. Klinische Wochenschrift 60: 207-210, 1982
- Olkkola KT. Effect of charcoal-drug ratio on antidotal efficacy of oral activated charcoal in man. British Journal of Clinical Pharmacology 19: 767-773, 1985b
- Olkkola KT. Factors affecting the antidotal efficacy of oral activated charcoal. Academic dissertation, Helsinki, 1985a
- Olkkola KT, Neuvonen PJ. Do gastric contents modify antidotal efficacy of oral activated charcoal? British Journal of Clinical Pharmacology 18: 663-669, 1984b
- Olkkola KT, Neuvonen PJ. Effect of gastric pH on antidotal efficacy of activated charcoal in man. International Journal of Clinical Pharmacology, Therapy and Toxicology 22: 565-569, 1984a
- Park GD. Radomski L, Goldberg MJ, Spector R, Johnson GF, et al. Effects of size and frequency of oral doses of charcoal on theophylline clearance. Clinical Pharmacology and Therapeutics 34: 663-666, 1983
- Park GD, Spector R, Goldberg MJ, Johnson GF. Expanded role of charcoal therapy in the poisoned and overdosed patient. Archives of Internal Medicine 146: 969-973, 1986
- Park GD, Spector R, Goldberg MJ, Johnson GF, Feldman R, et al. Effect of the surface area of activated charcoal on theophylline clearance. Journal of Clinical Pharmacology 24: 289-292, 1984
- Picchioni AL. Activated charcoal, a neglected antidote. Pediatric Clinics of North America 17: 535-543, 1970
- Picchioni AL, Chin L, Gillespie T. Evaluation of activated charcoal sorbitol suspension as an antidote. Clinical Toxicology 19: 433-444, 1982
- Picchioni AL, Chin L, Verhulst HL, Dieterle B. Activated charcoal vs. 'universal antidote' as an antidote for poisons. Toxicology and Applied Pharmacology 8: 447-454, 1966
- Pollack MM, Dunbar BS, Holbrook PR, Fields AI. Aspiration of activated charcoal and gastric contents. Annals of Emergency Medicine 10: 528-529, 1981
- Pond S, Jacobs M, Marks J, Garner J, Goldschlager N, et al. Treatment of digitoxin overdose with oral activated charcoal. Lancet 2: 1177-1178, 1981
- Pond SM. Role of repeated oral doses of activated charcoal in clinical toxicology. Medical Toxicology 1: 3-11, 1986
- Pond SM, Olson KR, Osterloh JD, Tong TG. Randomised study of the treatment of phenobarbital overdose with repeated doses of activated charcoal. Journal of the American Medical Association 251: 3104-3108, 1984
- Prescott LF, Boye GL, Simpson D. Rapid drug removal after overdosage by gastrointestinal dialysis with activated charcoal. 3rd World Conference on Clinical Pharmacology and Therapeutics, Stockholm, Jul 27-Aug 1, 1986, Abstracts II, p. 270, No. 1431, 1986
- Radomski L, Park GD, Goldberg MJ, Spector R, Johnson GF, et al. Model for theophylline overdose treatment with oral activated charcoal. Clinical Pharmacology and Therapeutics 35: 402-408, 1984
- Reigart JR, Trammel Jr HL, Lindsey JM. Repetitive doses of activated charcoal in dapsone poisoning in a child. Journal of Toxicology – Clinical Toxicology 19: 1061-1066, 1982
- Rosenberg J, Benowitz L, Pound SM. Pharmacokinetics of drug overdose. Clinical Pharmacokinetics 6: 161-192, 1981

- Rumack BH (Ed.) Poisindex, Micromedex Incorporation, Englewood, 1980
- Rybolt TR, Burrell DE, Shults JM, Kelley AK. In vitro coadsorption of acetaminophen and N-acetylcysteine onto activated carbon powder. Journal of Pharmaceutical Sciences 75: 904-906, 1986
- Rygnestad T, Walstad RA, Dahl K. Self-poisoning with theophylline: the effect of repeated doses of oral charcoal on drug elimination. Acta Medica Scandinavica 219: 425-427, 1986
- Scheinin M, Virtanen R, Iisalo E. Effect of single and repeated doses of activated charcoal on the pharmacokinetics of doxepin. International Journal of Clinical Pharmacology, Therapy and Toxicology 23: 38-42, 1985
- Scholtz EC, Jaffe JM, Colaizzi JL. Evaluation of five activated charcoal formulations for inhibition of aspirin absorption and palatability in man. American Journal of Hospital Pharmacy 35: 1355-1359, 1978
- Scolding N, Ward MJ, Hutchings A, Routledge PA. Charcoal and isoniazid pharmacokinetics. Human Toxicology 5: 285-286, 1986
- Shannon M, Fish SS, Lovejoy Jr H. Cathartics and laxatives: do they still have a place in management of the poisoned patient? Medical Toxicology 1: 247-252, 1986
- Sintek C, Hendeles L, Weinberger M. Inhibition of theophylline absorption by activated charcoal. Journal of Pediatrics 94: 314-316, 1979
- Sketris IS, Mowry JB, Czajka PA, Anderson WH, Stafford DT. Saline catharsis: effect on aspirin bioavailability in combination with activated charcoal. Journal of Clinical Pharmacology 22: 59-64, 1982
- Smith RP, Gosselin RE, Henderson JA, Anderson DM. Comparison of the adsorptive properties of activated charcoal and Alaskan montmorillonate for some common poisons. Toxicology and Applied Pharmacology 10: 95-104, 1967
- Sorby DL. Effect of adsorbents on drug absorption, I: modification of promazine absorption by activated attapulgite and activated charcoal. Journal of Pharmaceutical Sciences 5: 677-683, 1965
- Spector R, Goldberg MJ, Johnson GF. Expanded role of charcoal therapy in the poisoned and overdosed patient. Archives of Internal Medicine 146: 969-973, 1986
- Swartz CM, Sherman A. The treatment of tricyclic antidepressant overdose with repeated charcoal. Journal of Clinical Psychopharmacology 4: 336-340, 1984
- Szabuniewicz M, Bailey EM, Wiersig DO. A new regimen for the treatment of ethylene glycol poisoning. IRCS Medical Science 3: 102, 1975
- Tenenbein M, Cohen S, Sitar DA. Whole bowel irrigation as a decontamination procedure after acute drug overdose. 3rd World Congress of the World Federation of Associations of Clinical Toxicology and Poison Control Centres. Brussels 27-30 August, 1986, abstract 092, 1986
- Teschke R. Therapie akuter Vergiftungen durch halogenierte aliphatische Kohlenwasserstoffe. Deutsche Medizinische Wochenschrift 109: 543-546, 1984
- Traeger SM, Haug MT. Reduction of diazepam serum half life and reversal of coma by activated charcoal in a patient with severe liver disease. Clinical Toxicology 24: 329-337, 1986
- Tsuchiya T, Levy G. Relationship between effect of activated charcoal on drug absorption in man and its drug adsorption characteristics *in vitro*. Journal of Pharmaceutical Sciences 61: 586-589, 1972
- Vale JA, Ruddock FS, Boldy DAR. Multiple doses of activated charcoal in the treatment of phenobarbitone and carbamazepine poisoning. 3rd World Congress of the World Federation of Associations of Clinical Toxicology and Poison Control Centres. Brussels 27-30 August, 1986, abstract 071, 1986
- Van de Graaff W, Thompson WL, Sunshine I, Fretthold D, Leickly F, et al. Adsorbent and cathartic inhibition of enteral drug ab-

sorption. Journal of Pharmacology and Experimental Therapeutics 221: 656-663, 1982

- Venho VMK, Salonen RO, Mattila MJ. Modification of the pharmacokinetics of doxycycline in man by ferrous sulphate or charcoal. European Journal of Clinical Pharmacology 14: 277-280, 1978
- Wheeler-Usher DH, Wanke LA, Bayer MJ. Gastric emptying: risk versus benefit in the treatment of acute poisoning. Medical Toxicology 1: 142-153, 1986
- Wogan J, Frommer D, Kulig K, Rumack B. Multiple dose activated charcoal for intravenous salicylate intoxication in a dog model. 3rd World Congress of the World Federation of Associations of Federation of Associations of Clinical Toxicology

and Poison Control Centers, Brussels 27-30 Aug, 1986, abstract 053, 1986

- Yancy RE, O'Barr TP, Corby DG. In vitro and in vivo evaluation of the effect of cherry flavouring on the adsorptive capacity of activated charcoal for salicylic acid. Veterinary and Human Toxicology 19: 163-165, 1977
- Yatzidis H. Activated charcoal rediscovered. British Medical Journal 7: 51, 1972

Authors' address: Dr *Pertti J. Neuvonen*, Department of Clinical Pharmacology, University of Helsinki and University Central Hospital, Paasikivenkatu 4, SF-00250 Helsinki (Finland).