# Immunomodulatory effects of opioids

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#### Abstract

**Objective** – To review the immunomodulatory effects of opioids. **Data Sources** – Original research publications and review articles using the PubMed search engine with the following keywords – opioids, morphine, immuomodulation, and immunosuppression.

**Veterinary and Human Data Synthesis** – Opioids have been shown to modulate the immune system in animal models by affecting both the acquired and innate arms of the immune system. Natural killer cell activity, T-cell proliferation, antibody production, phagocytic cell function, and cytokine production have all been shown to be affected by opioids. Many of these effects are reversed by opioid antagonists. Opioids have also been shown to induce sepsis in laboratory animals. Opioid administration alters immune parameters in healthy humans at analgesic doses and may increase the risk of infection in some patient populations.

**Conclusions** – While opioids remain the most powerful and widely used analgesics available, their negative effects on the immune system are well established in the laboratory setting. Thoughtful consideration should be given to the use of certain opioids in critically ill patients, especially those with pre-existing immunocompromise.

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## Introduction

The term opioid refers to any directly acting compound that is stereospecifically antagonized by naloxone.<sup>1</sup> The clinical use of opioids stems from more than 5000 years of medicinal use of opium, which is derived from the unripe seed capsule of poppy *Papaver somniferum*. Morphine, a pure mu receptor agonist opioid, was isolated from opium in 1805; it is also known as an opiate as it is not a synthetically compounded drug. Since that time other semisynthetic and synthetic opioids have been developed in an attempt to produce more efficacious compounds with reduced abuse potential and adverse effects.<sup>1</sup>

Opioids are the most powerful and effective analgesic drugs used in clinical medicine. They make up the backbone of pain management in human and veteri-

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nary patients and are used for the management of moderate to severe pain.<sup>2,3</sup> They are potent analgesics, have few clinically apparent adverse effects on other organ systems, and have consistent pharmacokinetics.<sup>4</sup> Opioids are naturally occurring alkaloids, related synthetic/semisynthetic compounds, and endogenous opioid peptides (enkephalins, endorphins, and dynorphins).<sup>5</sup> The exact physiologic function of endogenous opioids includes modulation of stress-induced analgesia and neurotransmitter, neuromodulator, and neurohormonal activity.<sup>6</sup> In addition to analgesic properties, opioid receptor agonists and antagonists modulate the immune system by interacting with opioid receptors in the central nervous system (CNS) and on immune cells.

#### Pharmacology

Opioid receptors are named for their endogenous ligands and identified by a numerical subscript corresponding to the chronological order of discovery by cloning and sequencing.<sup>1</sup> The receptors for which all opioids act as agonists have a generic designation of OP (Table 1). The opioid receptor is made up of an extracellular N-terminal domain, 7 transmembrane guanine domains connected by 3 extracellular and 3 intracellular loops and an intracellular C-terminal tail.<sup>7,8</sup> Each

International Union of Pharmacology recommendation	Pharmacology nomenclature	Molecular biology nomenclature			
OP <sub>1</sub>	δ	DOR			
OP <sub>2</sub>	κ	KOR			
OP <sub>3</sub>	μ	MOR			

Table 1: Opioid nomenclature

Adopted from Dhawan BN et al.145

DOR, delta opioid receptor; KOR, kappa opioid receptor; MOR, mu opioid receptor.

receptor has specific subtypes that have not been completely defined.<sup>9,10</sup> These subtypes may dictate the physiologic response to endogenous and exogenous opioids and explain variable analgesic and physiologic responses associated with OP<sub>3</sub> receptor opioids.<sup>11</sup>

The frequency and distribution of opioid receptors is organ and species dependent and nonuniform.<sup>5,8,12</sup> The opioid receptor has been shown to be distributed in many sites throughout the CNS including the cerebral cortex, thalamus, periaqueductal gray matter, and the spinal cord dorsal horn. In addition, opioid receptors have been demonstrated on peripheral sensory neurons following inflammation and nonneurologic tissue including the heart, gastrointestinal tract and cells of the immune system.<sup>6,9,10,13,14</sup> Opioid receptors in these different areas all exhibit similar molecular structure.<sup>13</sup>

Opioid receptors signal through a second messenger system (cyclic AMP) or ion channel (K<sup>+</sup>).<sup>6</sup> They work by reducing intracellular calcium concentrations leading to a decrease in presynaptic neurotransmitter release.<sup>6,10</sup> They may also enhance efflux of potassium leading to hyperpolarization of postsynaptic neurons.<sup>6</sup> Opioids inhibit  $\gamma$ -aminobutric acid (GABA)ergic transmission in a local circuit, ultimately preventing GABA inhibition of descending antinociceptive pathways.<sup>6,10</sup> Binding of endogenous and exogenous opioid ligands to the opioid receptor results in receptor activation and

a variety of effects, including analgesia, euphoria, feeding, hormone secretion, respiratory depression, inhibition of gastrointestinal motility, and anxiolysis.<sup>8</sup>

Opioids may interact with receptors as agonists, antagonists, or as partial agonists. Morphine is the prototypical agonist of the OP<sub>3</sub> receptor and is standard of comparison for other opioid analgesics.<sup>6,15,16</sup> Other drugs that act as agonists at this receptor include hydromorphone, oxymorphone, methadone, fentanyl, and meperidine (Table 2). Buprenorphine has traditionally been described as a partial agonist at the OP<sub>3</sub> receptor and an antagonist at the OP<sub>2</sub> receptor.<sup>17</sup> Opioid antagonists lack in vivo or in vitro agonist activity and may be used to reverse the effect of opioid antagonists.<sup>12</sup> Naloxone, naltrexone, and nalmephine are opioid antagonists that block central and peripheral opioid effects.<sup>12</sup> In addition, specific peripheral opioid antagonists have been developed and are used to block undesired peripheral effects (ie, altered gastrointestinal motility) without reversing central analgesic effects of the drugs.<sup>12</sup>

#### Immunomodulatory effects of Opioids

As early as the late 19th century, the immunosuppressive effects of opioids were recognized. Both exogenous and endogenous opioid peptides have been shown to cause immunosuppression or immunostimulation.<sup>23–25</sup> Morphine-induced immunomodulation has been investigated most extensively. However, other opioids have immunomodulatory properties.<sup>26</sup>

The immunosuppressive effects of opioids are independent of their antinociceptive properties and appear to be related to the molecular structure of the opioid. Some opioids (morphine, fentanyl, methadone, codeine) are more immunosuppressive than others (hydromorphone, tramadol, hydrocodone, and oxycodone) (Table 3).<sup>26–28</sup> Buprenorphine produces little to no negative immune alterations and may enhance immune function.<sup>26,27,29–31</sup> Antagonists at the OP<sub>3</sub> receptor also appear to enhance immune function.<sup>13,23,32</sup> Opioids

OP <sub>3</sub> receptor agonist	Administration route	Relative potency to morphine	Additional comments
Hydromorphone	IV, IM, SQ, PO	5–10 <sup>18</sup>	May cause panting in dogs. Minimal to no histamine release
Oxymorphone	IV, IM, SQ, PO	10 <sup>19</sup>	Expensive. Minimal to no histamine release
Fentanyl	IV, TD	80–100 <sup>20</sup>	Short acting. Usually given as a constant rate infusion to attain prolonged therapeutic effects. Transdermal application takes 12–24 hours to reach therapeutic levels
Methadone	IV, IM, SQ, PO	3.4–10 <sup>21,22</sup>	Also has NMDA antagonist properties.

Table 2: OP<sub>3</sub> receptor agonists and their relative potencies to morphine

IV, intravenous; IM, intramuscular; SQ, subcutaneous; TD, transdermal; PO, oral; NMDA, N-methyl-D-aspartic acid.

Table 3:	Immunomodulating	properties	of selected	opioids
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Opioid	Primary receptor activity	HPA axis	Corticosteroid secretion	NK cell activity	Additional information
Morphine	OP <sub>3</sub> agonist	Stimulates <sup>31,34</sup>	Increases <sup>31,34</sup>	Decrease <sup>28,31,35</sup>	Generally upregulates production of pro-inflammatory mediators and suppresses the production of anti- inflammatory mediators. Suppresses immune cell proliferation <sup>31,33</sup>
Fentanyl	OP <sub>3</sub> agonist	Stimulates <sup>34</sup>	Increases <sup>34</sup>	Decreases, has also been reported to increase. May be dose dependent <sup>29,36,37</sup>	Does not stimulate the release of nitric oxide. <sup>38</sup> Suppresses T cell proliferation, IFN- $\gamma$ and IL-2 production <sup>29,39</sup>
Hydromorphone	OP <sub>3</sub> agonist	No known reported effect	No known reported effect	No effect <sup>33</sup>	Has no effect on immune cell proliferation or IL-2 production
Buprenorphine	OP <sub>3</sub> partial agonist	No effect <sup>31,34</sup>	No effect, decreases <sup>29,31</sup>	No effect <sup>29,31</sup>	No effects on immune cell proliferation or cytokine activity <sup>29</sup>
Naloxone	OP₃ antagonist	Variable and dose dependent <sup>40</sup>	Variable <sup>40</sup>	Increase, decrease, or no effect <sup>41,42</sup>	Not well studied. Decreases IL-4 Increases the production of IL-2 and IFN-γ. Increases T lymphocyte proliferation <sup>42</sup>
Naltrexone	OP <sub>3</sub> antagonist	Variable and dose <sup>40</sup> dependent	Variable <sup>40</sup>	Increase, decrease, or no effect <sup>41</sup>	Not well studied. Effects likely similar to naloxone

HPA, hypothalamo-pituitary axis; NK, natural killer; IL, interleukin; IFN, interferon.

with no immunosuppressive activity have a carbonyl substitution at C<sub>6</sub>, a single bond between C<sub>7–8</sub> and a hydroxyl group at C<sub>14</sub>.<sup>27,33</sup> Molecules like morphine and fentanyl that carry the hydroxyl group at both C<sub>3</sub> and C<sub>6</sub> have the greatest immunomodulatory effects; the mechanism by which these structural differences dictate immune response is unknown.<sup>33</sup>

#### Mechanisms of Immunomodulation

The mechanism by which opioids affect the immune system is complex and incompletely understood.<sup>43</sup> Opioids alter immune response through their actions in the CNS and in the periphery. Central mechanisms involve interaction of opioids with opioid receptors in the CNS, which leads to changes in neuroendocrine and autonomic function.<sup>44</sup> The OP<sub>3</sub> receptor is primarily responsible for the central immunomodulatory effect of opioids; OP<sub>3</sub> receptor knockout mice are protected from these immunomodulatory effects.<sup>23,45,46</sup> Centrally mediated suppression is responsible for reduction in natural killer (NK) cell activity (associated with the sympathetic nervous system), reduction in lymphocyte proliferation and interferon (IFN)-γ activity.47,48 Direct stimulation of OP<sub>1</sub> and OP<sub>3</sub> receptors on immune cells including B lymphocytes, monocytes, and macrophages is primarily responsible for peripheral immunosuppression, although a naloxone-insensitive morphine receptor may also play a role.<sup>23</sup>

### Centrally mediated immunomodulation

Several studies have demonstrated a centrally mediated mechanism for opioid-related immunosuppresion.<sup>49–52</sup> Hernandez et al<sup>49</sup> demonstrated the role of the CNS by showing that systemic administration of morphine, which crosses the blood brain barrier, suppressed lymphocyte activity. In contrast, *N*-methylmorphine, an active morphine analog that does not cross the blood-brain barrier, did not produce any systemic changes in lymphocyte activity. The role of central OP<sub>3</sub> receptors was further demonstrated by the observation that injection of *N*-methylmorphine directly into the third ventricle suppressed peripheral lymphocyte and NK cell activity.<sup>49,50</sup>

The periaqueductal gray matter of the mesencephalon (PAG) is enriched with opiate receptors and endogenous opioids. Thus, it is not surprising that the PAG is also the primary area of the brain involved in opioid-induced immunomodulation.<sup>31,47,53</sup> Microinjections of morphine into the PAG cause rapid suppression of NK cell function, while injection into other neuroanatomical sites does not cause immunosuppression.<sup>53</sup> While there are OP<sub>1</sub>, OP<sub>2</sub>, and OP<sub>3</sub> receptors present in the PAG and in many areas of the CNS, OP<sub>3</sub> receptors appear to be primarily responsible for the central immunomodulatory effect of morphine.<sup>51</sup> Administration of high doses OP<sub>1</sub> and OP<sub>2</sub> receptor agonists into the CNS of mice only produced mild suppression of blood lymphocyte proliferation in comparison to OP<sub>3</sub> agonists.<sup>51</sup>

While the role of opioid receptors in immunosuppression has been established, the pathway by which activation of central opioid receptors results in immunosuppression is debated.<sup>54</sup> It has been proposed that central opioid immunomodulation is either through the neuroendocrine (hypothalamic-pituitary-adrenal [HPA]) axis or the sympathetic nervous system.<sup>53,55,56</sup> Both acute and chronic administration of morphine appears to alter the immune response through different pathways. Acute administration of exogenous opioids primarily affects immune function through the sympathetic nervous system while chronic administration may activate the HPA axis.<sup>55,56</sup>

Activation of the HPA axis results in the activation of a hormonal cascade from the hypothalamus, pituitary (ie, CRH, ACTH) and the secretion of immunosuppressive hormones from the adrenal glands (ie, glucocorticoids) which exert their effect peripherally.<sup>30,56,57</sup> Importantly, opioid agonist administration has been shown to activate the descending pathways of the HPA and hypophysectomy, adrenalectomy, and the use of steroid antagonists abolish the immunosuppressive effects of morphine.<sup>58–61</sup>

The sympathetic nervous system is another pathway through which central opioid receptor activation translates to immunomodulation. Primary (eg, spleen, thymus) and secondary (eg, Peyer's patches, lymph nodes) lymphoid organs are primarily innervated by the sympathetic nervous system.<sup>55</sup> Opioids activate the sympathetic nervous system and induce catecholamine release, including epinephrine from the CNS and norepinephrine from the sympathetic nerve terminals and adrenal medulla. Catecholamines act on primary and secondary lymphoid organs and suppress lymphocyte proliferation, NK cell, and macrophage activity.<sup>47,62–64</sup> Alpha-adrenergic receptor activation is responsible for suppressing NK cell activity while beta-adrenergic receptor activation decrease lymphocyte activity.<sup>47,65</sup>

#### Peripherally mediated immunomodulation

Opioid receptors as well as nonclassical opioid-like receptors are expressed on the surface of immune cells. Additionally, there is evidence of opioid receptors that have yet to be cloned that are responsible for altered immune cell function. For example, novel morphine-selective receptors have been identified on lymphocytes. These opioid receptors are thought to exist in low density on a restricted population of lymphocytes.<sup>25</sup>

The  $OP_1$  receptor is primarily responsible for peripherally mediated immunomodulation, although  $OP_2$  and  $OP_3$  receptors may also play a lesser role.<sup>43</sup>  $OP_1$  receptors on T cells and macrophages are involved in the

maintenance of immune homeostasis. Overstimulation of  $OP_1$  opioid receptors by exogenous or endogenous opioids may alter the production cytokines (ie, enhancing or suppressing cytokine production depending on the agonist and the dose used) and decrease antibody production.<sup>25,55,66</sup>

# Effects of opioids on specific cells of the immune system

Opioids mediate their immunosuppressive effects by acting on various cells of the immune system, affecting both the innate and the adaptive immune systems. Opioids interfere with the phagocytic activities of macrophages and neutrophils, inhibit B and T-cell antibody response, interfere with chemotaxis of immune cells and modulate inflammatory mediator production.<sup>56,67–70</sup>

**Bone marrow progenitor cells:** Roy et al<sup>71</sup> discovered in 1991 that bone marrow cells are differentially sensitive to chronic morphine treatment in vitro. Continuous in vivo morphine treatment compromised the ability of macrophage progenitor cells to proliferate in response to macrophage-colony stimulating factor (M-CSF) in mice. A 70% reduction in colony formation was seen as early as 36 hours after continuous morphine exposure although acute injection of morphine failed to suppress M-CSF induced colony formation. This effect was not permanent and M-CSF responsiveness returned within 5 days after cessation of morphine treatment.<sup>56,71</sup>

Macrophages: Morphine suppresses phagocytic functions of macrophages in vivo and in vitro. Acute and chronic exposure to morphine affects a variety of macrophage functions including phagocytosis, tumoricidal activity, nitric oxide (NO) production, superoxide formation, and cytokine expression.<sup>56,72–76</sup> Suppressed macrophage function is antagonized by OP<sub>3</sub> antagonists but not by OP1 or OP2 antagonists, emphasizing the importance of the OP<sub>3</sub> receptor in immunosuppression.<sup>72</sup> Morphine induced apoptosis in macrophages of mice, rats, and healthy human volunteers and this effect was reversed by naloxone.77 Morphineinduced macrophage apoptosis may be mediated through the generation of NO, since the administration of NO-synthase inhibitors attenuated this effect.<sup>77</sup> NO is thought to promote macrophage apoptosis through the generation of p53.78

**NK cells:** Acute and chronic administration of morphine affects the function of human, monkey, and rodent NK cells.<sup>56,79</sup> Administration of morphine in rats and healthy human volunteers decreased splenic NK cell activity within 2–3 hours. This effect was completely antagonized by naltrexone.<sup>48,79,80</sup> The mecha-

nism by which morphine affects NK cell activity is thought to be mediated through OP<sub>3</sub> receptors in the CNS.<sup>35,50,79</sup> Morphine does not affect NK cell function in vitro and administration of *N*-methyl morphine, a morphine analog that does not cross the blood brain barrier, fails to inhibit NK activity in vivo.<sup>50,61,81</sup>

A key aspect of NK cell function is the identification and eradication of neoplastic cells. Therefore, it is logical that opioid-induced NK suppression would lower defenses against neoplastic cell growth. However, in vivo evaluations of this phenomenon have yielded conflicting results. Ishikawa et al<sup>82</sup> demonstrated that morphine enhanced the growth of tumor cells in vivo and this effect was inhibited by naloxone. However, other investigators have demonstrated that morphine controlled metastasis and retarded the growth of tumors.<sup>83,84</sup> It is thought that the antitumor effect of morphine may be the result of analgesia.<sup>84</sup>

**T cells:** Chronic administration of morphine affects the function of T cells and their precursors, in part through the OP<sub>3</sub> receptor.<sup>55,85,86</sup> Chronic morphine administration interferes with the synthesis of interleukin (IL)-2 and IFN- $\gamma$ , while the production of IL-4 and IL-5 are upregulated.<sup>86,87</sup> This ultimately leads to the differentiation of T helper (Th) cells toward Th2 effector cells, via killing of Th1 cells in a Fas/FasL-dependent manner, resulting in reduced cellular immunity.<sup>86–88</sup> Chronic morphine administration decreases CD4/CD8 ratio in the spleen and thymus of mice.<sup>24,46</sup> Morphine also decreases T-cell responses to the mitogen concanavalin A, in mice.<sup>89</sup>

Inflammatory mediators: In general, morphine increases production of proinflammatory cytokines and reduces production of anti-inflammatory cytokines, although this effect is dose dependent. The OP<sub>3</sub> receptor is involved in morphine induced reduction of tumor necrosis factor (TNF) synthesis while the role of this receptor in the modulation of IL-1 $\beta$  and IL-6 synthesis is unclear.<sup>23,56,90</sup> It is speculated that the modulation of IL-1 $\beta$  and IL-6 synthesis could be mediated by OP<sub>2</sub> or OP<sub>3</sub> receptors or through a naloxone-insensitive morphine receptor.<sup>56</sup> OP<sub>1</sub> selective agonist, U50,488 inhibits the synthesis of IL-1 $\beta$  and TNF but not IL-6 in a macrophage cell line.<sup>25,91</sup> Modulation of cytokine expression is thought to be through the nuclear factor- $\kappa$ B pathway.<sup>92</sup>

Morphine inhibits production of IFN- $\gamma$ , an important T cell lymphokine responsible for enhancing the microbicidal activity of macrophages, peripheral blood mononuclear cells, and T lymphocytes.<sup>93,94</sup> Peng et al<sup>95</sup> demonstrated that morphine upregulated the production of proinflammatory cytokines, TNF, and IL-12, at the protein and the mRNA level, and naloxone blocked those effects. Morphine also inhibited the production of IL-10, an anti-inflammatory cytokine.<sup>95</sup> Morphine reduces IL-2 production while hydromorphone and the OP<sub>3</sub> antagonists, naloxone and naltrexone, increase or have no effect on IL-2 production.<sup>33</sup> In one in vitro study, morphine produced a dose dependent effect. Roy et al<sup>92</sup> demonstrated that high doses of morphine reduced the expression of inflammatory cytokines TNF and IL-6, while low doses of morphine increased their expression in vitro. Ultimately, the effect of morphine on inflammatory cytokine synthesis and secretion is variable depending on experimental conditions and the dose of morphine used.<sup>56</sup>

#### Opioids, infection, and sepsis

Certain opioids have been shown to increase infection rates in humans and animal models.<sup>13,43</sup> Morphine increased susceptibility to Klebsiella pneumoniae and Candida albicans infection in rodents, reduced splenic and thymic weights, and decreased T cell proliferative response to mitogens. Additionally, morphine promoted the translocation of intestinal bacteria into the peritoneal cavity inducing sepsis in mice.14,23,44,96 The initial serum concentration of morphine attained in these mice was higher than those achieved with typical analgesia doses in clinical practice, although those concentrations could be attained if higher doses of morphine are used.<sup>14</sup> However, after 48-96 hours serum morphine concentrations were approximately  $0.6 \,\mu g/$ mL, which is comparable to that obtained for analgesia in clinical practice.<sup>14,97</sup>

Along with inducing infection, morphine administration has detrimental consequences in experimental models of sepsis and endotoxemia. Cell-mediated immunity is vital to combat microorganisms during sepsis, and morphine attenuates cell-mediated immunity in murine models of sepsis.<sup>14,98,99</sup> Administration of morphine before administration of lipopolysaccharide (LPS) significantly enhanced development of hypotension, intravascular coagulation, leukocyte endothelial adhesion, and production of TNF, IL-1 $\beta$ , and IL-6 in rats.<sup>99</sup> Morphine hastened the progression of sepsis to septic shock in this group of rodents.<sup>99</sup>

Conversely, buprenorphine and opioid antag onists are beneficial in murine models of sepsis. Buprenorphine improved mean arterial blood pressure, pH, and base excess in endotoxemic rats and attenuated the development of fever after intrathecal injection of LPS in rats.<sup>98,100</sup> In pigs with *Escherichia coli*-induced sepsis, there was significant improvement in hemodynamic status when buprenorphine was administered.<sup>101</sup> Naltrexone reduced plasma TNF concentrations, improved microcirculation and reduced hepatic dysfunction in endotoxemic rats.<sup>102</sup> Additionally, administration of naltrexone protected mice from developing

LPS-induced shock by blunting TNF production.<sup>103</sup> The protective effect of naltrexone during endotoxemia was reversed when morphine was administered.<sup>103</sup> In human clinical trials, naloxone improved mean arterial blood pressure during septic shock.<sup>104–106</sup>

#### Evidence in humans

There is abundant evidence demonstrating the relationship between intravenous drug users (IVDU) and increased susceptibility to infectious disease.57,107 Opioid exposure alone, in the absence of other confounding factors, markedly affects a wide variety of immune parameters in this population.<sup>57,108</sup> While a large percentage of infections in IVDU are associated with high-risk behaviors (eg, use of unsterilized needles) and lifestyle practices, IVDU also have an increased incidence of infections (eg, tuberculosis, pneumocystic pneumonia, systemic candidiasis) that cannot be attributed to high-risk behaviors alone.57,108,109 Individuals that are infected with the human immunodeficiency virus (HIV) that are not IVDU have a lower frequency of infectious diseases and nonacquired immunodeficiency syndrome related deaths than HIV patients who are IVDU.<sup>110</sup>

Heroin users frequently develop lymphadenopathy, lymphocytopenia, hypergammaglobulinemia, and decreased monocyte adhesion and neutrophil function.<sup>57,111</sup> Additionally, IVDU develop opioid-induced endocrine and metabolic dysfunction.<sup>112</sup> Heroin, which activates the OP<sub>3</sub> receptor, attenuates the immune system and is thought to be a cofactor in the pathogenesis of HIV infection.<sup>30,113–115</sup>

Opioids have also been shown to affect the progression of disease in HIV infection. Opioids enhance immunodeficiency and increase the viral load in humans infected with HIV.<sup>114,116–118</sup> Cessation of IVDU in HIV-positive individuals resulted in a lower incidence of acquired immunodeficiency syndrome.<sup>119</sup> In vitro, morphine has been shown to promote production of HIV virus in lymphocytes.<sup>114</sup>

Healthy human volunteers exhibited a dose dependent reduction in NK cell activity 2 hours after receiving morphine at analgesic doses. This reduction in NK cell activity persisted for more than 24 hours.<sup>79</sup> Sacerdote et al<sup>28</sup> investigated the effect of morphine and tramadol in a group of human patients after abdominal tumor resection. Tramadol was shown to enhance NK cell activity while morphine suppressed lymphocyte proliferation in this population of patients. Opioids may also contribute to increased infection rates in hospitalized patients. For example, patients with mildto-moderate burn injuries are more likely to develop infectious complications when treated with opioids.<sup>120</sup> In another study, men that underwent radical prostatectomy surgery were given either systemic opioids or epidural analgesia for pain control. Patients who had an epidural anesthesia were 57% less likely to have tumor reoccurrence when compared with those who received systemic morphine for pain control.<sup>121</sup> The authors speculated that the use of regional anesthesia reduces the neuroendocrine stress response to surgery, blocked the descending efferent activation of the sympathetic nervous system, and reduced the amount of general anesthesia required (all of which contribute to immunosuppression).<sup>121</sup> Regional anesthesia also provides adequate pain relief and may obviate the need for postoperative opioids and the consequent adverse effects on immune function and tumor growth.<sup>121</sup>

Naloxone has been shown to improve mean arterial blood pressure during septic shock in rats and human patients. Authors of a meta-analysis evaluating the use of naloxone in septic shock in humans concluded that naloxone administration may have hemodynamic benefits.<sup>104</sup> In one study, continuous infusion of naloxone decreased mortality but this effect has not been confirmed in subsequent studies.<sup>104–106,122,123</sup> The use of naloxone in septic shock is still controversial.

#### Evidence in veterinary species

The effects of opioids on the immune system have primarily been studied in murine rodents. Opioid receptors are present in many species including dogs, cats, horses, cows, monkeys, and sheep.<sup>124–127</sup> There are few studies evaluating the effects of opioids on immunomodulation in other species. It is possible that other species will demonstrate the same response to opioids seen in murine and human models but more controlled studies need to be performed to establish the effects of opioids in other species.

The immunomodulatory properties of opioids have been evaluated in pigs and cats. Bearing in mind the limited evaluation, morphine appears to have similar immunomodulatory effects in pigs as it does in mice and humans. Administration of high-dose morphine (3.3 mg/kg) to pigs downregulates neutrophil and monocyte activity.<sup>128,129</sup> Morphine also reduces T lymphocyte and NK cell activity in pigs when administered before vaccination with the Bacille-Calmette-Guérin vaccine. This vaccine stimulates T lymphocyte proliferation in humans and mice and has the same effect in the pig.<sup>46,79</sup>

In cats, the impact of morphine administration and withdrawal on lentivirus-related disease progression has been evaluated.<sup>130</sup> Morphine-treated feline immunodeficiency virus (FIV)-positive cats did not experience increase incidence or severity of FIV-related diseases.<sup>130</sup> While this study did not demonstrate an opioid-related increase in susceptibility to FIV-related diseases, the results should be interpreted with caution because of small sample size and large variation in viremia severity.

#### **Clinical implications**

Most opioids used in clinical practice affect the immune system to varying degrees. Many ICU patients are immunocompromised and it is important to select analgesic drugs carefully to avoid promoting further immunosuppression.<sup>27</sup> A healthy patient would likely tolerate immune modulations without clinically important consequences but there are situations (young, elderly, or immunocompromised patients) where the risks associated with opioid-related immunomodulation may be of clinical concern. In critically ill patients or patients with risk factors for immune dysfunction, opioids with minimal or no immunomodulatory properties (buprenorphine, hydromorphone, tramadol, oxycodone, and oxymorphone) may be more advantageous compared with opioids with known immunomodulatory properties (morphine, fentanyl, methadone, and codeine).

While the risks of immunosuppression with the use of opioids should be considered in the critical patient, the importance of adequate analgesia cannot be overstated. Untreated pain produces a generalized sympthathetic response, leading to tachycardia, increased cardiac output and peripheral vasoconstriction that may cause hypertension and contribute to hemodynamic instability in the critical patient.<sup>131</sup> In some cases, pain may cause a generalized vagal response consisting of bradycardia and systemic hypotension, which may produce end-organ damage as a result of poor perfusion.<sup>132</sup> Indeed, untreated pain alone may ultimately lead to immunosuppression.133,134 Unfortunately, the ability of specific opioids to reverse paininduced immunosuppression has not been assessed clinically. Because opioids are the most effective analgesic drugs available for treating pain, their use in the critical patient is still advocated when pain is recognized or suspected.

Multimodal analgesia, using a combination of different classes of analgesics is an important strategy to improve the efficacy of pain management. In addition, this approach has the potential to reduce the amount of opioids required for analgesia and thus may help minimize potential immunosuppressive effects associated with the use of opioids. However, it should be noted that many analgesic medications have been shown to alter immune responses so careful consideration of each medication selection is recommended.<sup>135–138</sup> *N*-methyl-D-aspartate antagonists, sodium channel blockers and nonsteroidal anti-inflammatory drugs have been used with great success in human and veterinary medicine to provide multimodal analgesia.<sup>139,140</sup> Neuraxial analgesia will also significantly reduce the amount of systemic opioids needed and should be utilized as often as possible when managing the critical patient. The use of multimodal analgesia in veterinary species has been reviewed elsewhere.<sup>141–144</sup>

While there is compelling evidence demonstrating the immunomodulatory effects of endogenous and exogenous opioids in animal models and humans, a consensus on the appropriate use of opioids in critical patients has not been reached. In light of the fact that there is little clinical evidence evaluating the impact of opioids on outcome, additional research is needed in the clinical veterinary patient population. In the meantime, patients should be treated for pain, using opioids when clinically appropriate. Opioids that cause minimal immunosuppression should be considered in favor of opioids that cause immunosuppression whenever possible.

#### References

- Dhawan BN, Cesselin F, Raghubir R, et al. International union of pharmacology. Xii. Classification of opioid receptors. Pharmacol Rev 1996; 48(4):567–592.
- Carr DB, Goudas LC. Acute pain. Lancet 1999; 353(9169): 2051–2058.
- Schofferman J. Long-term opioid analgesic therapy for severe refractory lumbar spine pain. Clin J Pain 1999; 15(2):136–140.
- Weinert C, Kethireddy S, Roy S. Opioids and infections in the intensive care unit should clinicians and patients be concerned? J Neuroimmune Pharmacol 2008; 3(4):218–229.
- Stein C, Lang LJ. Peripheral mechanisms of opioid analgesia. Curr Opin Pharmacol 2009; 9(1):3–8.
- Inturrisi CE. Clinical pharmacology of opioids for pain. Clin J Pain 2002; 18(4):S3–S13.
- Lopez A, Salomé L. Membrane functional organisation and dynamic of μ-opioid receptors. Cell Mol Life Sci 2009; 66(13): 2093–2018.
- Waldhoer M, Bartlett SE, Whistler JL. Opioid receptors. Annu Rev Biochem 2004; 73(1):953–990.
- 9. Janson W, Stein C. Peripheral opioid analgesia. Curr Pharm Biotechnol 2003; 4(4):270–274.
- Barry U, Zuo Z. Opioids: old drugs for potential new applications. Curr Pharm Des 2005; 11(10):1343–1350.
- 11. Pasternak GW. Multiple opiate receptors: Déjà vu all over again. Neuropharmacology 2004; 47(suppl 1):312–323.
- Goodman Allan J, Le Bourdonnec B, Dolle Roland E. Mu opioid receptor antagonists: recent developments. ChemMedChem 2007; 2(11):1552–1570.
- McCarthy L, Wetzel M, Sliker JK, et al. Opioids, opioid receptors, and the immune response. Drug Alcohol Depend 2001; 62(2): 111–123.
- Hilburger ME, Adler MW, Truant AL, et al. Morphine induces sepsis in mice. J Infect Dis 1997; 176(1):183–188.
- McQuay H. Opioids in pain management. Lancet 1999; 353(9171):2229–2232.
- Stanway G, Taylor PM, Brodbelt DC. A preliminary investigation comparing pre-operative morphine and buprenorphine for postoperative analgesia and sedation in cats. Vet Anaesth Analg 2002; 29(1):29–35.
- Walsh SL, Preston KL, Bigelow GE, et al. Acute administration of buprenorphine in humans: partial agonist and blockade effects. J Pharmacol Exp Ther 1995; 274(1):361–372.

- 18. Lawlor P, Turner K, Hanson J, et al. Dose ratio between morphine and hydromorphone in patients with cancer pain: a retrospective study. Pain 1997; 72(1–2):79–85.
- Eddy NB, Lee LE Jr. The analgesic equivalence to morphine and relative side action liability of oxymorphone (4-hydroxydihydromorphinone). J Pharmacol Exp Ther 1959; 125(2): 116–121.
- Paix A, Coleman A, Lees J, et al. Subcutaneous fentanyl and sufentanil infusion substitution for morphine intolerance in cancer pain management. Pain 1995; 63(2):263–269.
- Collins RJ, Weeks JR. Relative potency of codeine, methadone and dihydromorphinone to morphine in self-maintained addict rats. Naunyn Schmiedebergs Arch Pharmacol 1965; 249(6): 509–514.
- Pereira J, Lawlor P, Vigano A, et al. Equianalgesic dose ratios for opioids: a critical review and proposals for long-term dosing. J Pain Symptom Manage 2001; 22(2):672–687.
- Roy S, Barke RA, Loh HH. Mu-opioid receptor-knockout mice: role of mu-opioid receptor in morphine mediated immune functions. Brain Res Mol Brain Res 1998; 61(1–2):190–194.
- Eisenstein TK, Hilburger ME. Opioid modulation of immune responses: effects on phagocyte and lymphoid cell populations. J Neuroimmunol 1998; 83(1–2):36–44.
- Bidlack JM. Detection and function of opioid receptors on cells from the immune system. Clin Diagn Lab Immunol 2000; 7(5):719–723.
- 26. Sacerdote P. Opioid-induced immunosuppression. Curr Opin Support Palliat Care 2008; 2(1):14–18.
- Budd K. Pain management: is opioid immunosuppression a clinical problem? Biomed Pharmacother 2006; 60(7):310–317.
- Sacerdote P, Bianchi M, Gaspani L, et al. The effects of tramadol and morphine on immune responses and pain after surgery in cancer patients. Anesth Analg 2000; 90(6):1411–1414.
- Martucci C, Panerai AE, Sacerdote P. Chronic fentanyl or buprenorphine infusion in the mouse: similar analgesic profile but different effects on immune responses. Pain 2004; 110 (1–2):385–392.
- Sacerdote P. Opioids and the immune system. Palliat Med 2006; 20(8 suppl):9–15.
- Gomez-Flores R, Weber RJ. Differential effects of buprenorphine and morphine on immune and neuroendocrine functions following acute administration in the rat mesencephalon periaqueductal gray. Immunopharmacology 2000; 48(2):145–156.
- 32. Wang J, Charboneau R, Balasubramanian S, et al. The immunosuppressive effects of chronic morphine treatment are partially dependent on corticosterone and mediated by the μ-opioid receptor. J Leukoc Biol 2002; 71(5):782–790.
- Paola S, Barbara M, Paolo M, et al. Antinociceptive and immunosuppressive effects of opiate drugs: a structure-related activity study. Br J Pharmacol 1997; 121(4):834–840.
- 34. Franchi S, Panerai AE, Sacerdote P. Buprenorphine ameliorates the effect of surgery on hypothalamus-pituitary-adrenal axis, natural killer cell activity and metastatic colonization in rats in comparison with morphine or fentanyl treatment. Brain Behav Immun 2007; 21(6):767–774.
- Saurer TB, Carrigan KA, Ijames SG, et al. Suppression of natural killer cell activity by morphine is mediated by the nucleus accumbens shell. J Neuroimmunol 2006; 173(12):3–11.
- Beilin B, Shavit Y, Hart J, et al. Effects of anesthesia based on large versus small doses of fentanyl on natural killer cell cytotoxicity in the perioperative period. Anesth Analg 1996; 82(3):492–497.
- 37. Forget P, Collet V, Lavand'homme P, et al. Does analgesia and condition influence immunity after surgery? Effects of fentanyl, ketamine and clonidine on natural killer activity at different ages. Eur J Anaesthesiol 2010; 27(3):233–240.
- Bilfinger TV, Fimiani C, Stefano GB. Morphine's immunoregulatory actions are not shared by fentanyl. Int J Cardiol 1998; 64(suppl 1):S61–S66.
- House RV, Thomas PT, Bhargava HN. In vitro evaluation of fentanyl and meperidine for immunomodulatory activity. Immunol Lett 1995; 46(1–2):117–124.

- 40. Pechnick RN. Effects of opioids on the hypothalamo-pituitaryadrenal axis. Annu Rev Pharmacol Toxicol 1993; 33(1):353–382.
- Martin-Kleiner I, Gabrilovac J. Naloxone modulates nk-cell activity of human peripheral blood lymphocytes like an opioid agonist. Immunopharmacol Immunotoxicol 1993; 15(2–3): 179–197.
- Sacerdote P, Manfredi B, Gaspani L, et al. The opioid antagonist naloxone induces a shift from type 2 to type 1 cytokine pattern in balb/cj mice. Blood 2000; 95(6):2031–2036.
- 43. Wei G, Moss J, Yuan C-S. Opioid-induced immunosuppression: is it centrally mediated or peripherally mediated? J Pharm Biomed Anal 2003; 65(11):1761–1766.
- Mellon RD, Bayer BM. Evidence for central opioid receptors in the immunomodulatory effects of morphine: review of potential mechanism(s) of action. J Neuroimmunol 1998; 83(1–2):19–28.
- Nelson CJ, Schneider GM, Lysle DT. Involvement of central mubut not delta- or kappa-opioid receptors in immunomodulation. Brain Behav Immun 2000; 14(3):170–184.
- Gaveriaux-Ruff C, Matthes HW, Peluso J, et al. Abolition of morphine-immunosuppression in mice lacking the mu-opioid receptor gene. Proc Natl Acad Sci USA 1998; 95(11):6326–6330.
- Hall DM, Suo JL, Weber RJ. Opioid mediated effects on the immune system: sympathetic nervous system involvement. J Neuroimmunol 1998; 83(1–2):29–35.
- Shavit Y, Lewis JW, Terman GW, et al. Opioid peptides mediate the suppressive effect of stress on natural killer cell cytotoxicity. Science 1984; 223(4632):188–190.
- Hernandez M, Flores L, Bayer B. Immunosuppression by morphine is mediated by central pathways. J Pharmacol Exp Ther 1993; 267(3):1336–1341.
- Shavit Y, Depaulis A, Martin FC, et al. Involvement of brain opiate receptors in the immune-suppressive effect of morphine. Proc Natl Acad Sci U S A 1986; 83(18):7114–7117.
- Mellon RD, Bayer BM. Role of central opioid receptor subtypes in morphine-induced alterations in peripheral lymphocyte activity. Brain Res 1998; 789(1):56–67.
- Liang-Suo J, Gomez-Flores R, Weber RJ. Immunosuppression induced by central action of morphine is not blocked by mifepristone (ru 486). Life Sci 2002; 71(22):2595–2602.
- Weber R, Pert A. The periaqueductal gray matter mediates opiate-induced immunosuppression. Science 1989; 245(4914): 188–190.
- Nelson CJ, Schneider GM, Lysle DT. Involvement of central [mu]but not [delta]- or [kappa]-opioid receptors in immunomodulation. Brain Behav Immun 2000; 14(3):170–184.
- Vallejo R, de Leon-Casasola O, Benyamin R. Opioid therapy and immunosuppression: a review. Am J Ther 2004; 11(5):354–365.
- Roy S, Wang J, Kelschenbach J, et al. Modulation of immune function by morphine: implications for susceptibility to infection. J Neuroimmune Pharmacol 2006; 1(1):77–89.
- 57. Alonzo NC, Bayer BM. Opioids, immunology, and host defenses of intravenous drug abusers. Infect Dis Clin North Am 2002; 16(3):553–569.
- Flores LR, Hernandez MC, Bayer BM. Acute immunosuppressive effects of morphine: lack of involvement of pituitary and adrenal factors. J Pharmacol Exp Ther 1994; 268(3):1129–1134.
- Bryant HU, Bernton EW, Kenner JR, et al. Role of adrenal cortical activation in the immunosuppressive effects of chronic morphine treatment. Endocrinology 1991; 128(6):3253–3258.
- Flores LR, Wahl SM, Bayer BM. Mechanisms of morphine-induced immunosuppression: effect of acute morphine administration on lymphocyte trafficking. J Pharmacol Exp Ther 1995; 272(3):1246–1251.
- Freier DO, Fuchs BA. A mechanism of action for morphine-induced immunosuppression: corticosterone mediates morphineinduced suppression of natural killer cell activity. J Pharmacol Exp Ther 1994; 270(3):1127–1133.
- Spengler R, Chensue S, Giacherio D, et al. Endogenous norepinephrine regulates tumor necrosis factor-alpha production from macrophages in vitro. J Immunol 1994; 152(6):3024–3031.
- Carr DJ, Gebhardt BM, Paul D. Alpha adrenergic and mu-2 opioid receptors are involved in morphine- induced suppression

of splenocyte natural killer activity. J Pharmacol Exp Ther 1993; 264(3):1179–1186.

- Flores LR, Dretchen KL, Bayer BM. Potential role of the autonomic nervous system in the immunosuppressive effects of acute morphine administration. Eur J Pharmacol 1996; 318(2–3):437– 446.
- Fecho K, Dykstra LA, Lysle DT. Evidence for beta adrenergic receptor involvement in the immunomodulatory effects of morphine. J Pharmacol Exp Ther 1993; 265(3):1079–1087.
- Radulovic J, Miljevic C, Djergovic D, et al. Opioid receptormediated suppression of humoral immune response in vivo and in vitro: involvement of kappa opioid receptors. J Neuroimmunol 1995; 57(1–2):55–62.
- Rojavin M, Szabo I, Bussiere JL, et al. Morphine treatment in vitro or in vivo decreases phagocytic functions of murine macrophages. Life Sci 1993; 53(12):997–1006.
- Chao CC, Hu S, Shark KB, et al. Activation of mu opioid receptors inhibits microglial cell chemotaxis. J Pharmacol Exp Ther 1997; 281(2):998–1004.
- Bussiere J, Adler M, Rogers T, et al. Cytokine reversal of morphine-induced suppression of the antibody response. J Pharmacol Exp Ther 1993; 264(2):591–597.
- Bhargava HN, Thomas PT, Thorat S, et al. Effects of morphine tolerance and abstinence on cellular immune function. Brain Res 1994; 642(1–2):1–10.
- Roy S, Ramakrishnan S, Loh HH, et al. Chronic morphine treatment selectively suppresses macrophage colony formation in bone marrow. Eur J Pharmacol 1991; 195(3):359–363.
- Szabo I, Rojavin M, Bussiere JL, et al. Suppression of peritoneal macrophage phagocytosis of *Candida albicans* by opioids. J Pharmacol Exp Ther 1993; 267(2):703–706.
- Sharp BM, Keane WF, Suh HJ, et al. Opioid peptides rapidly stimulate superoxide production by human polymorphonuclear leukocytes and macrophages. Endocrinology 1985; 117(2):793– 795.
- Bhat RS, Bhaskaran M, Mongia A, et al. Morphine-induced macrophage apoptosis: oxidative stress and strategies for modulation. J Leukoc Biol 2004; 75(6):1131–1138.
- Fecho K, Maslonek KA, Coussons-Read ME, et al. Macrophagederived nitric oxide is involved in the depressed concanavalin a responsiveness of splenic lymphocytes from rats administered morphine in vivo. J Immunol 1994; 152(12):5845–5852.
- Koff WC, Dunegan MA. Modulation of macrophage-mediated tumoricidal activity by neuropeptides and neurohormones. J Immunol 1985; 135(1):350–354.
- 77. Singhal PC, Sharma P, Kapasi AA, et al. Morphine enhances macrophage apoptosis. J Immunol 1998; 160(4):1886–1893.
- Messmer UK, Reed UK, Brune B. Bcl-2 protects macrophages from nitric oxide-induced apoptosis. J Biol Chem 1996; 271(33):20192–20197.
- Yeager MP, Colacchio TA, Yu CT, et al. Morphine inhibits spontaneous and cytokine-enhanced natural killer cell cytotoxicity in volunteers. Anesthesiology 1995; 83(3):500–508.
- Bayer BM, Daussin S, Hernandez M, et al. Morphine inhibition of lymphocyte activity is mediated by an opioid dependent mechanism. Neuropharmacology 1990; 29(4):369–374.
- Lysle DT, Hoffman KE, Dykstra LA. Evidence for the involvement of the caudal region of the periaqueductal gray in a subset of morphine-induced alterations of immune status. J Pharmacol Exp Ther 1996; 277(3):1533–1540.
- Ishikawa M, Tanno K, Kamo A, et al. Enhancement of tumor growth by morphine and its possible mechanism in mice. Biol Pharm Bull 1993; 16(8):762–766.
- Yeager MP, Colacchio TA. Effect of morphine on growth of metastatic colon cancer in vivo. Arch Surg 1991; 126(4):454–456.
- 84. Sasamura T, Nakamura S, Iida Y, et al. Morphine analgesia suppresses tumor growth and metastasis in a mouse model of cancer pain produced by orthotopic tumor inoculation. Eur J Pharmacol 2002; 441(3):185–191.
- Roy S, Loh H. Effects of opioids on the immune system. Neurochem Res 1996; 21(11):1375–1386.

- Roy S, Balasubramanian S, Sumandeep S, et al. Morphine directs T cells toward T differentiation. Surgery 2001; 130(2):304–309.
- Roy S, Wang J, Gupta S, et al. Chronic morphine treatment differentiates t helper cells to th2 effector cells by modulating transcription factors gata 3 and t-bet. J Neuroimmunol 2004; 147(1–2):78–81.
- Greeneltch KM, Kelly-Welch AE, Shi Y, et al. Chronic morphine treatment promotes specific th2 cytokine production by murine t cells in vitro via a fas/fas ligand-dependent mechanism. J Immunol 2005; 175(8):4999–5005.
- Bryant HU, Bernton EW, Holaday JW. Immunosuppressive effects of chronic morphine treatment in mice. Life Sci 1987; 41(14):1731–1738.
- Roy S, Charboneau R, Barke R, et al. Role of mu-opioid receptor in immune function. Neuroimmune Circuits, Drugs Abuse, Infect Dis 2001; 493:117–126.
- Belkowski SM, Alicea C, Eisenstein TK, et al. Inhibition of interleukin-1 and tumor necrosis factor-alpha synthesis following treatment of macrophages with the kappa opioid agonist u50, 488h. J Pharmacol Exp Ther 1995; 273(3):1491–1496.
- Roy S, Cain KJ, Chapin RB, et al. Morphine modulates nf[kappa]b activation in macrophages. Biochem Biophys Res Commun 1998; 245(2):392–396.
- Peterson P, Sharp B, Gekker G, et al. Opioid-mediated suppression of cultured peripheral blood mononuclear cell respiratory burst activity. J Immunol 1987; 138(11):3907–3912.
- 94. Wang J, Barke RA, Charboneau R, et al. Morphine negatively regulates interferon-gamma promoter activity in activated murine t cells through two distinct cyclic amp-dependent pathways. J Biol Chem 2003; 278(39):37622–37631.
- Peng X, Mosser DM, Adler MW, et al. Morphine enhances interleukin-12 and the production of other pro-inflammatory cytokines in mouse peritoneal macrophages. J Leukoc Biol 2000; 68(5):723–728.
- Bhaskaran M, Reddy K, Sharma S, et al. Morphine-induced degradation of the host defense barrier: role of macrophage injury. J Infect Dis 2001; 184(12):1524–1531.
- Lucas AN, Firth AM, Anderson GA, et al. Comparison of the effects of morphine administered by constant-rate intravenous infusion or intermittent intramuscular injection in dogs. J Am Vet Med Assoc 2001; 218(6):884–891.
- Tseng CS, Tso HS. Effects of opioid agonists and opioid antagonists in endotoxic shock in rats. Anaesthesiologica Sinica 1993; 31(1):1–8.
- Ocasio FM, Jiang Y, House SD, et al. Chronic morphine accelerates the progression of lipopolysaccharide-induced sepsis to septic shock. J Neuroimmunol 2004; 149(1–2):90–100.
- 100. Tsai S-M, Lin M-T, Wang J-J, et al. Pyrogens enhance beta endorphin release in hypothalamus and trigger fever that can be attenuated by buprenorphine. J Pharmacol Sci 2003; 93(2):155– 162.
- 101. Donaldson MD, Vesey CJ, Wilks M, et al. Beneficial effects of buprenorphine (a partial opiate agonist) in porcine *Escherichia coli* septicaemia: a comparison with naloxone. Circ Shock 1988; 25(3):209–221.
- Lin S-L, Lee Y-M, Chang H-Y, et al. Effects of naltrexone on lipopolysaccharide-induced sepsis in rats. J Biomed Sci 2005; 12(2):431–440.
- 103. Greeneltch KM, Haudenschild CC, Keegan AD, et al. The opioid antagonist naltrexone blocks acute endotoxic shock by inhibiting tumor necrosis factor-[alpha] production. Brain Behav Immun 2004; 18(5):476–484.
- 104. Boeuf B, Gauvin F, Guerguerian AM, et al. Therapy of shock with naloxone: a meta-analysis. Crit Care Med 1998; 26(11):1910–1916.
- Peters WP, Johnson MW, Friedman PA, et al. Pressor effect of naloxone in septic shock. The Lancet 1981; 1(8219):529–532.
- Rock P, Silverman H, Plump D, et al. Efficacy and safety of naloxone in septic shock. Crit Care Med 1985; 13(1):28–33.
- 107. Hussey HH, Katz S. Infections resulting from narcotic addiction; report of 102 cases. Am J Med 1950; 9(2):186–193.

- Friedman H, Newton C, Klein TW. Microbial infections, immunomodulation, and drugs of abuse. Clin Microbiol Rev 2003; 16(2):209–219.
- Reichman LB, Felton CP, Edsall JR. Drug dependence, a possible new risk factor for tuberculosis disease. Arch Intern Med 1979; 139(3):337–339.
- Des Jarlais DC. Potential cofactors in the outcomes of HIV infection in intravenous drug users. NIDA Res Monogr 1991; 109:115–123.
- 111. De Paoli P, Reitano M, Battistin S, et al. Immunological abnormalities in intravenous drug abusers and relationship to the prolonged generalized lymphadenopathy syndrome in Italy. Clin Exp Immunol 1986; 64(3):451–456.
- 112. Cooper O, Brown T, Dobs A. Opiate drug use: a potential contributor to the endocrine and metabolic complications in human immunodeficiency virus disease. Clin Infect Dis 2003; 37(s2): S132–S136.
- Nair MP, Schwartz SA, Polasani R, et al. Immunoregulatory effects of morphine on human lymphocytes. Clin Diagn Lab Immunol 1997; 4(2):127–132.
- Peterson PK, Sharp BM, Gekker G, et al. Morphine promotes the growth of HIV-1 in human peripheral blood mononuclear cell cocultures. AIDS 1990; 4(9):869–873.
- 115. Kitanaka N, Sora I, Kinsey S, et al. No heroin or morphine 6β-glucuronide analgesia in μ-opioid receptor knockout mice. Eur J Pharmacol 1998; 355(1):R1–R3.
- Lokensgard JR, Gekker G, Peterson PK. κ-opioid receptor agonist inhibition of HIV-1 envelope glycoprotein-mediated membrane fusion and CXCR4 expression on CD4+ lymphocytes. Biochem Pharmacol 2002; 63(6):1037–1041.
- 117. Hubbard RL, Marsden ME, Cavanaugh E, et al. Role of drugabuse treatment in limiting the spread of aids. Rev Infect Dis 1988; 10(2):377–384.
- Peterson PK, Gekker G, Lokensgard JR, et al. κ-opioid receptor agonist suppression of HIV-1 expression in CD4+lymphocytes. J Pharm Biomed Anal 2001; 61(9):1145–1151.
- 119. Weber R, Ledergerber B, Opravil M, et al. Progression of HIV infection in misusers of injected drugs who stop injecting or follow a programme of maintenance treatment with methadone. BMJ 1990; 301(6765):1362–1365.
- Schwacha MG, McGwin JG, Hutchinson CB, et al. The contribution of opiate analgesics to the development of infectious complications in burn patients. Am J Surg 2006; 192(1):82–86.
- 121. Biki B, Mascha E, Moriarty DC, et al. Anesthetic technique for radical prostatectomy surgery affects cancer recurrence: a retrospective analysis. Anesthesiology 2008; 109(2):180–187.
- Roberts D, Hall K, Dobson K, et al. Effects of prolonged naloxone infusion in septic shock. The Lancet 1988; 332(8613):699–702.
- Bonnet F, Bilaine J, Lhoste F, et al. Naloxone therapy of human septic shock. Crit Care Med 1985; 13(11):972–975.
- 124. Hellyer PW, Bai L, Supon J, et al. Comparison of opioid and alpha-2 adrenergic receptor binding in horse and dog brain using radioligand autoradiography. Vet Anaesth Analg 2003; 30(3):172– 182.
- 125. Clark WG, Ponder SW. Thermoregulatory effects of (d-ala2)-methionine-enkephalinamide in the cat. Evidence for multiple naloxone-sensitive opioid receptors. Brain Res Bull 1980; 5(4):415–420.
- 126. Castanas E, Bourhim N, Giraud P, et al. Interaction of opiates with opioid binding sites in the bovine adrenal medulla: Ii. Interaction with kappa sites. J Neurochem 1985; 45(3):688–699.
- 127. Bouaziz H, Tong C, Yoon Y, et al. Intravenous opioids stimulate norepinephrine and acetylcholine release in spinal cord dorsal

horn: systematic studies in sheep and an observation in a human. Anesthesiology 1996; 84(1):143–154.

- 128. Clark JD, Shi X, Li X, et al. Morphine reduces local cytokine expression and neutrophil infiltration after incision. Mol Pain 2007; 3(1):28.
- Bilfinger TV, Kushnerik V, Bundz S, et al. Evidence for morphine downregulating immunocytes during cardiopulmonary bypass in a porcine model. Int J Cardiol 1996; 53(suppl 1): S39–S46.
- Barr MC, Huitron-Resendiz S, Sanchez-Alavez M, et al. Escalating morphine exposures followed by withdrawal in feline immunodeficiency virus-infected cats: a model for HIV infection in chronic opiate abusers. Drug Alcohol Depend 2003; 72(2):141– 149.
- 131. McLachlan E. Recognizing pain. Am J Nurs 1974; 74(3):496-497.
- O'Gara PT. The hemodynamic consequences of pain and its management. J Intensive Care Med 1988; 3(1):3–5.
- Stucky CL, Gold MS, Zhang X. Mechanisms of pain. Natl Acad Sci USA 2001; 98(21):11845–11846.
- 134. Pergolizzi J, Böger RH, Budd K, et al. Opioids and the management of chronic severe pain in the elderly: consensus statement of an international expert panel with focus on the six clinically most often used World Health Organization step III opioids (buprenorphine, fentanyl, hydromorphone, methadone, morphine, oxycodone). Pain Pract 2008; 8(4):287–313.
- 135. DeClue AE, Cohn LA, Lechner ES, et al. Effects of subanesthetic doses of ketamine on hemodynamic and immunologic variables in dogs with experimentally induced endotoxemia. Am J Vet Res 2008; 69(2):228–232.
- 136. Furst SM, Komocsar WJ, Khan KN, et al. Screening new drugs for immunotoxic potential: I. Assessment of the effects of conventional nonsteroidal anti-inflammatory drugs and selective cox-2 inhibitors on in vitro and in vivo phagocytic activity. J Immunotoxicol 2005; 1(3):149–158.
- Toth PD, Hamburger SA, Hastings GH, et al. Benoxaprofen attenuation of lethal canine endotoxic shock. Circ Shock 1985; 15(2):89–103.
- 138. Son KA, Kang JH, Yang MP. Ketamine inhibits the phagocytic responses of canine peripheral blood polymorphonuclear cells through the upregulation of prostaglandin E<sub>2</sub> in peripheral blood mononuclear cells in vitro. Res Vet Sci 2009; 87(1): 41–46.
- 139. Schmid RL, Sandler AN, Katz J. Use and efficacy of low-dose ketamine in the management of acute postoperative pain: a review of current techniques and outcomes. Pain 1999; 82(2):111– 125.
- Smith LJ, Shih A, Miletic G, et al. Continual systemic infusion of lidocaine provides analgesia in an animal model of neuropathic pain. Pain 2002; 97(3):267–273.
- 141. Mathews KA. Non-steroidal anti-inflammatory analgesics: a review of current practice. J Vet Emerg Crit Care 2002; 12(2): 89–97.
- 142. Mathews KA. Pain assessment and general approach to management. Vet Clin North Am Small Anim Pract 2000; 30(4):729– 755, v.
- Corletto F. Multimodal and balanced analgesia. Vet Res Commun 2007; 31:59–63.
- 144. Jones RS. Epidural analgesia in the dog and cat. Vet J 2001; 161(2):123–131.
- Dhawan BN, Cesselin F, Raghubir R, et al. International union of pharmacology. XII. Classification of opioid receptors. Pharmacol Rev 1996; 48(4):567–592.

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