

Microbiological Water Safety Detection



The purpose of this presentation is to convey the work done by the MSWM subteam this semester. More information can be found on our team page on the AguaClara wiki page



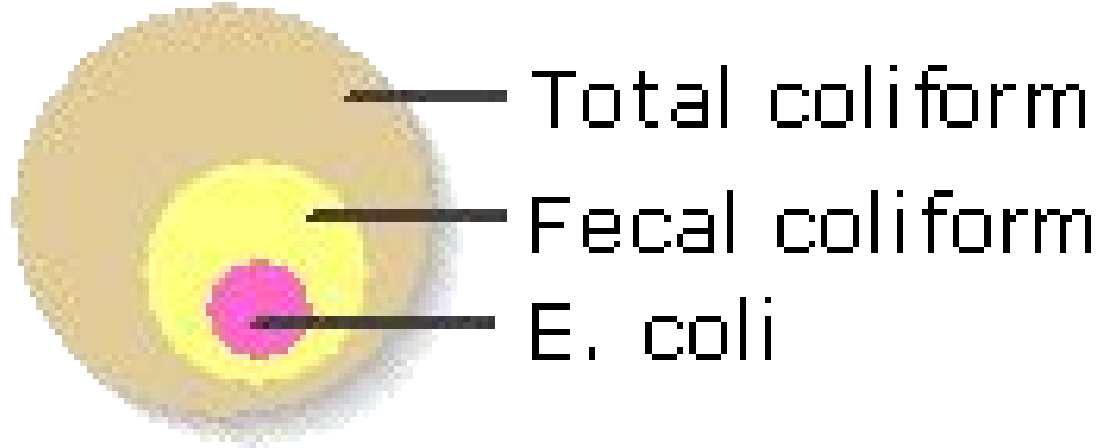
Problems

Causes of E.Coli and coliform growth:

- Leaks in pipes and tanks
- Improper cleaning of backwashing filters
- Increased precipitation

Unexpected events can take place during lifetime of plant operation

Coliform bacteria



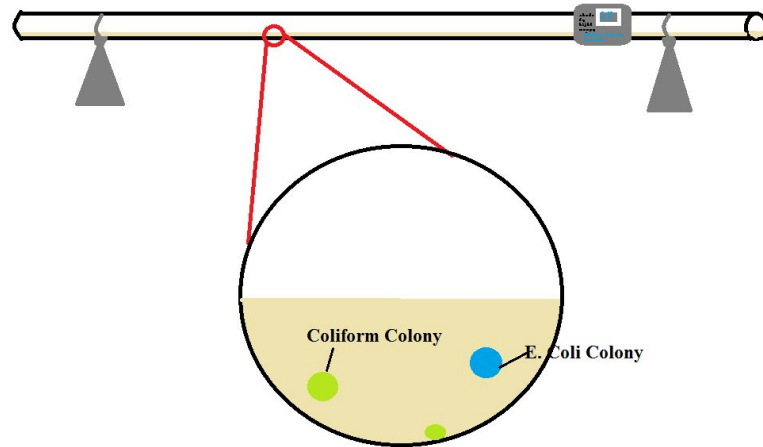
Goals

- Testing AguaClara water post-plant-treatment
- Creating a low cost test
- Creating a reliable test
- Creating a fast test
- Enumeration vs Presence/Absence

There are a plethora of pathogen detection methods, but few that can be used in low resource areas.



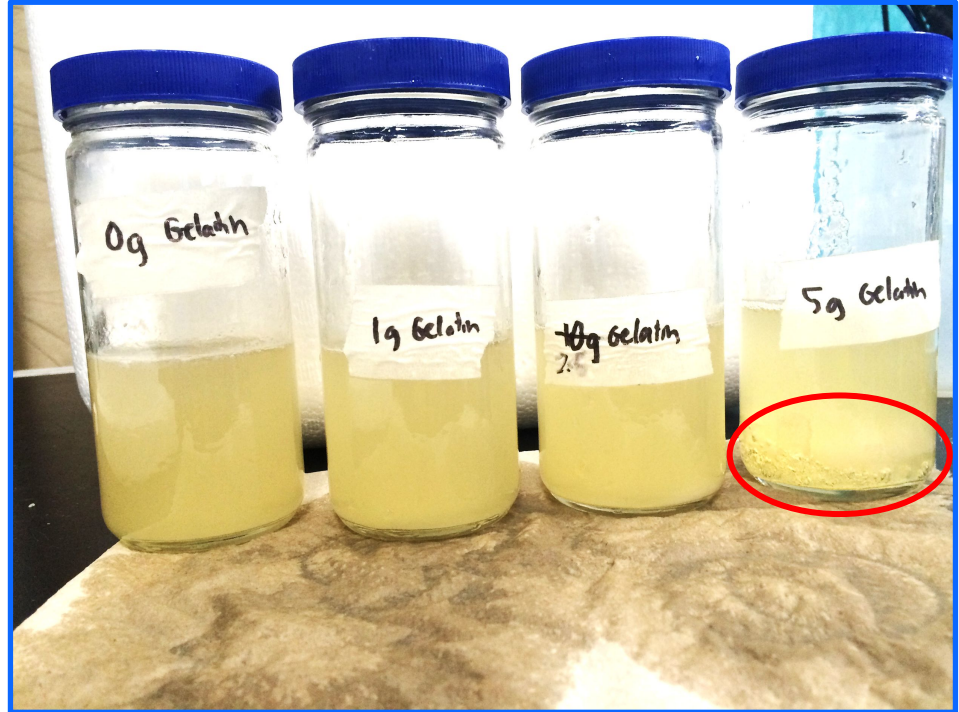
Sliding Colony Counter



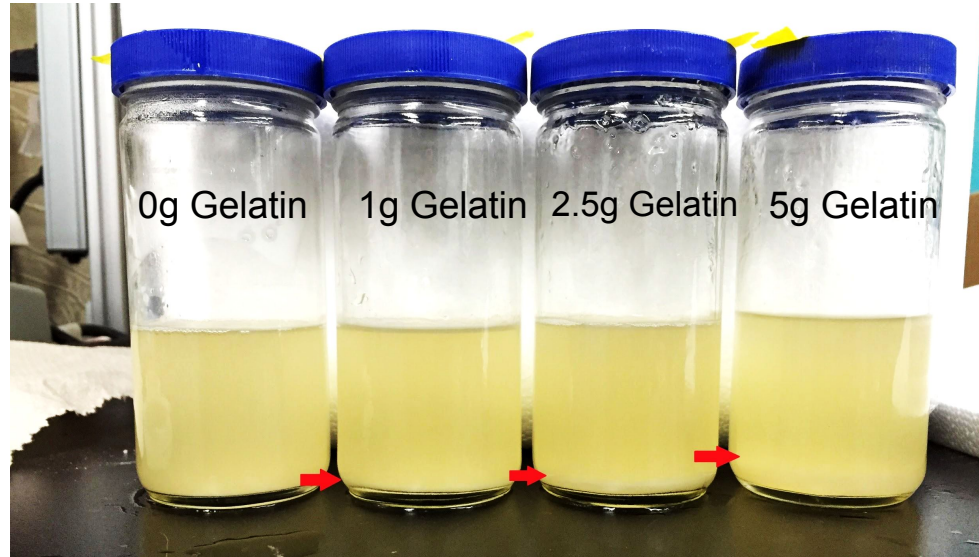
Design Goal: tube incubator with sliding colony counting device.

Methods

- Used water sample from creek
- Created LB Broth
- Heated up samples
- Mixed with gelatin
- Incubated



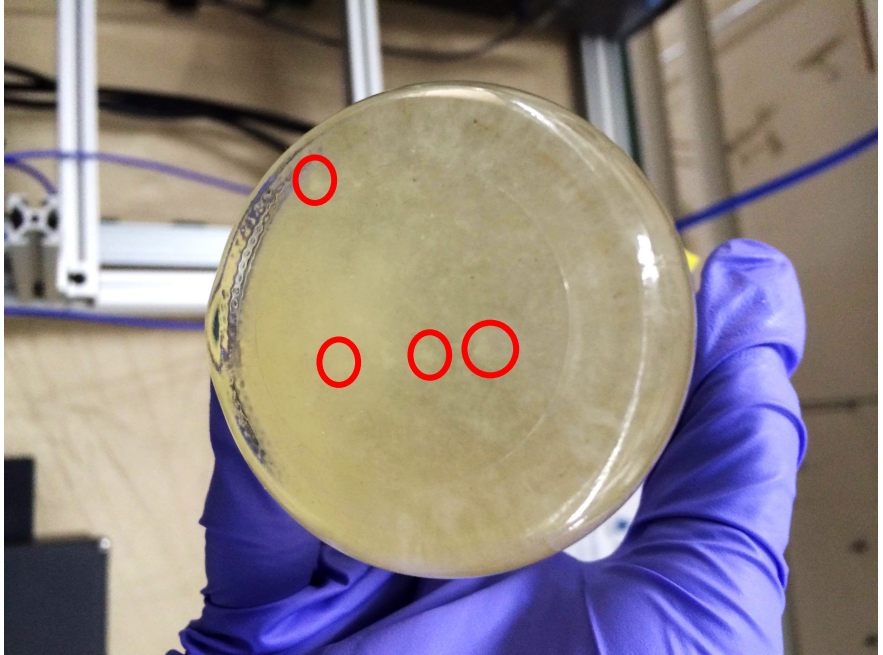
Difference in gelatin concentration correlated to difference in bacterial retention



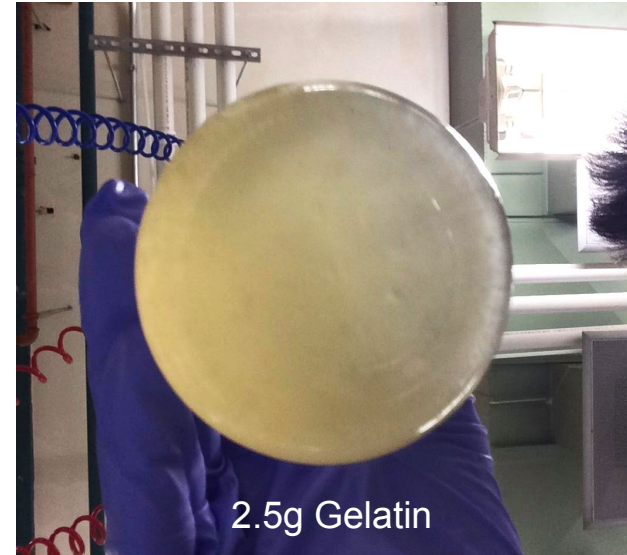
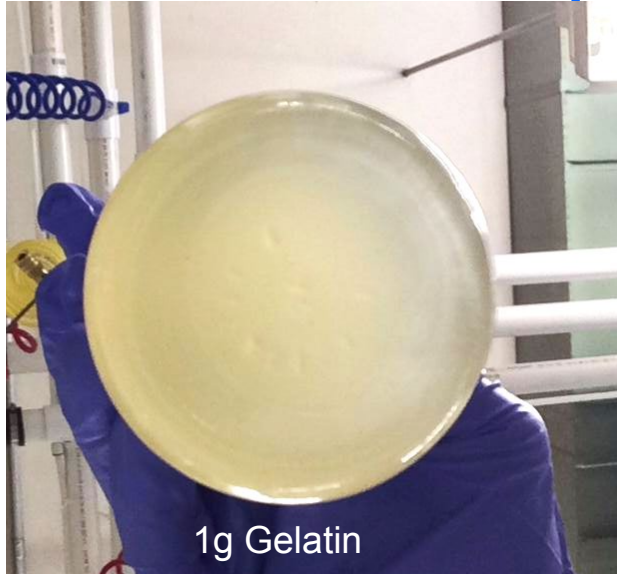
Greater gelatin concentrations tended to yield thicker layers on the bottom of the jars.

Isolation of colony forming units only occurred in 5g gelatin

- Control and 1g Gelatin were similar
- 2.5g gelatin: thicker layer but no isolation
- CFUs-colony forming units



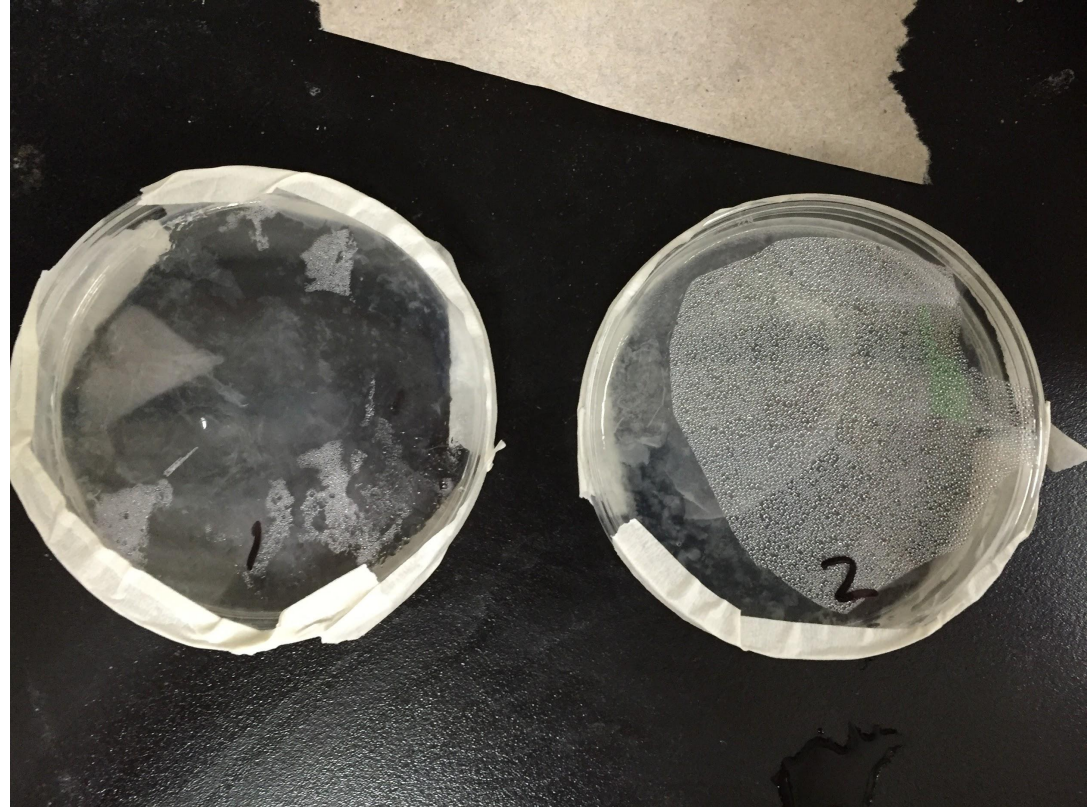
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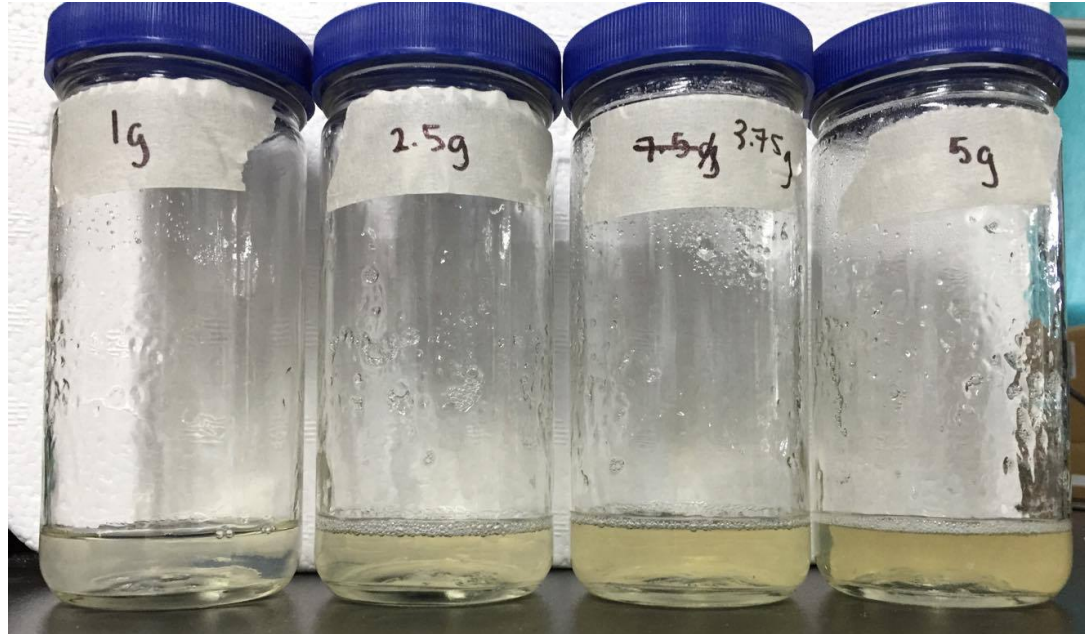
Compared to the 5g gelatin jars, the 1g and 2.5g gelatin samples only formed layers of bacteria rather than colonies.

Implementation of the pour plate method

- Thinner spread, more area
- Less sample water
- Greater ratio of gelatin to sample
- Bacterial films formed

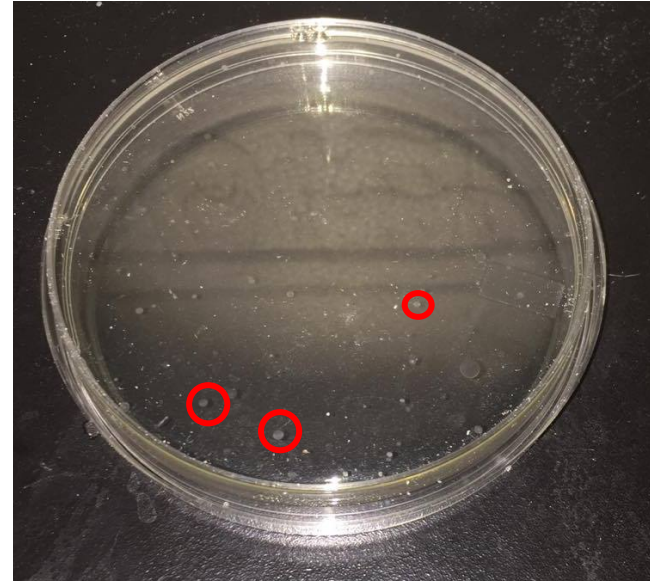
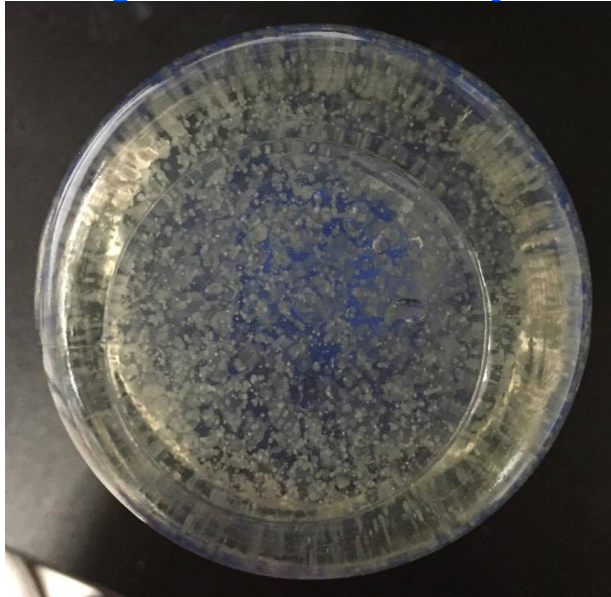


Exclusively gelatin experimentation



More gelatin → Darker media

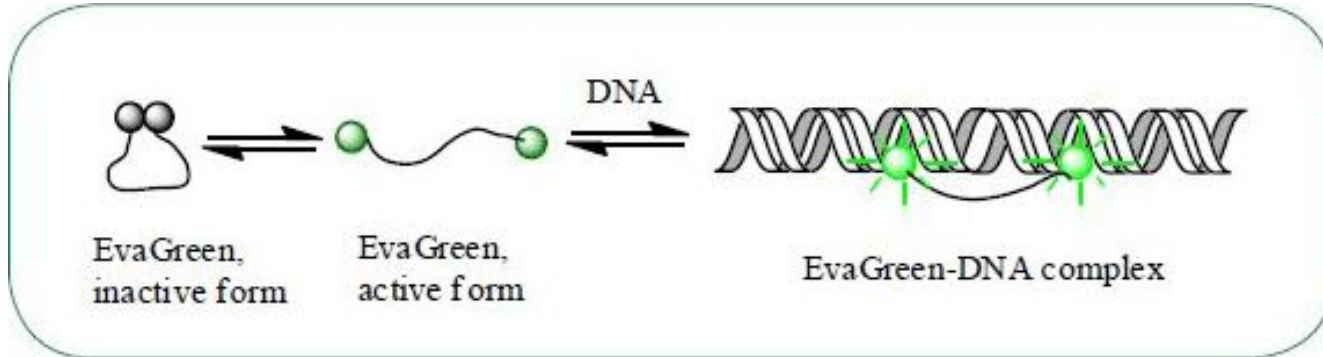
Combination of gelatin and media in both jars and plates



Used minimum required gelatin concentration found from previous experimentation with LB media instead of water

Future Work

- Color indication integration
- Investigate contamination
- Differing container shapes



An indicator will aid in identifying colony forming units

Questions and Recommendations

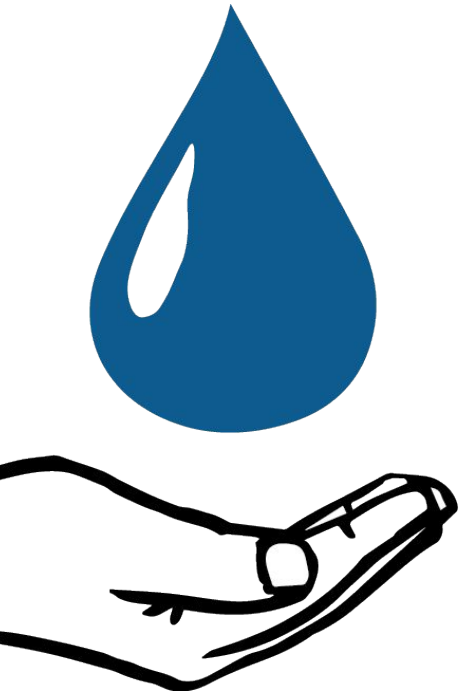


Jacqueline Dokko
Biological Engineering
jmd475@cornell.edu

Janak Shah
Chemical Engineering
jhs373@cornell.edu



Appendix Slides



Literature Review

A chart of all of the different types of detection.

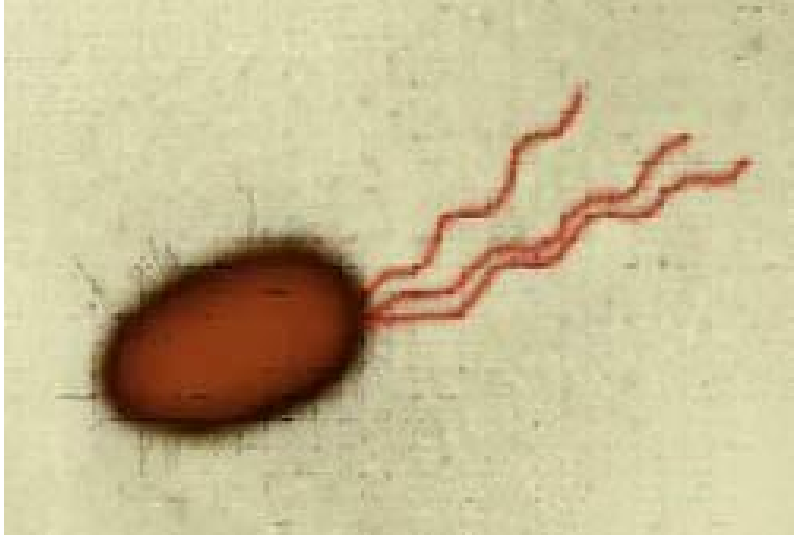
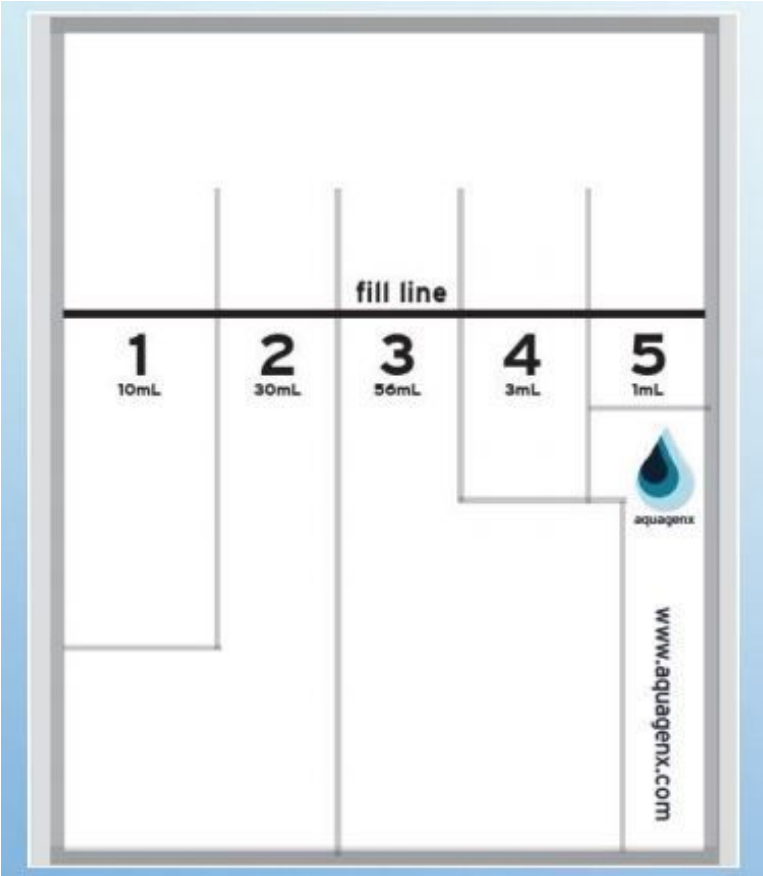
Table 4. Catalogue of microbial drinking water tests.

Type	Product	Resources required											Other			Settings		
		Cost per test	Cost of specialist equipment	Analysis time (min)	Trained technicians	Controlled incubation	Ultraviolet light	Sterilisation/disinfection	Deionised water	Cold storage	Transport	Disposal	Time to result (hrs)	Shelf life (months)	Temperature (°C)	Low resource	Medium resource	High resource
Hydrogen sulphide	Padlockgreen™	\$0.60	\$0	<5			x				S	24-72	12	RT				
	ITEX P ₆ 20 mL	\$0.80	\$0	<5			x				S	24-72	24	RT				
	ITEX™	\$2.40	\$100	<5			x				M	24-72	24	RT				
	ITEX M ₆ 100 mL	\$1.80	\$0	<5			x				M	24-72	12	RT				
	Local manufacture	Δ	\$0	<5			x				S	24-72	Δ	RT				
Presence Absence	Total Coliform	\$1.20	\$0	<5			x				S	44-48	24	RT				
	Coliform	\$0.80	\$100	<5		x	x			x	M	24	36	2-8				
	Coliform 10 mL	\$1.80	\$100	<5		x	x			x	S	24	12	4-80				
	Coliform 100 mL	\$8.00	\$100	<5		x	x			x	M	24	12	4-80				
	Coliform 8	\$8.00	\$100	<5		x	x			x	M	24	12	2-25				
	Coliform 18	\$8.00	\$100	<5		x	x			x	M	18	18	2-25				
	Indole test Coliform™	\$4.80	\$100	<5		x	x			x	M	16	22	4-80				
	E. coli and Total coliform	Watercheck™	\$5.00	\$2,700	<5		x	x			x	M	24	36	2-20			
	ReadyCount®	\$3.00	\$100	<5		x	x			x	M	24	36	18-25				
	FCCount	\$9.00	\$100	<5		x	x			x	M	20	12	RT				
	ECC Blue 100P	\$9.70	\$100	<5		x	x			x	M	24	12	RT				
	AquaCHECK™	\$2.40	\$0	<5		x	x			x	M	18	24	15-30				
	Hi-Selective™ E. coli	\$2.20	\$0	<5		x	x			x	M	24-48	12	2-8				
	Most Probable Number	Compartmentalized bag test	\$1.00	\$0	<5							S	24-72	6-9	RT			
		AquaTest™	\$4.00	\$100	5		x	x				M	24	24	RT			
Coligata™		\$7.50	\$200	10		x	x				L	24	36	2-20				
ECC Blue-Quant		\$5.80	\$100	5		x	x			x	L	24	12	RT				
Multiple tube (LTP-ECC-3,6,12)		\$9.50	\$200	30		x	x			x	S	48	36	RT				
Multiple tube (LTP-50,10)		\$2.10	\$200	30		x	x			x	S	36	36	RT				
Colitag-0.02%1400		\$5.77	\$0	10		x	x			x	L	16	22	4-30				
Colitag-Quant-Tray®		\$5.80	\$4,100	10		x	x			x	L	18,04	12	2-25				
Colitag-Quant-Tray® 2000		\$8.00	\$4,100	10		x	x			x	L	18,04	12	2-25				
Colony Count		Plate Methods	PlateCount™	\$1.30	\$100	<5		x			x	S	24	18	45			
	PlateCount™	\$0.70	\$100	<5		x				x	S	24	18	45				
	CFECOMag™	\$0.80	\$100	15		x				x	S	24	36	15-30				
	ECC	\$1.50	\$0	<5		x				x	S	24	24	1-30				
	Colitag Express	\$2.20	\$0	5		x				x	M	24	12	<5				
Gel based	ColiCol-PadlockGel	\$3.80	\$100	5		x	x			x	M	28	12	RT				
	Portable 140-58-5	\$0.80	\$2,700	20		x	x			x	S	24	48	RT				
Colony Count	Membrane Filtration	Portable 140-58-5	\$2.50	\$4,000	15		x			x	M	24	12	2-6				
	mem-Cellbag 24™	\$2.50	\$2,500	15		x	x			x	M	24	12	2-6				
	Colitag-302™	\$2.20	\$2,500	15		x	x			x	M	24	12	<5				
	mem-Bio	\$1.80	\$2,500	15		x	x			x	M	24	48	RT				
	mem-FC	\$1.80	\$2,500	15		x	x			x	M	24	48	RT				
	CFECOMag™	\$1.10	\$2,500	15		x	x			x	M	24	36	15-30				
	Liquid ECC	\$1.30	\$2,500	15		x	x			x	M	24	36	15-30				
	ECC	\$1.30	\$2,500	15		x	x			x	M	24	36	15-30				
	ME Agar	\$1.70	\$2,500	15		x	x			x	M	24	36	RT				
	Chlorococult	\$1.20	\$2,500	15		x	x			x	M	24	40	RT				
Papillat E. coli	7	\$2,500	15		x	x			x	M	24	7	RT					



The compartmentalized bag test.

Literature Review

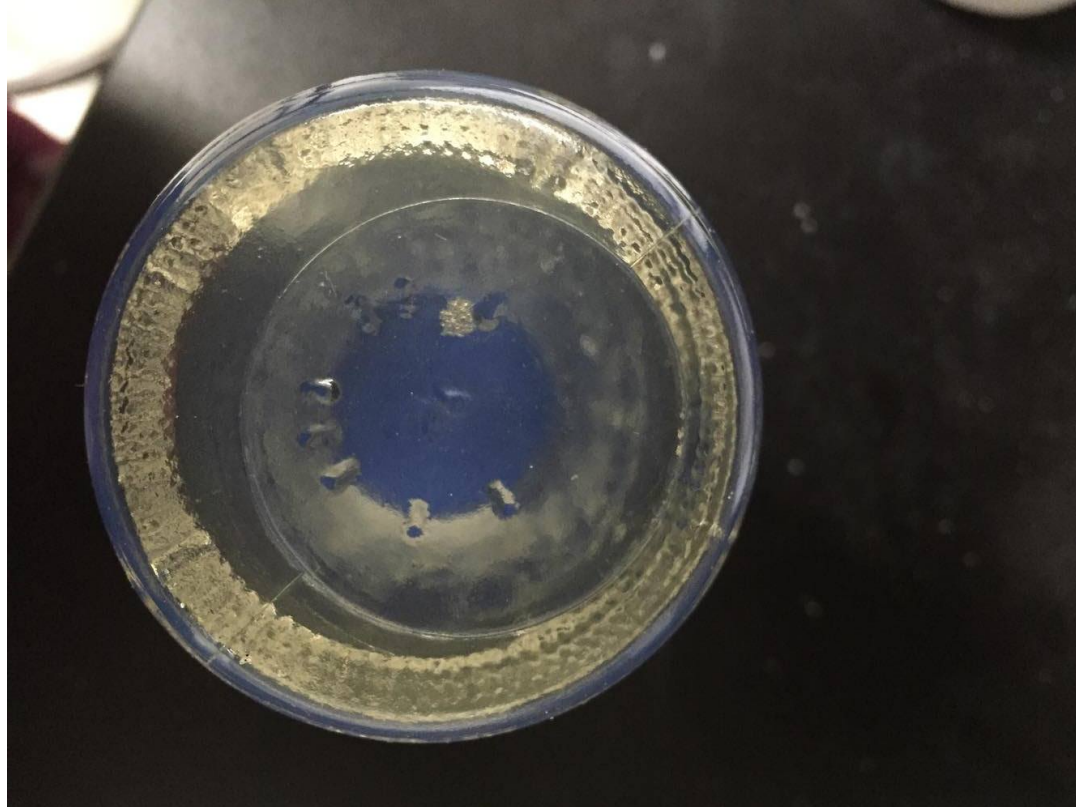


First Iteration



1g Gelatin

Sixth Iteration



2.5g Gelatin