# New cardiovascular biomarkers in animal models: what can be expected in the coming years?

Mark A. Oyama

Animal models of disease provide the basis for important scientific inquiry of proof of concept, drug safety and efficacy, and exploration of the mechanistic and pathophysiological aspects of disease. Animals with either induced cardiac disease or spontaneously occurring cardiac disease are test beds for new cardiovascular therapies or interventions. Biomarkers in animal models can help characterize disease, stratify risk of morbidity and mortality, and serve as surrogate endpoints. Biomarkers identify important similarities and differences between the animal model and the human condition. The future of biomarkers and animal models includes cross-organ investigation using multiorgan multimarker panels. Industry will use animal models and biomarkers to help shorten the time from drug discovery to market. Biomarker development has outpaced their acceptance in clinical use

Animal models of disease provide the basis for important scientific inquiry of proof of concept, drug safety and efficacy, and exploration of the mechanistic and pathophysiological aspects of disease. In particular, studies using animals with either induced cardiac disease or spontaneously occurring cardiac disease (SOCD) are of interest insofar as cardiovascular disease represents one of the leading causes of death and disability worldwide. The potential advantages of induced disease models include their neurohormonal and functional characteristics similar to human heart failure, reliability, and presence of a large amount of previously published data using these models. Commonly used induced models of myocardial or valvular disease include rapid ventricular pacing [1], microembolization [2], and chordae tendineae rupture [3]. The potential advantages of animals with SOCD include the ability to test interventions in subjects with the actual disease of interest, access to a large number of affected subjects, and avoidance of ethical and animal welfare considerations that accompany induced disease models. SOCD is relatively common in veterinary species such as the dog, and particular forms of SOCD in dogs are highly homologous to the corresponding human disease. Examples include myocardial diseases such as dilated cardiomyopathy [4,5] and arrhythmogenic right ventricular cardiomyopathy (ARVC) [6] and valvular diseases such as mitral valve prolapse [7] and myxomatous mitral valve disease (MMVD) [8]. Irrespective of whichever model is utilized, characterization of the disease state in individual subjects must be comprehensive, valid, and reliable. This is typically easier to accomplish in induced disease models where the subjects are often of the same age, sex and breed, and

and carefully designed clinical studies are needed to achieve their maximum potential. *Cardiovasc Endocrinol* 3:27-31 © 2014 Wolters Kluwer Health | Lippincott Williams & Wilkins.

Cardiovascular Endocrinology 2014, 3:27-31

Keywords: animal model, biomarkers, dog

Department of Clinical Studies, Philadelphia School of Veterinary Medicine, University of Pennsylvania, Philadelphia, Pennsylvania, USA

Correspondence to Mark A. Oyama, DVM, Department of Clinical Studies Philadelphia, School of Veterinary Medicine, University of Pennsylvania, 3900 Delancey St, Philadelphia, PA 19104, USA Tel: +1 215 898 8889; fax: +1 215 573 6232; e-mail: maoyama@vet.upenn.edu

Received 14 September 2013 Accepted 7 January 2014

whose diet, environment, and activity level can be closely controlled. The obvious disadvantage of induced disease models, from a biological standpoint, is that while similar, the induced disease often has important differences with respect to the rate of onset and degree of ventricular remodeling to the actual disease of interest.

Biomarkers are objective measures that predict the presence, risk, or outcome of disease. As such, they act as surrogates for clinically relevant conditions (i.e. presence of disease) or outcomes (i.e. heart failure, death). Cardiovascular biomarkers are increasingly being used to help diagnose, stage, monitor, and predict outcome of cardiovascular disease in both humans and animals [9,10]. The advantages of using biomarkers include their objectivity, potential cost-effectiveness, and in many cases, their pathophysiological link with the disease of interest, such that their study yields a deeper understanding of the mechanisms of disease [11]. Industry uses biomarkers as surrogate markers in animal models to help speed the development of new drugs, especially in diseases that progress slowly. The number and nature of potential cardiovascular biomarkers under investigation are both plentiful and diverse. This review will discuss future veterinary trends involving cardiovascular biomarkers and briefly describe some of the promising biomarkers under study in both human and animal models.

## Cardiac biomarkers in animal models of cardiovascular disease: 10 years hence

What will the use of biomarkers in animal models of disease look like 10 years into the future? What are the

DOI: 10.1097/XCE.00000000000026

Copyright © Lippincott Williams & Wilkins. Unauthorized reproduction of this article is prohibited.

most promising applications and where should we expend the majority of our resources to maximize return? As stated previously, induced disease animal models, by applying a uniform cardiac injury to each subject, offer homogeneity with respect to the severity and expected progression of disease across individuals. By controlling these variables, extraneous noise is reduced and the ability to make inferences on the effects of a treatment, for example, is increased. In contrast, SOCD models by nature are much more heterogeneous as the affected population includes individuals of varying ages, sexes, body habitus, severity of disease, duration of disease, and risk of future adverse events. These potential confounders can obscure the effect of an intervention and increase the number of subjects that must be studied.

If eligible subjects with SOCD could be better stratified into groups with similar clinical and biochemical characteristics, or if individuals at highest risk for adverse events could be selected from among the entire population, the use of SOCD models would be highly attractive. To illustrate this point, consider the case of a SOCD such as MMVD. In dogs, the clinical spectrum of MMVD ranges from mild mitral valve prolapse to ruptured chordae tendineae and torrential mitral regurgitation. In animals with preclinical MMVD, the rate of disease progression is highly variable and difficult to predict. In veterinary species with SOCD, biomarkers provide insights into the severity of disease, risk of morbidity and mortality, and response to therapy, such that populations of diseased individuals can be subdivided into groups with similar disease characteristics, risks, and outcomes [12,13]. Heart disease represents a significant societal burden with respect to healthcare costs and resources, and identification of individuals at highest risk for disease progression is of interest. The prediction of adverse events such as hospitalization for heart failure in an individual patient, be it dog or human, is difficult. The cardiac natriuretic peptide NT-proBNP, as has been discussed previously in this issue, stratifies the risk of morbidity and mortality in both dogs and humans. Characterization of study populations partly based on NT-proBNP or other relevant biomarkers could facilitate the study of a specific predisposed subgroup of humans by leveraging the ability to closely match them to their high-risk canine counterparts.

Biomarkers can also highlight the differences between SOCD and the corresponding human disease. An example is illustrated by ARVC. ARVC in the dog shows a high degree of clinical homology with human ARVC [6]. Both conditions are characterized by a high incidence of ventricular tachycardia, syncope, and sudden death, and variable degrees of myocardial failure and congestive heart failure. ARVC is particularly common in the Boxer breed of dog and is diagnosed on the basis of clinical signs of syncope and evidence of ventricular arrhythmias with right bundle branch block morphology [6]. The close clinical features between the two species suggest that new antiarrhythmic strategies in the dog could lead to better outcomes in humans. In the Boxer dog, a single genetic mutation associated with ARVC has been described [14]. However, this particular mutation, involving the *striatin* gene, is not one of the 12 different genetic subtypes of ARVC that have been described in humans. Here, the difference in biomarkers can be considered a glass half-empty (are findings from Boxer ARVC studies really applicable to humans?) or half-full (the potential for mutations in the *striatin* gene should be evaluated in humans with ARVC).

One trend in biomarker science likely to encompass both animal and human disease is the use of multiple complementary markers for a particular disease entity. The value of simultaneously assessing multiple biomarkers can be additive or synergistic. A study has shown that NT-proBNP and cardiac troponin-I provide complementary prognostic information in dogs with SOCD [12]. Dogs with elevations of both NT-proBNP and cardiac troponin-I fare worse than those with elevations of either biomarker alone. This latter group fares worse than dogs with neither biomarker elevated. In human patients, the multimarker approach has proven valuable in heart failure [15,16], myocardial infarction [17], and acute dyspnea [18]. Thus, carefully designed 'biomarker panels' that provide a cross-sectional profile of multiple markers will likely provide superior information than testing a single marker alone. Some of the most promising markers being studied in animal models and humans are listed in Table 1. These markers are grouped under broad headings that reflect their purported role in the pathophysiology of disease. Panels that include a sampling of markers across different pathophysiological categories likely provide superior information than testing multiple markers with a high degree of correlation within categories. The multimarker strategy for predicting likelihood of a theoretical outcome is most discriminative if their individual components, while all reflecting important aspects of the disease process, are only weakly correlative to each other (Fig. 1). This strategy is similar to looking at the disease from a variety of different 'viewpoints' in the hope of getting the most complete and accurate picture.

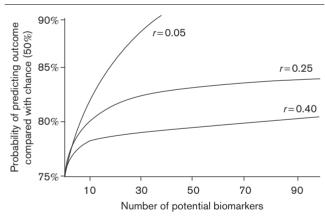
Regulation of the cardiovascular system involves a complex interplay between many different organ systems, including the heart, kidney, vasculature, and autonomic nervous system, and biomarker panels involving more than a single organ system are a subject of considerable interest. In diseases such as congestive heart failure or acute kidney injury, biomarkers with a wider 'cardiorenal' perspective have been proposed. Cardiorenal syndrome has been described as 'disorders of the heart and kidneys whereby acute or chronic dysfunction in one organ may

Copyright © Lippincott Williams & Wilkins. Unauthorized reproduction of this article is prohibited.

### Table 1 Incomplete list of potential circulating biomarkers in animal models and humans with cardiovascular disease

Acute and chronic heart failure NT-proBNP/BNP cTnl/hscTnl
Galectin-3 ST-2
Mid-regional proadrenomedullin Copeptin sST-2
Mid-regional proatrial natriuretic peptide (MR-proANP) MicroRNAs (mRNAs)
Growth differentiation factor-15 (GDF-15)
Extracellular matrix
Soluble endoglin
Osteopontin
PIIINP
MMPs
Glycoprotein 130 (gp130)
Inflammatory
Heat shock protein
Vascular cell adhesion molecule-1 (VCAM-1)
C-reactive protein Interleukin-1
Interieukin- i Metabolism
Leptin Branched chain amino acid metabolites
Cardiorenal disease
Neutrophil gelatinase-associated lipocalin (NGAL)
Cystatin C
Kidney injury molecule-1 (KIM-1)

Fig. 1



Incremental value in adding potential biomarkers of varying markermarker correlation (*r*). The *y*-axis shows the probability of the hypothetical outcome measure as compared with chance (50% probability). The *x*-axis shows the number of markers used to help determine the probability of the outcome. The greatest incremental gain in the probability occurs when the markers used are only weakly correlated with each other (r=0.05), as opposed to a lesser amount of gain if the markers are more strongly correlated to each other (r=0.25 or r=0.40). Adapted from Roberts *et al.* [19].

induce acute or chronic dysfunction of the other' [20]. Several cardiorenal biomarkers (Table 1) have been proposed as a means to specifically identify cardiorenal syndrome, stratify injury severity, and to guide subsequent therapy. New and existing markers will undoubtedly undergo testing in animal models of disease. In the author's opinion, a multiorgan multimarker strategy is one of the most tantalizing aspects of biomarker studies.

The science of new biomarkers in both animals and humans is evolving rapidly. Contributions to this evolution involve a variety of interested parties. Biopharmaceutical companies are interested in using biomarkers to assess toxicity as well as surrogate endpoints to speed drug development for slowly progressing diseases [21]. Physicians and veterinarians seek biomarkers that identify high-risk patients or patients who are most likely to respond favorably to a given therapy. Diagnostic testing companies are eager to bring new discoveries into the market. These and other forces are driving the search for new cardiovascular biomarkers, and the pace of discovery has largely overtaken the rate of acceptance. The excitement surrounding newly discovered biomarkers needs to be justified through carefully designed studies that validate the benefit of these markers in clinical practice.

#### New cardiovascular biomarkers

A broad variety of molecules involved in heart failure, tissue remodeling, inflammation, the extracellular matrix, and metabolism are being studied. An exhaustive review of emerging biomarkers is beyond the scope of this article, but a selection of some intriguing molecules is now presented.

#### MicroRNA

MicroRNA (miRNA) are short noncoding RNAs that block translation and speed breakdown of specific target messenger RNA. miRNA play important roles in many different disease processes including cardiovascular, kidney, neoplastic, and neurologic disease. miRNA help regulate a variety of aspects of cardiovascular function including hypertrophy, fibrosis, apoptosis, cardiac conduction, and energy utilization [22]. An example of miRNA in cardiac disease involves atrial fibrillation. A wellestablished animal model of atrial fibrillation is induced atrial fibrillation in dogs [23]. In affected dogs, miRNA-29 and miRNA-26 are downregulated and contribute toward pathological fibrotic atrial remodeling [24] and overexpression of the inward-rectifying K+ current [25], features that are important characteristics of atrial fibrillation in humans. Recently, the study of miRNA in heart disease has included dogs with SOCD [26,27].

#### Matrix metalloproteinases

Matrix metalloproteinases (MMPs) and their inhibitors belong to a family of endopeptidases capable of degrading and remodeling extracellular matrix. MMPs are involved in a wide variety of cell functions including apoptosis, differentiation, and proliferation. A role for MMPs in cardiac disease is well established in humans [28], induced-disease models [29,30], and SOCD [31,32]. Animal models of disease will be useful in determining whether pharmacological modulation of MMP activity can reduce pathological cardiac remodeling [33].

#### Leptin

Leptin is an adipose tissue-derived hormone that helps regulate energy expenditure, metabolism, and hunger. Leptin's effect on the cardiovascular system can be direct, through its effects on cardiomyocyte fatty acid utilization, hypertrophy, inflammation, and fibrosis, as well as indirect, through effects on renal function, insulin resistance, and body weight [34]. In dogs with SOCD, leptin gene transcription was increased in the myocardium and blood compared with healthy controls, and was highest in the dogs with the most severe heart failure [35]. In human patients, leptin expression has been associated with increased morbidity and mortality [36]. This effect is mediated, in part, by activation of inflammatory cytokines and increased redox stress within myocardiocytes [37].

#### Neutrophil gelatinase-associated lipocalin

Neutrophil gelatinase-associated lipocalin (NGAL) is a glycoprotein that is released by renal tubular cells secondary to kidney injury. Once in circulation, NGAL promotes apoptosis and inflammation of other organs, including the heart. In one study of human patients, NGAL was superior to NT-proBNP in predicting adverse cardiac events of acute decompensated heart failure within 30 days of discharge from the hospital [38]. Moreover, patients with elevations of both NGAL and NT-proBNP were identified to be at highest risk, indicating the potential advantage of a multimarker strategy. In another study, elevated urinary NGAL in patients being treated for acute heart failure was a superior determinant of worsening renal function at 72–96 h versus conventional means [39]. Elevated urinary NGAL has been reported in dogs with both spontaneous and acute kidney injury [40,41].

In summary, the use of biomarkers in animal models of disease is likely to increase. Testing of biomarkers in animals with SOCD will likely grow as awareness of these models increases within the medical community. Multimarker multiorgan approaches are particularly intriguing. The best chance of continuing success within the field of biomarkers will require cooperation between many different parties, including veterinary and medical researchers and clinicians and industry.

#### Acknowledgements Conflicts of interest

There are no conflicts of interest.

#### References

- 1 Yarbrough WM, Spinale FG. Large animal models of congestive heart failure: a critical step in translating basic observations into clinical applications. *J Nucl Cardiol* 2003; **10**:77–86.
- 2 Power JM, Tonkin AM. Large animal models of heart failure. *Aust N Z J Med* 1999; **29**:395–402.

- 3 Leroux AA, Moonen ML, Pierard LA, Kohl P, Amory H. Animal models of mitral regurgitation induced by mitral valve chordae tendineae rupture. *J Heart Valve Dis* 2012; 21:416–423.
- 4 Freeman LM, Rush JE. Nutrition and cardiomyopathy: lessons from spontaneous animal models. Curr Heart Fail Rep 2007; 4:84–90.
- 5 Hamlin RL. Animal models of ventricular arrhythmias. *Pharmacol Ther* 2007; **113**:276–295.
- 6 Basso C, Fox PR, Meurs KM, Towbin JA, Spier AW, Calabrese F, et al. Arrhythmogenic right ventricular cardiomyopathy causing sudden cardiac death in Boxer dogs: a new animal model of human disease. *Circulation* 2004; 109:1180–1185.
- 7 Pedersen HD, Haggstrom J. Mitral valve prolapse in the dog: a model of mitral valve prolapse in man. *Cardiovasc Res* 2000; **47**:234–243.
- 8 Aupperle H, Disatian S. Pathology, protein expression and signaling in myxomatous mitral valve degeneration: comparison of dogs and humans. J Vet Cardiol 2012; 14:59–71.
- 9 Silver MA, Maisel A, Yancy CW, McCullough PA, Burnett JC Jr, Francis GS, et al. BNP Consensus Panel 2004: a clinical approach for the diagnostic, prognostic, screening, treatment monitoring, and therapeutic roles of natriuretic peptides in cardiovascular diseases. *Congest Heart Fail* 2004; 10:1–30.
- 10 Oyama MA, Boswood A, Connolly DJ, Ettinger SJ, Fox PR, Gordon SG, et al. Clinical usefulness of an assay for measurement of circulating N-terminal pro-B-type natriuretic peptide concentration in dogs and cats with heart disease. J Am Vet Med Assoc 2013; 243:71–82.
- Mayeux R. Biomarkers: potential uses and limitations. *NeuroRx* 2004; 1:182–188.
- 12 Hezzell MJ, Boswood A, Chang YM, Moonarmart W, Souttar K, Elliot J, et al. The combined prognostic potential of serum high-sensitivity cardiac troponin I and N-terminal pro-B-type natriuretic peptide concentrations in dogs with degenerative mitral valve disease. J Vet Intern Med 2012; 26:302–311.
- 13 Wolf J, Gerlach N, Weber K, Klima A, Wess G. Lowered N-terminal pro-Btype natriuretic peptide levels in response to treatment predict survival in dogs with symptomatic mitral valve disease. *J Vet Cardiol* 2012; 14: 399–408.
- 14 Meurs KM, Mauceli E, Lahmers S, Acland GM, White SN, Lindblad-Toh K, et al. Genome-wide association identifies a deletion in the 3' untranslated region of striatin in a canine model of arrhythmogenic right ventricular cardiomyopathy. *Hum Genet* 2010; **128**:315–324.
- 15 Bjurman C, Jensen J, Petzold M, Hammarsten O, Fu ML, et al. Assessment of a multimarker strategy for prediction of mortality in older heart failure patients: a cohort study. *BMJ Open* 2013; **3**:e002254–e002254.
- 16 Alehagen U, Dahlstrom U, Rehfeld JF, Goetze JP. Pro-A-type natriuretic peptide, proadrenomedullin, and N-terminal pro-B-type natriuretic peptide used in a multimarker strategy in primary health care in risk assessment of patients with symptoms of heart failure. J Card Fail 2013; 19:31–39.
- 17 Fertin M, Dubois E, Belliard A, Amouyel P, Pinet F, Bauters C, et al. Usefulness of circulating biomarkers for the prediction of left ventricular remodeling after myocardial infarction. Am J Cardiol 2012; **110**:277–283.
- 18 Rehman SU, Martinez-Rumayor A, Mueller T, Januzzi JL Jr. Independent and incremental prognostic value of multimarker testing in acute dyspnea: results from the ProBNP Investigation of Dyspnea in the Emergency Department (PRIDE) study. *Clin Chim Acta* 2008; **392**:41–45.
- 19 Roberts LD, Gerszten RE. Toward new biomarkers of cardiometabolic diseases. Cell Metab 2013; 18:43–50.
- 20 Ronco C, McCullough P, Anker SD, Anand I, Aspromonte N, Bagshaw SM, et al. Cardio-renal syndromes: report from the consensus conference of the acute dialysis quality initiative. *Eur Heart J* 2010; **31**:703–711.
- 21 IBM Institute of Business Value. *IBM Global Business Services: the evolving role of biomarkers.* Somers, NY: IBM Global Services; 2007.
- 22 Small EM, Frost RJ, Olson EN. MicroRNAs add a new dimension to cardiovascular disease. *Circulation* 2010; **121**:1022–1032.
- 23 Urban JF, Gerhart RL, Krzeszak JR, Leet CR, Lentz LR, McClay CB, et al. Methods for the development and assessment of atrial fibrillation and heart failure dog models. J Geriatr Cardiol 2011; 8:133–140.
- 24 Dawson K, Wakili R, Ordog B, Clauss S, Chen Y, Iwasaki Y, et al. MicroRNA29: a mechanistic contributor and potential biomarker in atrial fibrillation. *Circulation* 2013; **127**:1466–1475, 1475e1–1475e28.
- 25 Luo X, Pan Z, Shan H, Xiao J, Sun X, Wang N, et al. MicroRNA-26 governs profibrillatory inward-rectifier potassium current changes in atrial fibrillation. J Clin Invest 2013; 123:1939–1951.
- 26 Steudemann C, Bauersachs S, Weber K, Wess G. Detection and comparison of microRNA expression in the serum of Doberman Pinschers with dilated cardiomyopathy and healthy controls. *BMC Vet Res* 2013; 9:12.

- 27 Von Deetzen MC, Schmeck B, Gruber AD, Klopfleisch R. Molecular quantification of canine specific microRNA species. *Res Vet Sci* 2013; 95:562–568.
- 28 Spinale FG. Matrix metalloproteinases: regulation and dysregulation in the failing heart. *Circ Res* 2002; **90**:520–530.
- 29 Ahmed FZ, Khattar RS, Zaidi AM, Neyses L, Oceandy D, Mamas M, et al. Pacing-induced cardiomyopathy: pathophysiological insights through matrix metalloproteinases. *Heart Fail Rev* 2013 [Epub ahead of print].
- 30 Bradham WS, Moe G, Wendt KA, Scott AA, Konig A, Romanova M, et al. TNF-alpha and myocardial matrix metalloproteinases in heart failure: relationship to LV remodeling. Am J Physiol Heart Circ Physiol 2002; 282:H1288–H1295.
- 31 Fonfara S, Hetzel U, Tew SR, Cripps P, Dukes-McEwan J, Clegg PD, et al. Expression of matrix metalloproteinases, their inhibitors, and lysyl oxidase in myocardial samples from dogs with end-stage systemic and cardiac diseases. Am J Vet Res 2013; 74:216–223.
- 32 Ljungvall I, Rajamaki MM, Crosara S, Olsen LH, Kvart C, Borgarelli M, et al. Evaluation of plasma activity of matrix metalloproteinase-2 and -9 in dogs with myxomatous mitral valve disease. Am J Vet Res 2011; 72: 1022–1028.
- 33 Briasoulis A, Tousoulis D, Papageorgiou N, Kampoli AM, Androulakis E, Antoniades C, *et al.* Novel therapeutic approaches targeting matrix metalloproteinases in cardiovascular disease. *Curr Top Med Chem* 2012; 12:1214–1221.
- 34 Abel ED, Sweeney G. Modulation of the cardiovascular system by leptin. Biochimie 2012; 94:2097–2103.

- 35 Fonfara S, Hetzel U, Tew SR, Dukes-McEwan J, Cripps P, Clegg PD, et al. Leptin expression in dogs with cardiac disease and congestive heart failure. J Vet Intern Med 2011; 25:1017–1024.
- 36 Yin WH, Wei J, Huang WP, Chen JE, Young MS, Lin SJ, et al. Prognostic value of circulating adipokine levels and expressions of adipokines in the myocardium of patients with chronic heart failure. Circ J 2012; 76:2139–2147.
- 37 Bobbert P, Jenke A, Bobbert T, Rauch U, Lassner D, Scheibenbogen C, et al. High leptin and resistin expression in chronic heart failure: adverse outcome in patients with dilated and inflammatory cardiomyopathy. Eur J Heart Fail 2012; 14:1265–1275.
- 38 Maisel AS, Mueller C, Fitzgerald R, Brikhan R, Hiestand BC, Iqgbal N, et al. Prognostic utility of plasma neutrophil gelatinase-associated lipocalin in patients with acute heart failure: the NGAL EvaLuation Along with B-type NaTriuretic Peptide in acutely decompensated heart failure (GALLANT) trial. *Eur J Heart Fail* 2011; **13**:846–851.
- 39 Collins SP, Hart KW, Lindsell CJ, Fermann GJ, Weintraub NL, Miller KF, et al. Elevated urinary neutrophil gelatinase-associated lipocalcin after acute heart failure treatment is associated with worsening renal function and adverse events. Eur J Heart Fail 2012; 14:1020–1029.
- 40 Kai K, Yamaguchi T, Yoshimatsu Y, Kinoshita J, Teranishi M, Takasaki W, et al. Neutrophil gelatinase-associated lipocalin, a sensitive urinary biomarker of acute kidney injury in dogs receiving gentamicin. J Toxicol Sci 2013; 38:269–277.
- 41 Lee YJ, Hu YY, Lin YS, Chang CT, Lin FY, Wong ML, *et al.* Urine neutrophil gelatinase-associated lipocalin (NGAL) as a biomarker for acute canine kidney injury. *BMC Vet Res* 2012; 8:248.